

**ANALISIS METABOLIT SEKUNDER DARI AKAR DAN DAUN  
HANJELI (*Coix lacryma-jobi L.*) KETAN DAN HANJELI PUTIH  
MENGGUNAKAN GC-MS**

**SKRIPSI**

diajukan sebagai salah satu syarat untuk memperoleh gelar Sarjana Sains Program Studi  
Biologi Departemen Pendidikan Biologi



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Sarjana Sains pada Departemen Pendidikan Biologi Fakultas Pendidikan  
Matematika dan Ilmu Pengetahuan Alam

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MENGGUNAKAN GC-MS

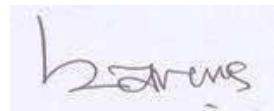
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## **PERNYATAAN**

Dengan ini saya menyatakan bahwa skripsi dengan judul “**Analisis Metabolit Sekunder dari Akar dan Daun Hanjeli (*Coix lacryma-Jobi L.*) Ketan dan Hanjeli Putih Menggunakan GC-MS**” ini beserta seluruh isinya merupakan karya saya sendiri. Saya tidak melakukan penjiplakan atau pengutipan dengan cara-cara yang tidak sesuai dengan etika ilmu yang berlaku dalam masyarakat keilmuan. Atas pernyataan ini, saya siap menanggung risiko/sanksi apabila di kemudian hari ditemukan adanya pelanggaran etika keilmuan atau ada klaim dari pihak lain terhadap keaslian karya saya ini.

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## **ANALISIS METABOLIT SEKUNDER DARI AKAR DAN DAUN HANJELI (*Coix lacryma-jobi L.*) KETAN DAN HANJELI PUTIH MENGGUNAKAN GC-MS**

### **ABSTRAK**

Hanjeli (*Coix lacryma-jobi L.*) digunakan sebagai obat tradisional di berbagai negara dan telah diketahui mengandung metabolit sekunder yang berbeda di setiap daerah. Di Indonesia, pemanfaatan hanjeli masih terbatas pada organ biji dalam jumlah yang sangat kecil, sedangkan organ akar dan daun seringkali tidak dimanfaatkan lebih lanjut. Penelitian ini bertujuan untuk menganalisis kandungan metabolit sekunder pada akar dan daun tanaman hanjeli ketan dan hanjeli putih sehingga diharapkan mampu meningkatkan pemanfaatan tanaman hanjeli di Indonesia. Sampel tanaman diambil dari Desa Cikadut, Kecamatan Cimenyan, Kabupaten Bandung, Jawa Barat. Sampel diekstrak menggunakan metode maserasi dengan pelarut etanol 96%. Analisis kandungan metabolit sekunder dilakukan menggunakan metode GC-MS. Data hasil GC-MS diidentifikasi dengan melihat indeks kesamaannya dengan data di pustaka *National Institute of Standards and Technology* (NIST). Hasil penelitian ini menunjukkan bahwa akar hanjeli ketan mengandung 19 senyawa, sedangkan akar hanjeli putih mengandung 18 senyawa. Kedua akar hanjeli mengandung coixol dengan konsentrasi tinggi dan banyak mengandung senyawa fenolik. Daun hanjeli ketan mengandung 9 senyawa dengan senyawa dominan 5-hidroksimetilfurfural, sedangkan daun hanjeli putih mengandung 15 senyawa dengan senyawa dominan asam palmitat. Terdapat empat senyawa yang sama dengan konsentrasi yang berbeda, yaitu coixol, asam palmitat, 2-metoksi-4-vinilfenol, dan asam ferulat. Akar dan daun hanjeli ketan dan hanjeli putih memiliki kandungan metabolit sekunder yang berbeda.

Kata kunci: akar, daun, GC-MS, hanjeli, metabolit sekunder

**ANALYSIS OF SECONDARY METABOLITE FROM ROOTS AND LEAVES  
OF HANJELI (*Coix lacryma-jobi L.*) KETAN AND HANJELI PUTIH  
USING GC-MS**

**ABSTRACT**

*Hanjeli (Coix lacryma-jobi L.) is used as traditional medicine in various countries and has been known to contain different secondary metabolites in each location. In Indonesia, the use of hanjeli is still limited to seeds in very small quantities, while roots and leaves are often not utilized further. This study aims to analyze the content of secondary metabolites in the roots and leaves of hanjeli ketan and hanjeli putih to increase the utilization of hanjeli in Indonesia. Plant samples were taken from Cikadut Village, Cimenyan District, Bandung Regency, West Java. Samples were extracted using the maceration method with 96% ethanol as solvent. The secondary metabolite content analysis was carried out using the GC-MS method. The GC-MS data were identified by looking at their similarity index with the National Institute of Standards and Technology (NIST) library data. The results of this study showed that the root of hanjeli ketan contained 19 compounds, while the root of hanjeli putih contained 18 compounds. Both hanjeli roots contain high concentrations of coixol and contain many phenolic compounds. Hanjeli ketan leaves contain 9 compounds with the dominant compound 5 hydroxymethylfurfural, while hanjeli putih leaves contain 15 compounds with the dominant compound palmitic acid. There are four of the same compounds with different concentrations, namely coixol, palmitic acid, 2-methoxy-4-vinylphenol, and ferulic acid. The roots and leaves of hanjeli ketan and hanjeli putih contain different secondary metabolites.*

*Keywords:* roots, leaves, GC-MS, hanjeli, secondary metabolites

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## DAFTAR PUSTAKA

- Abdulmalik, O., Safo, M. K., Chen, Q. K., Yang, J. S., Brugnara, C., Ohene-Frempong, K., Abraham, D. J., & Asakura, T. (2005). 5-Hydroxymethyl-2-furfural modifies intracellular sickle haemoglobin and inhibits sickling of red blood cells. *Br. J. Haematol.* 128, 552– 561.
- Aftab, T. & Hakeem, K. R. (2021). Medicinal and Aromatic Plants, Healthcare and Industrial Applications. Switzerland: Springer Nature. doi: <https://doi.org/10.1007/978-2-020-58975-2>.
- Aid, F. (2020). Plant Lipid Metabolism. *Advances in Lipid Metabolism*. doi:[10.5772/intechopen.81355](https://doi.org/10.5772/intechopen.81355)
- Ahmed, E., Arshad, M., Khan, M. Z., Amzad, M. S., Sadaf, H. M., Riaz, I., Sabir, S., Ahmad, N., & Sabaoon. (2017). Secondary Metabolites and Their Multidimensional Prospective in Plant Life. *Journal of Pharmacognosy and Phytochemistry*, 6(2). 205-214
- Almeida, D. T., Costa, M. M., & Feitosa, S. (2014). General Aspects Of Palmitic Acid. Palmitic Acid: Occurrence, Biochemistry and Health Effects. Nova Science Publishers, 64-93
- Altieri, C., Cardillo, D., Bevilacqua, A., & Sinigaglia, M. (2007). Inhibition of *Aspergillus spp.* and *Penicillium spp.* by Fatty Acids and Their Monoglycerides. *Journal of Food Protection*, 70(5), 1206– 1212. doi:[10.4315/0362-028X-70.5.1206](https://doi.org/10.4315/0362-028X-70.5.1206)
- Al-Snafi, A. E. (2021). Medicinal Plants Alkaloids, As a Promising Therapeutics- A Review (Part 1). *IOSR Journal of Pharmacy*, 11(2) 2021, 51-67.
- Amen, Y., Arung, E. T., Afifi, M. S., Halim, A. F., Ashour, A., Fujimoto, R., Goto, T., & Shimizu, K. (2017). Melanogenesis inhibitors from *Coix lacryma-jobi* seeds in B16-F10 melanoma cells. *Natural Product Research*, 31(23), 2712– 2718. doi: <https://doi.org/10.1080/14786419.2017.1292270>
- Anulika, N. P., Ignatius, E. S., Raymond, E. S., Osasere, O. I., & Abiola ,A., H. (2016). The Chemistry Of Natural Product: Plant Secondary Metabolites. *International Journal Of Technology Enhancements And Emerging Engineering Research*, 4(8), 1-8.
- Arif, Y., Lombarkia, N., & Souici, F. (2018). Analysis of the volatiles compounds of three date palm, (*Phoenix dactylifera L.*) fruits varieties via SPME-GCMS at two maturation stages and their effect on *Ectomyelois ceratoniae*

- (Lepidoptera: Pyralidae) oviposition behavior. *Journal of Entomological Research*, 42(2), 151-155. doi: [10.5958/0974-4576.2018.00025](https://doi.org/10.5958/0974-4576.2018.00025)
- Arora, R. K. (1977). Job's-tears (*Coix lacryma-jobi*)—a minor food and fodder crop of northeastern India. *Economic Botany*, 31(3), 358–366. doi:[10.1007/bf02866887](https://doi.org/10.1007/bf02866887)
- Asgari Lajayer, B., Ghorbanpour, M., & Nikabadi, S. (2017). Heavy metals in contaminated environment: Destiny of secondary metabolite biosynthesis, oxidative status and phytoextraction in medicinal plants. *Ecotoxicology and Environmental Safety*, 145, 377–390. doi:[10.1016/j.ecoenv.2017.07.035](https://doi.org/10.1016/j.ecoenv.2017.07.035)
- Awuchi, Godswill, C., & Kate, E. C. (2019). Current Developments in Sugar Alcohols: Chemistry, Nutrition, and Health Concerns Of Sorbitol, Xylitol, Glycerol, Arabitol, Inositol, Maltitol, and Lactitol. *International Journal of Advanced Academic Research*, 5(11), 1-33.
- Aziz, T., Febrizky, S., & Mario, A. D. (2014). Pengaruh Jenis Pelarut Terhadap Persen Yield alkaloid dari Daun Salam India (*Murraya Koenigii*). *Teknik Kimia*, 2(20). 1-6
- Balitkabi (Balai Penelitian Tanaman Aneka Kacang dan Umbi). (2017). GCMS Terpasang, Penelitian harus Lebih Maju. [Online]. Diakses dari [http://balitkabi.litbang.pertanian.go.id/berita/gcms-terpasang-penelitian-harus-lebih-maju/#:~:text=GCMS%20\(Gas%20Chromatografi%20Mass%20Spectrometry,yang%20berbeda%20dalam%20analisis%20sampel](http://balitkabi.litbang.pertanian.go.id/berita/gcms-terpasang-penelitian-harus-lebih-maju/#:~:text=GCMS%20(Gas%20Chromatografi%20Mass%20Spectrometry,yang%20berbeda%20dalam%20analisis%20sampel)
- Ban, J. O., Hwang, I. G., Kim, T. M., Hwang, B. Y., Lee, U. S., Jeong, H.-S., ... & Hong, J. T. (2007). Anti-proliferate and pro-apoptotic effects of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyranone through inactivation of NF-κB in Human Colon Cancer Cells. *Archives of Pharmacal Research*, 30(11), 1455–1463. doi:[10.1007/bf02977371](https://doi.org/10.1007/bf02977371)
- Bennour, N., Mighri, H., Eljani, H., Zammouri, T., & Akroud. (2020). Effect of Solvent Evaporation Method on Phenolic Compounds and The Antioxidant Activity of *Moringa oleifera* cultivated in Southern Tunisia. *South African Journal of Botany*, 129, 181-190.
- Bhattarai, H., Saikawa, E., Wan, X., Zhu, H., Ram, K., Gao, S., ..., & Cong, Z. (2019). Levoglucosan as a tracer of biomass burning: Recent progress and perspectives. *Atmospheric Research*. doi:[10.1016/j.atmosres.2019.01.004](https://doi.org/10.1016/j.atmosres.2019.01.004)

- Böttger, A., Vothknecht, U., Bolle, C., & Wolf, A. (2018). Plant Secondary Metabolites and Their General Function in Plants. *Learning Materials in Biosciences*, 3–17. doi:[10.1007/978-3-319-99546-5\\_1](https://doi.org/10.1007/978-3-319-99546-5_1)
- Bourgaud, F., Gravot, A., Milesi, S., & Gontier, E. (2001). Production of plant secondary metabolites: a historical perspective. *Plant Science*, 161(5), 839–851. doi:[10.1016/s0168-9452\(01\)00490-3](https://doi.org/10.1016/s0168-9452(01)00490-3)
- Breitmaier, E. (2006). *Terpenes: Flavors, Fragrances, Pharmaca, Pheromones* Germany : Wiley-VCH
- Burdock, G. A. & Carabin, I. G. (2007). Safety assessment of myristic acid as a food ingredient. *Food and Chemical Toxicology*, 45(4), 517–529. doi:[10.1016/j.fct.2006.10.009](https://doi.org/10.1016/j.fct.2006.10.009)
- Burow, M., Wittstock, U., & Gershenson, J. (2008). Sulfur-Containing Secondary Metabolites and Their Role in Plant Defense. *Advances in Photosynthesis and Respiration*, Springer. 27, 201–222. doi:[10.1007/978-1-4020-6863-8\\_11](https://doi.org/10.1007/978-1-4020-6863-8_11)
- Cai, B., Jack, A. M., Lewis, R. S., Dewey, R. E., & Bush, L. P. (2013). (R)-nicotine biosynthesis, metabolism and translocation in tobacco as determined by nicotine demethylase mutants. *Phytochemistry*, 95, 188–196
- Cheemanapalli, S., Mopuri, R., Golla, R., C.M., Anuradha, Kumar, S., & Chitta. (2018). Syringic acid (SA) – A Review of Its Occurrence, Biosynthesis, Pharmacological and Industrial Importance. *Biomedicine & Pharmacotherapy*, 108, 547-557. doi: <https://doi.org/10.1016/j.biopha.2018.09.069>
- Chen, H.-H., Chiang, W., Chang, J.-Y., Chien, Y.-L., Lee, C.-K., Liu, K.-J., ...& Kuo, C.-C. (2011). Antimutagenic Constituents of Adlay (*Coix lacryma-jobi* L. var.*ma-yuen* Stapf) with Potential Cancer Chemopreventive Activity. *Journal of Agricultural and Food Chemistry*, 59(12), 6444–6452. doi:[10.1021/jf200539r](https://doi.org/10.1021/jf200539r)
- Cho, J.-Y., Moon, J.-H., Seong, K.-Y., & Park, K.-H. (1998). Antimicrobial Activity of 4-Hydroxybenzoic Acid and trans-4-Hydroxycinnamic Acid Isolated and Identified from Rice Hull. *Bioscience, Biotechnology, and Biochemistry*, 62(11), 2273–2276. doi:[10.1271/bbb.62.2273](https://doi.org/10.1271/bbb.62.2273)
- Christoph, R., Schmidt, B., Steinberner, U., Dilla, W., & Karinen, R. (2006). Glycerol. *Ullmann's Encyclopedia of Industrial Chemistry*. doi:[10.1002/14356007.a12\\_477.pub2](https://doi.org/10.1002/14356007.a12_477.pub2)

- Chrzanowska, A., Roszkowski, P., Bielenica, A., Olejarz, W., Stępień, K., & Struga, M. (2019). Anticancer and antimicrobial effects of novel ciprofloxacin fatty acids conjugates. *European Journal of Medicinal Chemistry*, 111810. doi:[10.1016/j.ejmech.2019.111810](https://doi.org/10.1016/j.ejmech.2019.111810)
- Corke, H., Huang, Y., & Li, Y. (2016). The Cereal Grains: *Coix* overview. *Encyclopedia of Food Grains, Second Edition*.
- Costa, B.P., Nassr, M.T., Diz, F.M., Antunes Fernandes, K.H., Antunes, G.L., Grun, L.K., Barbé-Tuana, F.M., Nunes, F.B., Branchini, G., & Rodrigues de Oliveira, J. (2020). Methoxyeugenol regulates the p53/p21 pathway and suppresses human endometrial cancer cell proliferation. *Journal of Ethnopharmacology*. doi: <https://doi.org/10.1016/j.jep.2020.113645>.
- Cronquist, A.J. (1981). *An Integrated System of Classification of Flowering Plants*. Colombia: Colombia University Press.
- Crozier, A., Jaganath, I. B., & Clifford, M. N. (2006). *Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet*. Oxford: Blackwell Publishing Ltd
- Daayf, F. & Lattanzio, V. (2008). *Recent Advences in Polyphenol Research*. United Kingdom: Blackwell Publishing.
- Daderot. (2009). *Coix lacryma-jobi - Oslo botanical garden*. [Online]. Diakses dari [https://commons.wikimedia.org/wiki/File:Coix\\_lacryma-jobi\\_-\\_Oslo\\_botanical\\_garden\\_-\\_IMG\\_8913.jpg](https://commons.wikimedia.org/wiki/File:Coix_lacryma-jobi_-_Oslo_botanical_garden_-_IMG_8913.jpg)
- Das, S., Akhter, R., Khandaker, S., Huque, S., Das, P., Anwar, M., Tanni, K., Shabnaz, S., & Shahriar, M. (2017). Phytochemical screening, antibacterial and anthelmintic activities of leaf and seed extracts of *Coix lacryma-jobi* L. *Journal of Coastal Life Medicine*, 5(8): 360-364
- Daulay, A. S., Ridwanto, Syahputra, R. A., & Nafitri, A. (2021). Antioxidant Activity Test of Chayote (*Sechium edule* (Jacq.) Swartz) Ethanol Extract using DPPH Method. *Journal of Physics: Conference Series*. 1-8. doi:[10.1088/1742-6596/1819/1/012035](https://doi.org/10.1088/1742-6596/1819/1/012035)
- David, G. W. (2005). *Analisis Farmasi, Edisi kedua*. Jakarta: EGC

- Delgoda, R. & Murray, J. E. (2017). Evolutionary Perspectives on the Role of Plant Secondary Metabolites. *Pharmacognosy*, 93–100. doi:[10.1016/b978-0-12-802104-0.00007-x](https://doi.org/10.1016/b978-0-12-802104-0.00007-x)
- Dev, A., Tapas, S., Pratap, S., & Kumar, P. (2012). Structure and function of enzymes of shikimate pathway. *Curr Bioinform*, 7, 374–391
- Devaraj, R. D., Jeppipalli, S. P. K., & Xu, B. (2020). Phytochemistry and health promoting effects of Job's tears (*Coix lacryma-jobi*) - A critical review. *Food Bioscience*, 100537. doi:[10.1016/j.fbio.2020.100537](https://doi.org/10.1016/j.fbio.2020.100537)
- Dewick, P. M. (2009). *Medicinal Natural Products: A Biosynthetic Approach, 3rd Edition*. West Sussex, UK: John Wiley & Sons, Ltd .
- Dick, R., Rattei, T., Haslbeck, M., Schwab, W., Gierl, A., & Frey, M. (2012). Comparative Analysis of Benzoxazinoid Biosynthesis in Monocots and Dicots: Independent Recruitment of Stabilization and Activation Functions. *The Plant Cell*, 24(3), 915–928. doi:[10.1105/tpc.112.096461](https://doi.org/10.1105/tpc.112.096461)
- Diningrat, S. D., Risfandi, M., Harahap, N. S., Sari, A. N., Kusdianti, & Siregar, H. K. (2020). Phytochemical Screening and Antibacterial Activity *Coix lacryma-jobi* Oil. *J Plant Biotechnol*, 47: 100-106. doi: <https://doi.org/10.5010/JPB.2020.47.1.100>
- Diningrat, S. D., Sipayung, S. A., Restuati, M., Marwani, E., Sari, A. N., & Kusdianti. (2018). Isolasi Senyawa Bioinsektisida Pada Ekstrak Etanol Tumbuhan Buasbuas (*Premna Pubescens Blume*) Dengan Metode GCMS. *Prosiding Seminar Nasional Biotik*, 5(1).
- Dirar, A. I., Alsaadi, D. H. M., Wada, M., Mohamed, M. A., Watanabe, T., & Devkota, H. P. (2018). Effects of extraction solvents on total phenolic and flavonoid contents and biological activities of extracts from Sudanese medicinal plants. *South African Journal of Botany*. doi:[10.1016/j.sajb.2018.07.003](https://doi.org/10.1016/j.sajb.2018.07.003)
- El-Demerdash, E. (2011). Anti-inflammatory and antifibrotic effects of methyl palmitate. *Toxicology and Applied Pharmacology*, 254(3), 238–244. doi:[10.1016/j.taap.2011.04.016](https://doi.org/10.1016/j.taap.2011.04.016)
- Encyclopedia Britannica. (2016). *Caryopsis*. [Online]. Diakses dari <https://www.britannica.com/science/caryopsis>

- European Medicine Agency (EMA). (2015). 5-Hydroxymethylfurfural. [Online]. Diakses dari EU/3/15/1441 | European Medicines Agency ([europa.eu](http://europa.eu))
- Figueiredo, A. C., Barroso, J. G., Pedro, L. G., & Scheffer, J. J. C. (2008). Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour and Fragrance Journal*, 23(4), 213–226. doi:[10.1002/ffj.1875](https://doi.org/10.1002/ffj.1875)
- Gao, J., Yu, H., Guo, W., Kong, Y., Gu, lina, Li, Q., Yang, S., Zang, Y., & Wang, Y. (2018). The anticancer effects of ferulic acid is associated with induction of cell cycle arrest and autophagy in cervical cancer cells. *Cancer Cell International*, 18(1). doi:[10.1186/s12935-018-0595-y](https://doi.org/10.1186/s12935-018-0595-y)
- Gao, L., Shen, G., Zhang, L., Qi, J., Zhang, C., Ma, C., Li, J., Wang, L., Malook, S. U., & Wu, J. (2019). An efficient system composed of maize protoplast transfection and HPLC-MS for studying the biosynthesis and regulation of maize benzoxazinoids. *Plant Methods*, 15(144), 1-13. doi: <https://doi.org/10.1186/s13007-019-0529-2>
- Ge, Y., Lu, H., Zhang, J., Wang, C., & Gao, X. (2020). Phytoliths in Inflorescence Bracts: Preliminary Results of an Investigation on Common Panicoideae Plants in China. *Front. Plant Sci.* 10. doi: [10.3389/fpls.2019.01736](https://doi.org/10.3389/fpls.2019.01736)
- George, L. O., Radha, H. R., & Somasekariah, B. V. (2018). In vitro antidiabetic activity and GC-MS analysis of bioactive compounds present in the methanol extract of *Kalanchoe pinnata*. *Indian Journal of Chemistry*, 57B, 1213-1221.
- Goyal, S., Lambert, C., Cluzet, S., Mérillon, J. M., & Ramawat, K. G. (2011). Secondary Metabolites and Plant Defence. *Plant Defence: Biological Control*, 109–138. doi:[10.1007/978-94-007-1933-0\\_5](https://doi.org/10.1007/978-94-007-1933-0_5)
- Graf, E. (1992). Antioxidant potential of ferulic acid. *Free Radical Biology and Medicine*, 13(4), 435–448. doi:[10.1016/0891-5849\(92\)90184-i](https://doi.org/10.1016/0891-5849(92)90184-i)
- Guven, M., Aras, A. B., Topaloglu, N., Ozkan, A., Sen, H. M., Kalkan, Y., Okuyucu, A., Akbal, A., Gokmen, F., & Cosar, M. (2015). The protective effect of syringic acid on ischemia injury in rat brain. *Turkish Journal of Medical Sciences*, 45, 233-240. doi: [10.3906/sag-1402-71](https://doi.org/10.3906/sag-1402-71)
- Hakkim, F. L., Arivazhagan, G., & Boopathy, R. B. (2008). Antioxidant property of selected *Ocimum* species and their secondary metabolite content. *Journal of Medicinal Plants Research*, 2(9), 250-257.
- Haleshappa, R., Patil, S. J., Usha, T., & Murthy, K. S. (2020). Phytochemicals, Antioxidant Profile and GC-MS Analysis of Ethanol Extract of *Simarouba glauca* Seeds. *Asian Journal of Biological and Life Sciences*, 9(3).

- Ham, J. R., Lee, H.I., Choi, R. Y., Sim, M.O., Seo, K.I., & Lee, M. K. (2016). Document details - Anti-steatotic and anti-inflammatory roles of syringic acid in high-fat diet-induced obese mice. *Food and Function*, 7(2), 689-697. doi: [10.1039/c5fo01329a](https://doi.org/10.1039/c5fo01329a)
- Handayani, F., Sumarmiyati, & Rahayu, S. (2019). Karakterisasi Morfologi Jelai (*Coix lacryma-jobi*) Lokal Kalimantan Timur. *PROS SEM NAS MASY BIODIV INDON*, 5(2): 228-233.
- Hartmann, T. (1996). Diversity and variability of plant secondary metabolism: a mechanistic view. *Proceedings of the 9th International Symposium on Insect-Plant Relationships*, 177–188. doi:[10.1007/978-94-009-1720-0\\_42](https://doi.org/10.1007/978-94-009-1720-0_42)
- Hassanpour, S., Maheris-sis, N., Eshratkah, B., & Mehmandar, F. B. (2011). Plants and secondary metabolites (Tannins): A Review. *International Journal of Forest, Soil and Erosion*, 1(1). 47-53.
- Hayashi, H., Takegawa, M., Tzuzuki, M., Matsuzawa, K., Yoshizawa, M., Barla, F., Yukino, T., Mieda, T., Inui, H., Nakano, Y., & Enomoto, T. (2009). Effects of Administration of Adlay Leaves on 2,4,6-trinitro-1-chlorobenzene-induced Chronic Dermatitis in Mice. *Food Sci. Technol. Res*, 15(5).
- Hayashi, T., Gotoh, K., Ohnishi, K., Okamura, K., & Asamizu, T. (1994). 6-METHOXY-2-Benzoxazolinone In *Scoparia Dulcis* and Its Production By Cultured Tissues. *Phytochemistry*, 37(6), 1611-1614.
- Hirooka, K., Miyamoto, O., Jinming, P., Du, Y., Itano, T., Baba, T., Tokuda, M., & Shiraga, F. (2006). Neuroprotective Effects of d-Allose against Retinal Ischemia-Reperfusion Injury. *Investigative Ophthalmology & Visual Science*, 47, 1653-1657. doi: <https://doi.org/10.1167/iovs.05-1018>
- Hu, J., Huang, W., Zhang, F. Luo, X., Chen, Y., & Xie, J. (2020). Variability of Volatile Compounds in The Medicinal Plant *Dendrobium officinale* from Different Regions. *Molecules*, 25, 1-10. doi: <https://doi.org/10.3390/molecules25215046>
- Hu, L., Robert, C. A. M., Cadot, S., Zhang, X., Ye, M., Li, B., Manzo, D., Chervet, N., Steinger, T., Heijden, M. G. A., Schlaeppi, K., & Erb, M. (2018). Root exudate metabolites drive plant-soil feedbacks on growth and defense by shaping the rhizosphere microbiota. *Nature Communications*, 9(2738), 1-13. doi: [10.1038/s41467-018-05122-7](https://doi.org/10.1038/s41467-018-05122-7)
- Iglesias, A. & Meins, F. Jr. (2000). Vacuoles and plant defense. In: Robinson DG, Rogers JC (eds) Annual plant reviews: vacuolar compartments, vol 5. Sheff

- Iida, T. & Okuma, K. (2013). Properties of Three Rare Sugars D-psicose, D-allose, D-tagatose and Their Applications. *Japan Oil Chemists Society*, 13(9), 435–440.
- Ismail, N. Z., Arsal, H., Samian, M. R., & Hamdan, M. R. (2017). Determination of Phenolic and Flavonoid Contents, Antioxidants Activities and GC-MS Analysis of *Clinacanthus nutans* (Acanthaceae) in Different Locations. *Agrivita Journal of Agricultural Science*, 39(3), 335-344. doi: <http://doi.org/10.17503/agrivita.v39i3.1076>
- Jadhav, V., Kalase, V., & Patil, P. (2014). GC-MS analysis of bioactive compounds in methanolic extract of *Holigarna grahamii* (wight) Kurz. *International Journal of Herbal Medicine*, 2(4), 35-39.
- Jansen, P.C.M. (2006). *Coix lacryma-jobi* L. PROTA. [Online]. Diakses dari [https://uses.plantnet-project.org/en/Coix\\_lacryma-jobi\\_\(PROTA\)](https://uses.plantnet-project.org/en/Coix_lacryma-jobi_(PROTA))
- Jeong, J. B. & Jeong, H. J. (2010). 2-Methoxy-4-vinylphenol can induce cell cycle arrest by blocking the hyper-phosphorylation of retinoblastoma protein in benzo[a]pyrene-treated NIH3T3 cells. *Biochemical and Biophysical Research Communications*, 400(4), 752–757. doi:[10.1016/j.bbrc.2010.08.142](https://doi.org/10.1016/j.bbrc.2010.08.142)
- Jeong, J. B., Hong, S. C., Jeong, H. J., & Koo, J. S. (2011). Anti-inflammatory effect of 2-methoxy-4-vinylphenol via the suppression of NF-κB and MAPK activation, and acetylation of histone H3. *Archives of pharmacal research*, 34(12), 2109–2116. doi: <https://doi.org/10.1007/s12272-011-1214-9>
- Jiang, H., Wang, B., Li, X., Lü, E., & Li, C. (2008). A consideration of the involukrum remains of *Coix lacryma-jobi* L. (Poaceae) in the Sampula Cemetery (2000 years BP), Xinjiang, China. *Journal of Archaeological Science*, 35(5), 1311–1316. doi:[10.1016/j.jas.2007.09.006](https://doi.org/10.1016/j.jas.2007.09.006)
- Jiang, Z., Ma, Y., Li, M., Zhan, X., Zhang, X., & Wang, M. (2015). 5-Hydroxymethylfurfural protects against ER stress-induced apoptosis in GalN/TNF-α-injured L02 hepatocytes through regulating the PERK-eIF2 $\alpha$  signaling pathway. *Chinese Journal of Natural Medicines*, 13(12), 896–905. doi:[10.1016/s1875-5364\(15\)30095-9](https://doi.org/10.1016/s1875-5364(15)30095-9)
- Johnson, M. & Abugri, D. A. (2014). Occurrence, Biochemical, Antimicrobial And Health Effects Of Palmitic Acid. *Palmitic Acid: Occurrence, Biochemistry and Health Effects*. Nova Science Publishers, Inc. 18-43

- Jones, W. P. & Kinghorn, A. D. (2012). Extraction of Plant Secondary Metabolites. *Natural Products Isolation*, 864, 341–366. doi:[10.1007/978-1-61779-624-1\\_13](https://doi.org/10.1007/978-1-61779-624-1_13)
- Julianto, T. (2019). *Fitokimia Tinjauan Metabolit Sekunder dan Skrining Fitokimia*. Yogyakata: Universitas Islam Indonesia
- Jungermann, E. & Sonntag, N. O. V. (1991). *Glycerine: A Key Cosmetic Ingredient*. New York: Marcel Dekker Inc.
- Junior, I., Alevar do Nascimento, M., de Souza, R. O. M. A., Dufour, A., & Wojcieszak, R. (2020). Levoglucosan: A promising platform molecule? *Green Chemistry*, 22, 5859-5880. doi:[10.1039/d0gc01490g](https://doi.org/10.1039/d0gc01490g)
- Kaluarachchi, M., Lewis, M. R., & Lindon, J. C. (2017). Standardized Protocols for MS-Based Metabolic Phenotyping. *Encyclopedia of Spectroscopy and Spectrometry*, 224–231. doi:[10.1016/b978-0-12-409547-2.12134-1](https://doi.org/10.1016/b978-0-12-409547-2.12134-1)
- Kang, S., Kim, B., Choi, B., Lee, H. O., Kim, N., Lee, S., Kim, H. S., Shin, M. J., Nam, K., Kang, K. D., Kwon, S., Oh, T., Lee, S., & Kim, C. (2020). Genome Assembly and Annotation of Soft-Shelled Adlay (*Coix lacryma-jobi* variety *ma-yuen*) A Cereal and Medicinal Crop in The Poaceae Family. *Frontiers in Plant Science*, 11, 1-14.
- Kaur, R. & Arora, S. (2015). Alkaloids—Important Therapeutic Secondary Metabolites of Plant Origin. *J. Crit. Rev*, 2, 1–8
- Kchaou, W., Abbès, F., Blecker, C., Attia, H., & Besbes, S. (2013). Effects of extraction solvents on phenolic contents and antioxidant activities of Tunisian date varieties (*Phoenix dactylifera* L.). *Industrial Crops and Products* 45, 262–269.
- Keisotyo. (2005). *Coix lacryma-jobi inflorescence*. [Online]. Diakses dari [https://commons.wikimedia.org/wiki/File:Coix\\_lacrymajobi\\_inflorescence.jpg](https://commons.wikimedia.org/wiki/File:Coix_lacrymajobi_inflorescence.jpg)
- Kessler, A. & Halitschke, R. (2007). Specificity And Complexity: The Impact of Herbivore-Induced Plant Responses on Arthropod Community Structure. *Curr. Opin. Plant Biol.* 10(4):409–14
- Khandaker, S., Das, S., Opo FADM., Akhter, R., & Shahriar, M. (2016). In vivo pharmacological investigations of the crude extracts of *Calamus viminalis* (L.). *J Pharmacogn Phytochem*, 5(3).263-269.
- Kim, D.-H., Han, S.-I., GO, B., Oh, U. H., Kim, C.-S., Jung, Y.-H., ... & Kim, J.-H. (2019). 2-Methoxy-4-vinylphenol Attenuates Migration of Human Pancreatic

- Cancer Cells via Blockade of FAK and AKT Signaling. *Anticancer Research*, 39(12), 6685–6691. doi:[10.21873/anticanres.13883](https://doi.org/10.21873/anticanres.13883)
- Kim, J. H., Lee, H., Cho, Y., Kim, J., Chun, J., Choi, J., Lee, Y., & Jung, H. W. (2014). A Vanillin Derivative Causes Mitochondrial Dysfunction and Triggers Oxidative Stress in *Cryptococcus neoformans*. *PLoS One*, 9(2). doi: [10.1371/journal.pone.0089122](https://doi.org/10.1371/journal.pone.0089122)
- Kocacaliskan, I., Talan, I., & Terzi, I. (2006). Antimicrobial Activity of Catechol and Pyrogallol as Allelochemicals. *Zeitschrift für Naturforschung C*, 61(9-10). 6439-642
- Kumar, S. & Pandey, A. K. (2013). Phytochemical profiling of leaf, stem, and tuber Parts of *Solena amlexiacaulis* (Lam.) Gandhi using GC-MS. *International Scholarly Research Notices*, 1-16. doi: <https://doi.org/10.1155/2013/162750>
- Kumar, N. & Pruthi, V. (2014). Potential applications of ferulic acid from natural sources. *Biotechnology Reports*, 4, 86-93. doi: <http://dx.doi.org/10.1016/j.btre.2014.09.002>
- Kurniawan, H. (2014). *Hanjeli Dan Potensinya Sebagai Bahan Pangan*. [Online]. Diakses dari <http://biogen.litbang.pertanian.go.id/?p=57097>
- Kutchan, T. M. (2005). A role for intra- and intercellular translocation in natural product biosynthesis. *Curr Opin Plant Biol*, 8. 292–300
- Kushwaha, P., Yadav, S. S., Singh, V., & Dwivedi, L. K. (2019). GC-MS Analysis Of Bio-Active Compounds In Methanolic Extract Of *Ziziphus Mauritiana* Fruit. *International Journal of Pharmaceutical Sciences and Research*, 10(6), 2911-2916. doi: [http://dx.doi.org/10.13040/IJPSR.0975-8232.10\(6\).2911-16](http://dx.doi.org/10.13040/IJPSR.0975-8232.10(6).2911-16)
- Lee, H. J., Ryu, J., Park, S. H., Seo, E.-K., Han, A.-R., Lee, S. K., & Lee, C. J. (2014). Suppressive effects of coixol, glyceryl trilinoleate and natural products derived from *Coix lacryma-Jobi* var. *ma-yuen* on gene expression, production and secretion of airway MUC5AC mucin. *Archives of Pharmacal Research*, 38(5), 620–627. doi:[10.1007/s12272-014-0377-6](https://doi.org/10.1007/s12272-014-0377-6)
- Li, H., Liu, Y., Yuan, W., & Dai, X. (2009). Determination of coixol in *Coix lacryma-jobi* L.var. *mayuen* Stapf by HPLC. *West China J Pharm Sci*. doi: [24\(5\):530–532](https://doi.org/10.1007/s12272-014-0377-6)
- Li, S. (1981). *Bencao Gangmu (Material Medica, Arranged According to Drug Descriptions and Technical Aspects)*. Beijing: Public Health Press
- Li, Y. (1979). *Anatomy and morphology of Graminae*. Shanghai: Shanghai Sci Technol Press

- Li, Z.-H., Wang, Q., Ruan, X., Pan, C.-D., & Jiang, D.-A. (2010). Phenolics and Plant Allelopathy. *Molecules*, 15(12), 8933–8952. doi:[10.3390/molecules15128933](https://doi.org/10.3390/molecules15128933)
- Li, X.-X., Du, L.-D., & Du, G.-H. (2018). Arbutin. *Natural Small Molecule Drugs from Plants*, 667–670. doi:[10.1007/978-981-10-8022-7\\_107](https://doi.org/10.1007/978-981-10-8022-7_107)
- Lim, T. K. (2012). *Coix lacryma-jobi*. Edible Medicinal And Non-Medicinal Plants. *Fruits*, 5, 243–261. doi:[10.1007/978-94-007-5653-3\\_14](https://doi.org/10.1007/978-94-007-5653-3_14)
- Liu, B., Peng, X., Han, L., Hou, L., & Li, B. (2020). Effects of Exogenous Spermidine on Root Metabolism of Cucumber Seedlings under Salt Stress by GC-MS. *Agronomy*, 10(4), 459. doi:[10.3390/agronomy10040459](https://doi.org/10.3390/agronomy10040459)
- Liu, L., Duncan, N. A., Chen, X., & Cui, J. (2018). Exploitation of Job's Tears in Paleolithic and Neolithic China: Methodological Problems and Solutions. *Quaternary International*. doi: [10.1016/j.quaint.2018.11.019](https://doi.org/10.1016/j.quaint.2018.11.019)
- Liu, Q.-M., Peng, W.-X., Wu, Y.-X., Xie, X.-M., & Guang, X.-S. (2009). Analysis of Biomedical Components of *Camellia oleifera* Leaf and Kernel Hull by GC/MS. *2009 3rd International Conference on Bioinformatics and Biomedical Engineering*. doi:[10.1109/icbbe.2009.5162344](https://doi.org/10.1109/icbbe.2009.5162344)
- Liu, S., Ruan, W., Li, J., Xu, H., Wang, J., Gao, Y., & Wang, J. (2008). Biological Control of Phytopathogenic Fungi by Fatty Acids. *Mycopathologia*, 166(2), 93–102. doi:[10.1007/s11046-008-9124-1](https://doi.org/10.1007/s11046-008-9124-1)
- Loman, A. A. & Ju, L.-K. (2015). Inhibitory effects of arabitol on caries-associated microbiologic parameters of oral *Streptococci* and *Lactobacilli*. *Archives of Oral Biology*, 60(12), 1721–1728. doi:[10.1016/j.archoralbio.2015.09.004](https://doi.org/10.1016/j.archoralbio.2015.09.004)
- Lucas, A., Ao-ieong, E. S. Y., Williams, A. T., Jani, V. P., Muller, C. R., Yaicin, O., & Cabrales, P. (2019). Increased Hemoglobin Oxygen Affinity with 5-Hydroxymethylfurfural Supports Cardiac Function During Severe Hypoxia. *Frontiers in Physiology*, 10(1350). 1-15. doi: [10.3389/fphys.2019.01350](https://doi.org/10.3389/fphys.2019.01350)
- Majoul, T., Bouabdallah, F., Hammami, M., Satouri, H., Tabbene, O., Slimen, A., Elkhoui, S., Limam, S., & Aouani, E. (2019). Chemical Composition and Biological Activities of Seeds from Four *Vitis vinifera* Tunisian Varieties. *Journal of Food Chemistry and Nutrition*.
- Makowska, B., Bakera, B., & Rakoczy-Trojanowska, M. (2015). The genetic background of benzoxazinoid biosynthesis in cereals. *Acta Physiol Plant* 37(176), 1-12. doi: [10.1007/s11738-015-1927-3](https://doi.org/10.1007/s11738-015-1927-3)
- Manosroi, A., Sainakham, M., Chankhampan, C., Manosroi, W., & Manosroi, J. (2016). In vitro anti-cancer activities of Job's tears (*Coix lacryma-jobi* Linn.)

- extracts on human colon adenocarcinoma. *Saudi Journal of Biological Sciences*, 16(2): 1-23
- Mariska, I. (2013). *Metabolit Sekunder: Jalur pembentukan dan kegunaannya*. Balai Besar Penelitian Bioteknologi dan Sumberdaya Genetik.
- Markwirth. (2020). *Job's tears: Coix lacryma-jobi*. [Online]. Diakses dari <https://flora-obscura.de/en/portfolio-item/jobs-tears-coix-lacryma-jobi/>
- Mazid, M., Khan, T. A., & Mohammad, F. (2011). Role of Secondary Metabolites in Defence Mechanisms of Plants. *Biol. Med.* 3(22), 232-249
- Mendes, R. D. F. F. & Maia, E. R. S. (1995). The use of levoglucosan for the treatment of infectious or inflammatory conditions. *Patent Cooperation Treaty*. No. WO1996015796A1. World Intellectual Property Organization.
- Miyazawa, M., Anzai, J., Fujioka, J., & Isikawa, Y. (2003). Insecticidal Compounds against *Drosophila Melanogaster* from *Cornus Officinalis Sieb. Et Zucc. Natural Product Research*, 17(5), 337–339. doi:[10.1080/1057563031000072587](https://doi.org/10.1080/1057563031000072587)
- Morrison, L. R. (2000). Glycerol. *Kirk-Othmer Encyclopedia of Chemical Technology*. doi:[10.1002/0471238961.0712250313151818.a0](https://doi.org/10.1002/0471238961.0712250313151818.a0)
- Mukhriani. (2014) Ekstraksi, Pemisahan Senyawa, dan Identifikasi Senyawa Aktif. *Jurnal Kesehatan*, 7(2), 361-367
- Muniz, D. F., Dos Santos Barbosa, C. R., de Menezes, I. R. A., de Sousa, E. O., Pereira, R. L. S., Júnior, J. T. C., ... & Tintino, S. R. (2020). In vitro and in silico inhibitory effects of synthetic and natural eugenol derivatives against the NorA efflux pump in *Staphylococcus aureus*. *Food Chemistry*, 337, 127776. doi:[10.1016/j.foodchem.2020.127776](https://doi.org/10.1016/j.foodchem.2020.127776)
- Muthukumaran, J., Srinivasan, S., Venkatesan, R. S., Ramachandran, V., & Muruganathan, U. (2013). Syringic acid, a novel natural phenolic acid, normalizes hyperglycemia with special reference to glycoprotein components in experimental diabetic rats. *Journal of Acute Disease*, 2(4), 304-309. doi: [https://doi.org/10.1016/S2221-6189\(13\)60149-3](https://doi.org/10.1016/S2221-6189(13)60149-3)
- Nagata, Y., Ishizaki, I., Waki, M., Ide, Y., Hossen, M. A., Ohnishi, K., ..., & Setou, M. (2015). Palmitic acid, verified by lipid profiling using secondary ion mass spectrometry, demonstrates anti-multiple myeloma activity. *Leukemia Research*, 39(6), 638–645. doi:[10.1016/j.leukres.2015.02.011](https://doi.org/10.1016/j.leukres.2015.02.011)

- Nakamura, T., Tanaka, S., Hirooka, K., Toyoshima, T., Kawai, N., Tamiya, T., Shiraga, F., Tokuda, M., Keep, R. F., Itano, T., & Miyamoto, O. (2011). Anti-oxidative effects of d-allose, a rare sugar, on ischemia-reperfusion damage following focal cerebral ischemia in rat. *Neuroscience Letters*, 487(1), 103-106. doi: <https://doi.org/10.1016/j.neulet.2010.10.004>
- Nandikar, M. D., Kumbhalkar, B. B., & Gurav, R. V. (2018). GC-MS Analysis of Phytochemical Compounds in The Crude Methanolic Extract Roots of *Murdannia lanuginose* and *M. simplex* (Commelinaceae). *Indian Journal of Natural Products and Resources*, 9(3), 229-234.
- National Parks Board (NParks). (2020). *Coix lacryma-jobi*. [Online]. Diakses dari <https://www.nparks.gov.sg/florafaunaweb/flora/1/8/1834>
- Nes, W.D. & Zhou, W. (2001). Terpenoids: Higher. *Encyclopedia of Life Sciences*. doi: <https://doi.org/10.1038/npg.els.0001916>
- Ngo, T.V., Scarlett, C. J., Bowyer, M. C., Ngo, P. D., & Vuong, Q.V. (2017). Impact of different extraction solvents on bioactive compounds and antioxidant capacity from the root of *Salacia chinensis* L. *Journal of Food Quality*, 1-8.
- Noegrohati. (1996). *Prinsip Dasar dan Aplikasi Kromatografi Gas*. Yogyakarta: Laboratorium Analisa Kimia dan Fisika Pusat Universitas Gadjah Mada.
- Nugroho, A. (2017). *Buku Ajar: Teknologi Bahan Alam*. Banjarmasin: Lambung Mangkurat University Press
- Nur, Y., Cahyotomo, A., Nanda, & Fistoro, N. (2020). Profil GC-MS Senyawa Metabolit Sekunder dari Jahe Merah (*Zingiber officinale*) dengan Metode Ekstraksi Etil Asetat, Etanol dan Destilasi. *Jurnal Sains dan Kesehatan*, 2(3). doi: <https://doi.org/10.25026/jsk.v2i3.115>
- Nurhadianty, V., Cahyani, C., Nirwana, W. O. C., & Dewi, L. K. (2018). *Pengantar Teknologi Fermentasi Skala Industri*. Malang: UB Press
- Nurmala, T. (2011). Potensi dan Prospek Pengembangan Hanjeli (*Coix lacryma jobi* L) sebagai Pangan Bergizi Kaya Lemak untuk Mendukung Diversifikasi Pangan Menuju Ketahanan Pangan Mandiri. *Pangan*, 20(1), 41-48
- Nurmala, T., Ruminta, & Wicaksono, F. (2017). Pengembangan Pangan Lokal Hanjeli Sebagai Pangan Multifungsi dalam Rangka Pemberdayaan Petani di Lahan Marjinal. Prosiding Fakultas Pertanian UNPAD
- Oliveira, M. S. C., Morais, S. M. de, Magalhães, D. V., Batista, W. P., Vieira, I. G. P., Craveiro, A. A., & de Lima, G. P. G. (2011). Antioxidant, larvicidal and

- antiacetylcholinesterase activities of cashew nut shell liquid constituents. *Acta Tropica*, 117(3), 165–170. doi:[10.1016/j.actatropica.2010.08.003](https://doi.org/10.1016/j.actatropica.2010.08.003)
- Ormeño, E., Mévy, J. P., Vila, B., Bousquet-Mélou, A., Greff, S., Bonin, G., & Fernandez, C. (2007). Water deficit stress induces different monoterpene and sesquiterpene emission changes in Mediterranean species-Relationship between terpene emissions and plant water potential. *Chemosphere*, 67(2), 276–284. doi:[10.1016/j.chemosphere.2006.10.029](https://doi.org/10.1016/j.chemosphere.2006.10.029)
- Osbourn, A. E., Qi, X., Townsend, B., & Qin, B. (2003). Dissecting plant secondary metabolism - constitutive chemical defences in cereals. *New Phytologist*, 159(1), 101–108. doi:[10.1046/j.1469-8137.2003.00759.x](https://doi.org/10.1046/j.1469-8137.2003.00759.x)
- Paduch, R., Szerszen, M. K., Trytek, M., & Fiedurek, J. (2007). Terpenes: substances useful in human healthcare. *Arch. Immunol. Ther.* 55, 315-327. doi: <https://doi.org/10.1007/s00005-007-0039-1>
- Pagare, S., Bhatia, M., Tripathi, N., Pagare, S., & Bansal, Y. (2015). Secondary Metabolites of Plants and their Role: Overview. *Trends in Biotechnology and Pharmacy*, 9(3), 293-304.
- Pal, A., Das, D., Panda, P., Rath, M., & Sharma, T. (2015). GC-MS Analysis of bioactive compounds in the methanol extract of *Clerodendrum viscosum* leaves. *Pharmacognosy Research*, 7(1), 110.
- Papadimitropoulos, M.-E. P., Vasilopoulou, C. G., Maga-Nteve, C., & Klapa, M. I. (2018). Untargeted GC-MS Metabolomics. *Metabolic Profiling*, 1738, 133–147. doi:[10.1007/978-1-4939-7643-0\\_9](https://doi.org/10.1007/978-1-4939-7643-0_9)
- Picman, A. K. (1986). Biological activities of sesquiterpene lactones. *Biochemical systematics and Ecology*. 14. 255-281
- Pillaiyar, T., Namasivayam, V., Manickam, M., & Jung, S.-H. (2018). Inhibitors of Melanogenesis: An Updated Review. *Journal of Medicinal Chemistry*. doi:[10.1021/acs.jmedchem.7b00967](https://doi.org/10.1021/acs.jmedchem.7b00967)
- Pinelo, M., Rubilar, M., Jerez, M., Sineiro, J., & Nunez, M. J. (2005). Effect of Solvent, Temperature, and Solvent-to-Solid Ratio on the Total Phenolic Content and Antiradical Activity of Extracts from Different Components of Grape Pomace. *Agricultural and Food Chemistry*, 53. 2111-2117.
- Plantamor. (2020). Hanjeli (*Coix lacryma-jobi* L.). [Online]. Diakses dari <http://plantamor.com/species/info/coix/lacryma-jobi>
- Prayitno, S.A., J. Kusnadi, E.S. & Murtini. (2016). Antioxidant activity of red betel leaves extract (*Piper crocatum* Ruiz and Pav.) by different concentration o

- solvents. *Journal of Pharmaceutical, Biological and Chemical Science* 7(5). 1836-1843
- Pubchem. (2021). *Compound Summary. (Online)*. Diakses dari <https://pubchem.ncbi.nlm.nih.gov/compound>.
- Puspitasari, A. D. & Proyogo, L., S. (2010). Perbandingan Metode Ekstraksi Maserasi dan Sokletasi terhadap Kadar Flavonoid Total Ekstrak Etanol Daun Kersen (*Muntingia calabura*). *Publikasi Ilmiah Unhawas*
- Rada-Mendoza, M., Olano, A., & Villamiel, M. (2002). Determination of Hydroxymethylfurfural In Commercial Jams and in Fruits-Base Infant Foods. *Food Chem.* 79. 513-516. doi: [10.1016/s0308-8146\(02\)00217-0](https://doi.org/10.1016/s0308-8146(02)00217-0)
- Rahman, M. (2018). Application of Computational Methods in Isolation of Plant Secondary Metabolites. *Computational Phytochemistry*, 107–139. doi:[10.1016/b978-0-12-812364-5.00004-3](https://doi.org/10.1016/b978-0-12-812364-5.00004-3)
- Rajasekharan, R. & Nachiappan, V. (2010). Fatty Acid Biosynthesis and Regulation in Plants. *Plant Developmental Biology – Biotechnological Perspectives*, 2, 105-115. doi: [10.1007/978-3-642-04670-4\\_6](https://doi.org/10.1007/978-3-642-04670-4_6),
- Rajesh, K. B. R., Bharathb, Rao, C. V., Bhat, K.I., K.S., Bhat, C., & Bath, P. (2017). Neutralization of *Naja naja* venom induced lethality, edema and myonecrosis by ethanolic root extract of *Coix lacryma-jobi*. *Toxicology Reports*, 4, 637-645
- Ramadoss, D. P. & Sivalingam, N. (2019). Vanillin extracted from Proso and Barnyard millets induce apoptotic cell death in HT-29 human colon cancer cell line. *Nutrition and Cancer*. doi: <https://doi.org/10.1080/01635581.2019.1672763>
- Rastogi S. C. & Johansen, J. D. Significant exposures to isoeugenol derivatives in perfumes. *Contact Dermatitis*, 58(5), 278-281. doi: [10.1111/j.1600-0536.2007.01283.x](https://doi.org/10.1111/j.1600-0536.2007.01283.x)
- Ratnasari, E. (2019). *Kandungan Metabolit Akar dan Daun Hanjeli (Coix Lacryma-Jobi L.) Liar dan Budidaya dengan GC-MS*. (Skripsi). Departemen Pendidikan Biologi, Universitas Pendidikan Indonesia.
- Rohmer, M., Knani, M., Simonin, P., Sutter, B., & Sahm, H. (1993). Isoprenoid Biosynthesis In Bacteria: A Novel Pathway For The Early Steps Leading To Isopentenyl Diphosphate. *Biochem. J.* 295.517-524
- Rubab, M., Chelliah, R., Saravanakumar, K., Barathikannan, K., Wei, S., Kim, J. R., Yoo, D., Wang, M. H., & Oh, D. H. (2020). Bioactive Potential of 2-Methoxy-4-vinylphenol and Benzofuran from *Brassica oleracea* L. var.

- capitate f. rubra* (Red Cabbage) on Oxidative and Microbiological Stability of Beef Meat. *Foods (Basel, Switzerland)*, 9(5), 568. doi: <https://doi.org/10.3390/foods9050568>
- Ruiz, M. A. & Sotelo, A. (2001). Chemical Composition, Nutritive, Value, and Toxicology Evaluation Of Mexican Wild Lupins. *J. Agric. Food Chem.* 49, 5336-5339.
- Saenong, M. S. (2016). Tumbuhan Indonesia Potensial Sebagai Insektisida Nabati Untuk Mengendalikan Hama Kumbang Bubuk Jagung (*Sitophilus spp.*). *Jurnal Litbang Pertanian*, 35(3), 131-142. doi: [10.21082/jp3.v35n3.2016.p131-142](https://doi.org/10.21082/jp3.v35n3.2016.p131-142)
- Saifudin, N. (2014). *Senyawa Alam Metabolit Sekunder: Teori, Konsep, dan Teknik Pemurnian*. Yogyakarta: Deepublish
- Salau, V. F., Erukainure, O. L., Ibeji, C. U., Olasehinde, T. A., Koorbanally, N. A., & Islam, M. S. (2020). Vanillin and vanillic acid modulate antioxidant defense system via amelioration of metabolic complications linked to Fe<sup>2+</sup>-induced brain tissues damage. *Metabolic Brain Disease*, 35, 727-738. doi: <https://doi.org/10.1007/s11011-020-00545-y>
- Sampaio, B. L., Edrada-Ebel, R., & Costa, F. (2016). Effect of The Environment on The Secondary Metabolic Profile of *Tithonia diversifolia*: A Model for Environmental Metabolomics of Plants. *Scientific Reports*, 6, 1-11. doi: <http://doi.org/10.1038/srep29265>
- Sasaki, Y. & Nagano, Y. (2004). Plant acetyl-CoA carboxylase: structure, biosynthesis, regulation, and gene manipulation for plant breeding. *Biosci Biotechnol Biochem*, 68, 1175–1184.
- Setyowati, H., Zharfa, H., Angela, I., Agnes, J. K., Muawanah, S. & Aliyah, N. (2013). *Isolasi dan Standarisasi Bahan Alam Gas Chromatography Mass Spectrometry GC-MS*. Semarang: Sekolah Tinggi Ilmu Farmasi.
- Schkolnik, G. & Rudich, Y., (2006). Detection and quantification of levoglucosan in atmospheric aerosols: a review. *Anal. Bioanal. Chem.* 385, 26–33. doi: <https://doi.org/10.1007/s00216-005-0168-5>
- Senja, R.Y., Issusilaningtyas, E., Nugroho, A.K., & Setyowati, E.P. (2014). Perbandingan Metode Ekstraksi dan Variasi Pelarut terhadap Rendemen dan Aktivitas Antioksidan Ekstrak Kubis Ungu (*Brassica oleracea* L. Var. *Capitata f. Rubra*). *Traditional Medicine Journal*, 19, 43-48.

- Shrikumar, S., & Ravi, T. K. (2007). Approaches Towards Development and Promotion of Herbal Drugs. *Pharmacognosy Reviews*, 1(1), 180–184.
- Shouliang, C. & Phillips, S. (2006). *Coix Linnaeus. Flora of China*, 22: 648-649
- Sidorov, R. A., Zhukov, A. V., Pchelkin, V. P., & Tsydendambaev, V. D. (2014). Palmitic Acid in Higher Plant Lipids. Palmitic Acid: Occurrence, Biochemistry and Health Effects. *Nova Science Publishers*, Inc. 126-143
- Simoneit, B. R. T., Schauer, J. J., Nolte, C. G., Oros, D. R., Elias, V. O., Fraser, M. P., Rogge, W. F., & Cass, G. R. (1999). Levoglucosan, a tracer for cellulose in biomass burning and atmospheric particles. *Atmospheric Environment*, 33(2), 173–182. doi:[10.1016/s1352-2310\(98\)00145-9](https://doi.org/10.1016/s1352-2310(98)00145-9)
- Son, E., Kim, Y., Park, C., Park, K., Jeong, S., Park, J., & Kim, S. (2017). *Coix lacryma-jobi* var. *ma-yuen* Stapf sprout extract has anti-metastatic activity in colon cancer cells in vitro. *BMC Complementary Medicine and Therapies*, 17
- Son, E., Kim, Y., Park, C., Park, K., Jeong, S., Park, J., & Kim, S. (2019). *Coix lacryma-jobi* var. *ma-yuen* Stapf sprout extract induces cell cycle arrest and apoptosis in human cervical carcinoma cells. *BMC Complementary Medicine and Therapies*, 19
- Srinivasan, M., Sudheer, A. R., & Menon, V. P. (2007). Ferulic Acid: Therapeutic Potential Through Its Antioxidant Property. *J Clin Biochem Nutr*, 40(2), 92-100. doi: [10.3164/jcbn.40.92](https://doi.org/10.3164/jcbn.40.92)
- Stashenko, E. & Martínez, J. (2012). GC-MS Analysis of Volatile Plant Secondary Metabolites, Gas Chromatography in Plant Science, Wine Technology, Toxicology and Some Specific Application. *InTechOpen*. doi: [10.5772/32246](https://doi.org/10.5772/32246).
- Stochmal, A., Kus, J., Martyniuk, S., & Oleszek, W. (2006). Concentration of benzoxazinoids in roots of field-grown wheat (*Triticum aestivum* L.) varieties. *J. Agric. Food Chem.* 54, 1016-1022. doi: [10.1021/jf050899](https://doi.org/10.1021/jf050899)
- Sugih, A. & Hengky, H. (2013). Pengujian dan Peningkatan Masa Simpan Produk Mie Instan Berbasis Hanjeli. *Hibah Monodisiplin UNPAR 2013*.
- Suhendra, C. P., Widarta, I. W. R., & Wiadnyani, A. A. I. S. (2019). Pengaruh Konsentrasi Etanol Terhadap Aktivitas Antioksidan Ekstrak Rimpang Ilalang (*Imperata Cylindrica* (L) Beauv.) pada Ekstraksi Menggunakan Gelombang Ultrasonik. *Jurnal Ilmu dan Teknologi Pangan*, 8(1). 27-35.

- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of Extraction Solvent/Technique on the Antioxidant Activity of Selected Medicinal Plant Extracts. *Molecules*, 14(6), 2167–2180. doi:[10.3390/molecules14062167](https://doi.org/10.3390/molecules14062167)
- Sun C.S. & Wang D.Y. (1989). *Coix lacryma-jobi L.* (Jobstears): In Vitro Regeneration. In: Bajaj Y.P.S. (eds) Medicinal and Aromatic Plants II. *Biotechnology in Agriculture and Forestry*, vol 7. Springer, Berlin, Heidelberg, 99-109. doi: [https://doi.org/10.1007/978-3-642-73617-9\\_6](https://doi.org/10.1007/978-3-642-73617-9_6)
- Taiz, L. & Zeiger, E. (2002). *Plant Physiology Third Edition*. Sunderland: Sinauer Associates, Inc., Publisher
- Takigawa, H., Nakagawa, H., Kuzukawa, M., Mori, H., & Imokawa, G. (2005). Deficient Production of Hexadecenoic Acid in the Skin Is Associated in Part with the Vulnerability of Atopic Dermatitis Patients to Colonization by *Staphylococcus aureus*. *Dermatology*, 211(3), 240–248. doi:[10.1159/000087018](https://doi.org/10.1159/000087018)
- Tan, H. W., Abdul Aziz, A. R., & Aroua, M. K. (2013). Glycerol production and its applications as a raw material: A review. *Renewable and Sustainable Energy Reviews*, 27, 118–127. doi:[10.1016/j.rser.2013.06.035](https://doi.org/10.1016/j.rser.2013.06.035)
- Thoo, Y. Y., Ho, S. K., Liang, J. Y., Ho, C. W., & Tan, C. P. (2010). Effects of binary solvent extraction system, extraction time and extraction temperature on phenolic antioxidants and antioxidant capacity from mengkudu (*Morinda citrifolia*). *Food Chemistry*, 120(1), 290-295. doi:[10.1016/j.foodchem.2009.09](https://doi.org/10.1016/j.foodchem.2009.09)
- Tjokrosetio, V. C. & Budiarjo, S. B. (2017). Efek Antibakteri dan Antijamur Allicin yang Terkandung dalam Bawang Putih: Telaah Pustaka. Paper presented at Pertemuan Ilmiah Nasional Ilmu Kedokteran Gigi Anak ke-10, Jakarta, Indonesia
- Topal, F. (2019). Anticholinergic and antidiabetic effects of isoeugenol from clove (*Eugenia caryophylata*) oil. *International Journal of Food Properties*, 22(1), 583–592. doi:[10.1080/10942912.2019.1597882](https://doi.org/10.1080/10942912.2019.1597882)
- Torres, F., Eric, L., & Gonzales, K. (2016). Exploring the Mechanical Properties of Hard Botanical Structures of Two Tropical Plants. *Bioinspired, Biomimetic and Nanobiomaterials*. 5(3), 1-10. doi: [5. 10.1680/jbibn.15.00023](https://doi.org/10.1680/jbibn.15.00023).
- Velu, G., Palanichamy, V., & Rajan, A. P. (2018). Phytochemical and Pharmacological Importance of Plant Secondary Metabolites in Modern Medicine. *BioorganicPhase in Natural Food: An Overview*, 135-156.

- Vergeer, L. H. T., Aarts, T. L., & de Groot, J. D. (1995). The “wasting disease” and the effect of abiotic factors (light intensity, temperature, salinity) and infection with *Labyrinthula zosterae* on the phenolic content of *Zostera marina* shoots. *Aquatic Botany*, 52(1-2), 35–44. doi:[10.1016/0304-3770\(95\)00480-n](https://doi.org/10.1016/0304-3770(95)00480-n)
- Vickers CE, Gershenson J, Lerdau MT, & Loreto F. (2009). A unified mechanism of action for volatile isoprenoids in plant abiotic stress. *Nature Chemical Biology*, 5:283–291
- Vijayakumar, K. & Thirunanasambandham, R. (2020). 5-Hydroxymethylfurfural inhibits *Acinetobacter baumannii* biofilms: an in vitro study. *Archives of Microbiology*. doi:[10.1007/s00203-020-02061-0](https://doi.org/10.1007/s00203-020-02061-0)
- Viladomat, F. & Bastida, J. (2015). General Overview of Plant Secondary Metabolism. *Plant Biology and Biotechnology*, 539–568. doi:[10.1007/978-81-322-2286-6\\_21](https://doi.org/10.1007/978-81-322-2286-6_21)
- Vom Endt, D., Kijne, J. W., & Memelink, J. (2002). Transcription factors controlling plant secondary metabolism: what regulates the regulators?. *Phytochemistry*, 61(2), 107–114. doi:[10.1016/s0031-9422\(02\)00185-1](https://doi.org/10.1016/s0031-9422(02)00185-1)
- Wang, L., Chen, J., Xie, H., Ju, X., & Liu, R. (2013). Phytochemical Profiles and Antioxidant Activity of Adlay Varieties. *Journal of Agricultural and Food Chemistry*.
- Wiater, M. K. (2015). Production of arabitol by yeasts: current status and future prospects. *Journal of Applied Microbiology*, 119, 303–314. doi:[10.1111/jam.1280](https://doi.org/10.1111/jam.1280)
- Willy, T. (2018). Iskemia. (*Online*). Diakses dari <https://www.alodokter.com/iskemia#:~:text=Iskemia%20pada%20otak%20merupakan%20salah,kerusakan%20atau%20kematian%20sel%20otak>.
- Wink, M. (2010). *Annual plant reviews, vol 40, 2nd edn, Biochemistry of plant secondary metabolism*. Oxford: Wiley- Blackwell
- Woo, J.-H., Dapeng, L., Dapeng, L., Orita, H., Coulter, J., Tully, E., ..., & Gabrielson, E. (2007). Coix seed extract, A commonly used treatment for cancer in china, inhibits NF $\kappa$ B and protein kinase C signaling. *Cancer Biology & Therapy*, 6(12), 2005–2011. doi:[10.4161/cbt.6.12.5168](https://doi.org/10.4161/cbt.6.12.5168)

- Wu, H., Xue, R., Dong, L., Liu, T., Deng, C., Zeng, H., & Shen, X. (2019). Metabolomic Profiling of Human Urine in Hepatocellular Carcinoma Patients using Gas Chromatography/Mass Spectrometry. *Analytica Chimica Acta*, 648, 98-104.
- Wu, S., Lyu, G., & Lou, R. (2012). Applications of Chromatography Hyphenated Techniques in the Field of Lignin Pyrolysis. *Applications of Gas Chromatography*
- Yang, C.-Q., Fang, X., Wu, X.-M., Mao, Y.-B., Wang, L.-J., & Chen, X.-Y. (2012). Transcriptional Regulation of Plant Secondary Metabolism. *Journal of Integrative Plant Biology*, 54(10), 703–712. doi:[10.1111/j.1744-7909.2012.01161.x](https://doi.org/10.1111/j.1744-7909.2012.01161.x)
- Yen, J. (2011). *Coix lacryma-jobi*. [Online]. Diakses dari <https://www.nparks.gov.sg/florafaunaweb/flora/1/8/1834#gallery-9>
- Yu, F., Li Y., Zhang, J., & Liu C. (2014). *Coix lacryma-jobi* L. var. *ma-yuen* (Roman.) Stapf (Yiyiren, Jobstears). in Dietary Chinese Herbs: Chemistry, Pharmacology, and Clinical Evidence. *Springer Wien Heidelberg New York, Dordrecht, London*: 339-346.
- Yu, X., Zhao, M., Liu, F., Zeng, S., & Hu, J. (2013). Identification of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose-histidine Maillard reaction products. *Food Research International*, 51(1), 397–403. doi:[10.1016/j.foodres.2012.12.044](https://doi.org/10.1016/j.foodres.2012.12.044)
- Yu, Q., Ye, G., Li, R., Li, T., & Huang, S. (2021). Extraction And Characterization Of Cycloartenol Isolated From Stems And Leaves of *Coix Lacryma-jobi* L. And Its Potential Cytotoxic Activity. *Research Square*, 1-16. doi: <https://doi.org/10.21203/rs.3.rs-611931/v1>
- Zasada, I. A., Meyer, S. L. F, Halbrendt, J.M, & Rice, C. (2005). Activity of hydroxamic acids from Secale cereale against the plant-parasitic nematodes *Meloidogyne incognita* and *Xiphinema americanum*. *Phytopathology* 95(10), 1116–1121. doi: [10.1094/PHYTO-95-1116](https://doi.org/10.1094/PHYTO-95-1116)
- Zheng, R., Su, S., Li, J., Zhao, Z., Wei, J., Fu, X., & Liu, R. H. (2017). Recovery of phenolics from the ethanolic extract of sugarcane (*Saccharum officinarum* L.) bagasse and evaluation of the antioxidant and antiproliferative activities. *Industrial Crops and Products*, 107, 360–369. doi:[10.1016/j.indcrop.2017.05.050](https://doi.org/10.1016/j.indcrop.2017.05.050)

- Zhang, L., Shan, K., Tang, R., & Putheti. (2009). Ultrasound-assisted extraction flavonoid of lotus (*Nelumbo nucifera* Gaertn) leaf and evaluation of its anti-fatigue activity. *International Journal of Physical Science* 4(8). 418-422.
- Zhang, Q., Lin, L., & Ye, Wen. (2018). Techniques for extraction and isolation of natural products: a comprehensive review. *Chin Med*, 13(20). doi: <https://dx.doi.org/10.1186%2Fs13020-018-0177-x>
- Zhao, L., Chen, J., Su, J., Li, L., Hu, S., Li, B., & Chen, T. (2013). In Vitro Antioxidant and Antiproliferative Activities of 5-Hydroxymethylfurfural. *Journal of Agricultural and Food Chemistry*, 61(44), 10604–10611. doi:10.1021/jf403098y
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