

**KANDUNGAN METABOLIT DAN AKTIVITAS ANTIOKSIDAN PADA
KALUS DAN PLANLET TANAMAN TEBU (*Saccharum officinarum* L.)**

SKRIPSI

diajukan untuk memenuhi sebagian syarat untuk memperoleh gelar Sarjana Sains
Program Studi Biologi



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memperoleh gelar Sarjana Sains pada Program Studi Biologi Fakultas Pendidikan
Matematika dan Ilmu Pengetahuan Alam

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LEMBAR PERNYATAAN

Dengan ini saya menyatakan bahwa skripsi dengan judul “**Kandungan Metabolit dan Aktivitas Antioksidan pada Kalus dan Planlet Tanaman Tebu (*Saccharum officinarum* L.)**” ini beserta seluruh isinya adalah benar-benar karya saya sendiri. Saya tidak melakukan penjiplakan atau pengutipan dengan cara-cara yang tidak sesuai dengan etika ilmu yang berlaku dalam masyarakat keilmuan. Atas pernyataan ini, saya siap menanggung risiko/sanksi apabila di kemudian hari ditemukan adanya pelanggaran etika keilmuan atau ada klaim dari pihak lain terhadap keaslian karya saya ini.

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ABSTRAK

Tebu (*Saccharum officinarum* L.) telah banyak digunakan untuk mengobati berbagai penyakit karena kandungan metabolit yang dimilikinya. Kultur jaringan merupakan salah satu metode yang dapat digunakan untuk memproduksi metabolit dan merupakan alternatif bioteknologi sebagai bentuk strategi konservasi. Dalam memenuhi kebutuhan nutrisinya maka diperlukan teknik subkultur. Perbedaan media kultur dan subkultur yang berulang dapat mempengaruhi kandungan metabolit dan aktivitas antioksidan yang dihasilkan. Penelitian ini bertujuan untuk mendapatkan informasi mengenai kandungan metabolit serta aktivitas antioksidan pada kalus tebu yang dikultur pada media yang sama dan pada planlet tebu yang dikultur pada media berbeda dengan frekuensi subkultur yang berbeda. Kalus dan planlet tebu diambil dari Balai Pengembangan dan Produksi Benih Perkebunan (BPPBP) Jawa Barat, dan diekstrak menggunakan metode maserasi dengan pelarut etanol p.a 70%. Metabolit dianalisis dengan menggunakan *Gas Chromatography Mass-Spectrometry* (GC-MS) dan aktivitas antioksidan dianalisis menggunakan metode 2,2-difenil-1-pikrilhidrazil (DPPH). Hasil penelitian menunjukkan kalus subkultur ke-3 dan ke-4 yang dikultur pada media MS + 2,4-D mengandung 2 metabolit, planlet subkultur ke-10 yang dikultur pada media MS + Kinetin + Air Kelapa mengandung 5 metabolit, dan planlet subkultur ke-11 yang dikultur pada media MS + Kinetin + TDZ mengandung 7 metabolit. Aktivitas antioksidan pada kalus dan planlet tebu tergolong kategori sangat lemah. Penelitian ini menunjukkan adanya perbedaan kandungan metabolit dan aktivitas antioksidan pada kalus yang dikultur pada media sama dan planlet yang dikultur pada media berbeda dengan frekuensi subkultur berbeda.

Kata Kunci : Kalus, Planlet, Metabolit, Antioksidan, Subkultur.

METABOLITE CONTENT AND ANTIOXIDANT ACTIVITY IN CALLUS AND PLANTLETS OF SUGARCANE (*Saccharum officinarum* L.)

ABSTRACT

*Sugarcane (*Saccharum officinarum* L.) has been widely used to cure various diseases due to its metabolite content. Tissue culture is one method that can be used to produce metabolites and is an alternative biotechnology as a form of conservation strategy. In order to meet its nutritional needs, subculture techniques are needed. Differences in culture media and repeated subcultures can affect the metabolite content and antioxidant activity produced. This study aims to obtain information regarding metabolite content and antioxidant activity in sugarcane callus cultured on the same media and on sugarcane plantlets cultured on different media with different subculture frequencies. Sugarcane callus and plantlets were taken from Balai Pengembangan dan Produksi Benih Perkebunan (BPPBP) Jawa Barat, and extracted using the maceration method with 70% p.a. ethanol solvent. Metabolites were analyzed using Gas Chromatography Mass-Spectrometry (GC-MS) and antioxidant activity was analyzed using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method. The results showed that the 3rd and 4th subculture callus cultured on MS + 2,4-D media contained 2 metabolites, the 10th subculture plantlet cultured on MS + Kinetin + Coconut Water media contained 5 metabolites, and the 11th subculture plantlet cultured on MS + Kinetin + TDZ media contained 7 metabolites. Antioxidant activity in sugarcane callus and plantlets classified as very weak category. This study showed differences in metabolite content and antioxidant activity in callus cultured on the same media and plantlets cultured on different media with different subculture frequencies.*

Keywords: *Callus, Plantlet, Metabolite, Antioxidant, Subculture.*

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DAFTAR PUSTAKA

- Abubakar, A. R. & Haque, M. (2020). Preparation of Medicinal Plants: Basic Extraction and Fractionation Procedures for Experimental Purposes. *Journal of Pharmacy and Bioallied Sciences*, 12(1), 1-10. doi: [10.4103/jpbs.JPBS.175.19](https://doi.org/10.4103/jpbs.JPBS.175.19)
- Afgatiani, P. M., Husni, A., & Budhiyanti, S. A. (2020). Aktivitas Antioksidan Bubuk *Sargassum hystrix* selama Penyimpanan pada Suhu Berbeda. *agriTECH*, 40(3), 175-181. doi: [10.22146/agritech.18134](https://doi.org/10.22146/agritech.18134)
- Aguilar-Rivera, N. (2019). A Framework for the Analysis of Socioeconomic and Geographic Sugarcane Agroindustry Sustainability. *Socio-Economic Planning Sciences*, 66, 149-160. doi: <https://doi.org/10.1016/j.seps.2018.07>
- Ahmad, N., Fazal, H., Abbasi, B. H., Rahmah, I. U., Anwar, S., Khan, M.A., ... Khan, K. Y. (2011). DPPH-scavenging Antioxidant Potential in Regenerated Tissues of *Stevia rebaudiana*, *Citrus sinensis* and *Saccharum officinarum*. *Journal of Medicinal Plants Research*, 5(14), 3293-3297.
- Ahmad, N., Fazal, H., Abbasi, B. H., Rashid, M., Mahmood, T., & Fatima, N. (2010). Efficient Regeneration and Antioxidant Potential in Regenerated Tissues of *Piper nigrum* L. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 102, 129-134. doi: [10.1007/s11240-010-9712-x](https://doi.org/10.1007/s11240-010-9712-x)
- Akafity, G., Kumi, N., & Ashong, J. (2024). Diagnosis and Management of Malaria in the Intensive Care Unit. *Journal of Intensive Medicine*, 4(01), 3-15. doi: <https://doi.org/10.1016/j.jointm.2023.09.002>
- Ali, S. E., El Gedaily, R. A., Mocan, A., Farag, M. A., & El-Seedi, H. R. (2019). Profiling Metabolites and Biological Activities of Sugarcane (*Saccharum officinarum* Linn.) Juice and Its Product Molasses Via a Multiplex Metabolomics Approach. *Molecules*, 24(5), 934. doi: [10.3390/molecules24050934](https://doi.org/10.3390/molecules24050934)
- Al-Marzoqi, A. H., Hameed, I. H., & Idan, S. A. (2015). Analysis of Bioactive Chemical Components of Two Medicinal Plants (*Coriandrum sativum* and *Melia azedarach*) Leaves Using Gas Chromatography-Mass Spectrometry (GC-MS). *African Journal of Biotechnology*, 14(40), 2812-2830.
- Alqurashi, A. S., Al Masoudi, L. M., Hamdi, H., & Abu Zaid, A. (2022). Chemical Composition and Antioxidant, Antiviral, Antifungal, Antibacterial and Anticancer Potentials of *Opuntia ficus-indica* Seed Oil. *Molecules*, 27(17), 5453. doi: <https://doi.org/10.3390/molecules27175453>
- Anggraito, Y. U., Susanti, R., Iswari, R.S. Yuniastuti, A., Lisdiana., Nugrahaningsih., Habibah., N.A., & Bintari, S.H. (2019). *Metabolit Sekunder dari Tanaman : Aplikasi dan Produksi*. Semarang : Universitas Negeri Semarang.
- Antari, N. M. R. O., Wartini, N. M., & Mulyani, S. (2015). Pengaruh Ukuran Partikel dan Lama Ekstraksi terhadap Karakteristik Ekstrak Warna Alami

- Buah Pandan (*Pandanus tectorius*). *Jurnal Rekayasa dan Manajemen Agroindustri*, 3(4), 30-40.
- Anwar, T., Qureshi, H., Mahnashi, M. H., Kabir, F., Parveen, N., Ahmed, D., ... & Alhaider, H. A. (2021). Bioherbicidal Ability and Weed Management of Allelopathic methyl esters from *Lantana camara*. *Saudi Journal of Biological Sciences*, 28(8), 4365-4374. doi: <https://doi.org/10.1016/j.sjbs.2021.04.026>
- Apak, R., Gorinstein, S., Böhm, V., Schaich, K. M., Özyürek, M., & Güçlü, K. (2013). Methods of Measurement and Evaluation of Natural Antioxidant Capacity/Activity (IUPAC Technical Report). *Pure and Applied Chemistry*, 85(5), 957-998. doi: <http://dx.doi.org/10.1351/PAC-REP-12-07-15>
- Araújo, L. A. D., Addor, F., & Campos, P. M. B. G. M. (2016). Use of Silicon for Skin and Hair Care: An Approach of Chemical Forms Available and Efficacy. *Anais brasileiros de dermatologia*, 91, 0331-0335. doi: <http://dx.doi.org/10.1590/abd1806-4841.20163986>
- Arora, S. & Kumar, G. (2017). Gas Chromatography-Mass Spectrometry (GC-MS) Determination of Bioactive Constituents from the Methanolic and Ethyl Acetate Extract of *Cenchrus setigerus* Vahl (Poaceae). *Pharma Innov J*, 6(11), 635-40.
- Arulandhu, A. C. M. A., Viji. M., Aishwariya, S., Vijayalakshmi, P., Rajagopal, P., & Rajalakshmi, M. (2025). Exploring the Antioxidant Activity of Phytol from the *Scoparia dulcis* Through In-silico Analysis. *Texila International Journal of Public Health*. doi: [10.21522/TIJPH.2013.SE.25.01.Art013](https://doi.org/10.21522/TIJPH.2013.SE.25.01.Art013)
- Ashar, J. R., Farhanah, A., Hamzah, P., Ismayanti, R., Tuhuteru, S., Yusuf, R., Yulianti, R. & Mardaleni, M. (2023). *Pengantar Kultur Jaringan Tanaman*. Bandung : Widina Media Utama.
- Asih, D. J., Warditiani, N. K., & Wiarsana, I. G. S. (2022). Review artikel: Aktivitas Antioksidan Ekstrak Amla (*Phyllanthus emblica/Embolica officinalis*). *Humantech: Jurnal Ilmiah Multidisiplin Indonesia*, 1(6), 674-687. doi: <https://doi.org/10.32670/ht.v1i6.1533>
- Astuti, W. Y. & Respatie, D. W. (2022). Kajian Senyawa Metabolit Sekunder pada Mentimun (*Cucumis sativus* L.). *Vegetalika*, 11(2), 122-134. doi: <https://doi.org/10.22146/veg.60886>
- Badan Pusat Statistik. (2023). *Luas Tanaman Perkebunan Besar Menurut Jenis Tanaman (Ribuan Hektar), 2021-2023*. [Online]. Diakses dari <https://www.bps.go.id/id/statistics-table/2/MTg0NyMy/luas-tanaman-perkebunan-besar-menurut-jenis-tanaman.html>.
- Badaring, D. R., Sari, S. P. M., Nurhabiba, S., Wulan, W., & Lembang, S. A. R. (2020). Uji Ekstrak Daun Maja (*Aegle marmelos* L.) terhadap Pertumbuhan Bakteri *Escherichia coli* dan *Staphylococcus aureus*. *Indonesian Journal of Fundamental Sciences*, 6(1), 16.

- Badawy, S., Liu, Y., Guo, M., Liu, Z., Xie, C., Marawan, M. A., ... & Martínez, M. A. (2023). Conjugated Linoleic Acid (CLA) as A Functional Food: Is it Beneficial or Not? *Food Research International*, 172, 113158. doi: <https://doi.org/10.1016/j.foodres.2023.113158>
- Badoni, R., Semwal, D. K., & Rawat, U. (2010). Fatty Acid Composition and Antimicrobial Activity of *Celtis australis* L. Fruits. *Journal of Scientific Research*, 2(2), 397-402. doi: [10.3329/jsr.v2i2.4056](https://doi.org/10.3329/jsr.v2i2.4056)
- Bairu, M. W., Aremu, A. O., & Van Staden, J. (2011). Somaclonal Variation in Plants: Causes and Detection Methods. *Plant Growth Regulation*, 63, 147-173. doi: [10.1007/s10725-010-9554-x](https://doi.org/10.1007/s10725-010-9554-x)
- Baliyan, S., Mukherjee, R., Priyadarshini, A., Vibhuti, A., Gupta, A., Pandey, R. P., & Chang, C. M. (2022). Determination of Antioxidants by DPPH Radical Scavenging Activity and Quantitative Phytochemical Analysis of *Ficus religiosa*. *Molecules*, 27(4), 1326. doi: <https://doi.org/10.3390/molecules27041326>
- Bazinet, R. P. & Layé, S. (2014). Polyunsaturated Fatty Acids and their Metabolites in Brain Function and Disease. *Nature reviews neuroscience*, 15(12), 771-785.
- Beppu, F., Hosokawa, M., Tanaka, L., Kohno, H., Tanaka, T., & Miyashita, K. (2006). Potent Inhibitory Effect of trans9, trans11 isomer of Conjugated Linoleic Acid on the Growth of Human Colon Cancer Cells. *The Journal of Nutritional Biochemistry*, 17(12), 830-836. doi: [10.1016/j.jnutbio.2006.01.007](https://doi.org/10.1016/j.jnutbio.2006.01.007)
- Bharathithasan, M., Kotra, V., Abbas, S. A., & Mathews, A. (2024). Review on Biologically Active Natural Insecticides from Malaysian Tropical Plants against *Aedes aegypti* and *Aedes albopictus*. *Arabian Journal of Chemistry*, 17(1), 105345. doi: <https://doi.org/10.1016/j.arabjc.2023.105345>
- Biswas, A., Dey, S., Huang, S., Xiao, A., Birhanie, Z. M., Deng, Y., ... & Li, D. (2022). Phenolic Profiling and Antioxidant Activity of Different Anatomical Parts of *C. Olitorius* and *C. Capsularis* During Different Phonological Stages. *SSRN Electronic Journal*. doi: [10.2139/ssrn.4254448](https://doi.org/10.2139/ssrn.4254448)
- Blankson, H., Stakkestad, J. A., Fagertun, H., Thom, E., Wadstein, J., & Gudmundsen, O. (2000). Conjugated Linoleic Acid Reduces Body Fat Mass in Overweight and Obese Humans. *The Journal of nutrition*, 130(12), 2943-2948.
- Boligon, A. A., Machado, M. M., & Athayde, M. L. (2014). Technical Evaluation of Antioxidant Activity. *Med. Chem*, 4(7), 517-522. doi: [10.4172/2161-0444.1000188](https://doi.org/10.4172/2161-0444.1000188)
- Breeta, R. D. I. E., Grace, V. M. B., & Wilson, D. D. (2021). Methyl Palmitate—A Suitable Adjuvant for Sorafenib Therapy to Reduce in vivo Toxicity and to Enhance Anti-cancer Effects on Hepatocellular Carcinoma Cells. *Basic & Clinical Pharmacology & Toxicology*, 128(3), 366-378. doi: [10.1111/bcpt.13525](https://doi.org/10.1111/bcpt.13525)

- Buhaescu, I. & Izzedine, H. (2007). Mevalonate Pathway: A Review of Clinical and Therapeutical Implications. *Clinical biochemistry*, 40(9-10), 575-584. doi: <https://doi.org/10.1016/j.clinbiochem.2007.03.016>
- Calabrese, M. N., Tianco, C. A. D., Garate, A. M. C., Tan, M. C. S., Ting, J., Galian, R. A. F., ... & Chang, A. C. G. (2024). Evaluation of Spirulina spp. Crude Extract Revealed Antimicrobial, Antioxidant and UV Photoprotective Properties. *Trends in Sciences*, 21(6), 7582-7582. doi: <https://doi.org/10.48048/tis.2024.7582>
- Candido, C.J., Figueiredo, P.S., Silva, R. D. C., Portugal, L.C., Jaques, J. A. D. S., Almeida, J. A., ... & Hiane, P. A. (2019). Protective Effect of α -linolenic Acid on Non-alcoholic Hepatic Steatosis and Interleukin-6 and-10 in Wistar Rats. *Nutrients*, 12(1), 9. doi:10.3390/nu12010009
- Candraningrat, I. D. A. A., Santika, A. A. G. J., Dharmayanti, I. A. M. S., & Prayascita, P. W. (2021). Review Kemampuan Metode GC-MS dalam Identifikasi Flunitrazepam terkait dengan Aspek Forensik dan Klinik. *Jurnal Kimia*, 15(1), 12. doi: <https://doi.org/10.24843/JCHEM.2021.v15.i01.p03>
- Carlisle, S. M., Trainor, P. J., Yin, X., Doll, M. A., Stepp, M. W., States, J. C., ... & Hein, D. W. (2016). Untargeted Polar Metabolomics of Transformed MDA-MB-231 Breast Cancer Cells Expressing Varying Levels of Human Arylamine N-acetyltransferase 1. *Metabolomics*, 12, 1-12. doi: <https://doi.org/10.1007/s11306-016-1056-z>
- Carvalho, M. J., Oliveira, A. L., Pedrosa, S. S., Pintado, M., & Madureira, A. R. (2021). Potential of Sugarcane Extracts as Cosmetic and Skincare Ingredients. *Industrial Crops and Products*, 169, 113625. doi: <https://doi.org/10.1016/j.indcrop.2021.113625>
- Chairunnisa, S., Wartini, N. M., & Suhendra, L. (2019). Pengaruh Suhu dan Waktu Maserasi terhadap Karakteristik Ekstrak Daun Bidara (*Ziziphus mauritiana* L.) sebagai Sumber Saponin. *Jurnal Rekayasa Dan Manajemen Agroindustri*, 7(4), 551-560.
- Chanchula, N., Jaruwattanaphan, T., & Jala, A. (2014). Differential Effects of Sucrose and Plant Growth Regulator on Shoot Multiplication and Bulbil Formation in *Oxalis Versicolour* In vitro. *ITJEMAST*, 5(4), 227-234.
- Chandra, C. & Wahyuniari, I. A. I. (2023). The Potential Effect of Alpha-linolenic Acid against Oxidative Stres in Non-alcoholic Fatty Liver Disease. *Indonesia Journal of Biomedical Science*, 17(2), 230-234.
- Charlet, R., Le Danvic, C., Sendid, B., Nagnan-Le Meillour, P., & Jawhara, S. (2022). Oleic acid and Palmitic Acid from *Bacteroides thetaiotaomicron* and *Lactobacillus johnsonii* Exhibit Anti-Inflammatory and Antifungal Properties. *Microorganisms*, 10(9), 1803. doi: <https://doi.org/10.3390/microorganisms10091803>
- Chatri, M., Jumjunidang, J., Aini, Z., & Suryendra, F. D. (2022). Aktivitas Antifungi Ekstrak Daun *Melastoma malabathricum* terhadap *Fusarium oxysporum* dan

- Sclerotium rolfsii* secara In vitro. *Jurnal Agrotek Tropika*, 10(3), 395-401. doi: <http://dx.doi.org/10.23960/jat.v10i3.5713>
- Chaturvedi, S. & Gupta, P. (2021). Plant Secondary Metabolites for Preferential Targeting among Various Stresors of Metabolic Syndrome. *Studies in Natural Products Chemistry*, 71, 221-261. doi: <https://doi.org/10.1016/B978-0-323-91095-8.00012-X>
- Chaudhari, G. R., Prakash, K., Patel, S. R., Chavda, D., Shekhawat, N., Ragi, S., & DS, S. R. (2023). Secondary Metabolites Production in Plant Tissue Culture. *The Pharma Innovation*, 12(9):2198-2208. doi: <https://doi.org/10.22271/tpi.2023.v12.i9Sac.23144>
- Chen, Y., Yamori, W., Tanaka, A., Tanaka, R., & Ito, H. (2021). Degradation of the Photosystem II Core Complex is Independent of Chlorophyll Degradation Mediated by Stay-Green Mg²⁺ Dechelataase in *Arabidopsis*. *Plant Science*, 307, 110902. doi: <https://doi.org/10.1016/j.plantsci.2021.110902>
- Cronquist, A. (1981). *An Integrated System of Classification of Flowering Plants*. New York : Columbia University Press.
- Cui, C., Zhu, L., Wang, Q., Liu, R., Xie, D., Guo, Y., ... & Jiang, P. (2022). A GC–MS-based Untargeted Metabolomics Approach for Comprehensive Metabolic Profiling of Vancomycin-Induced Toxicity in Mice. *Heliyon*, 8(7). doi: <https://doi.org/10.1016/j.heliyon.2022.e09869>
- Daba, G. M., Elkhateeb, W., ELDien, A. N., Fadl, E., Elhagrasi, A., Fayad, W., & Wen, T. C. (2020). Therapeutic Potentials of n-hexane Extracts of the Three Medicinal Mushrooms Regarding their Anti-Colon Cancer, Antioxidant, And Hypocholesterolemic Capabilities. *Biodiversitas Journal of Biological Diversity*, 21(6). doi: [10.13057/biodiv/d210615](https://doi.org/10.13057/biodiv/d210615)
- Dalimunthe, C. I. & Rachmawan, A. (2017). Prospek Pemanfaatan Metabolit Sekunder Tumbuhan sebagai Pestisida Nabati untuk Pengendalian Patogen pada Tanaman Karet. *Warta Perkaretan*, 36(1), 15-28.
- Damayanti, A., Hakim, A. R., & Saputri, R. (2023). GC-MS Analysis of Metabolite Composition in Edible Bird's Nest From Jenamas Central Kalimantan. *International Journal of Pharmacy and Applied Health Sciences*, 1(1), 6-9.
- Damayanti, D., Sudarsono, S., Mariska, I., & Herman, M. (2007). Regenerasi Pepaya Melalui Kultur In vitro. *Jurnal AgroBiogen*, 3(2), 49-54. doi: <http://dx.doi.org/10.21082/jbio.v3n2.2007.p49-54>
- Dasenaki, I., Betsi, P. C., Raptopoulos, D., & Konstantopoulou, M. (2022). Insecticidal Effect of *Pistacia lentiscus* (Anacardiaceae) Metabolites against *Lobesia botrana* (Lepidoptera: Tortricidae). *Agronomy*, 12(4), 755. doi: <https://doi.org/10.3390/agronomy12040755>
- Debeaujon, I., Peeters, A. J., Léon-Kloosterziel, K. M., & Koornneef, M. (2001). The TRANSPARENT TESTA12 Gene of *Arabidopsis* Encodes a Multidrug Secondary Transporter-Like Protein Required for Flavonoid Sequestration in

- Vacuoles of The Seed Coat Endothelium. *The Plant Cell*, 13(4), 853-871. doi: <https://doi.org/10.1105/tpc.13.4.853>
- Del Cueto, J., Ionescu, I. A., Pičmanová, M., Gericke, O., Motawia, M. S., Olsen, C. E., Campoy, J.A., Dicenta, F., Moller, B.L. & Sánchez-Pérez, R. (2017). Cyanogenic Glucosides and Derivatives in Almond and Sweet Cherry Flower Buds from Dormancy to Flowering. *Frontiers in Plant Science*, 8, 800. doi: <https://doi.org/10.3389/fpls.2017.00800>
- Dewick, P.M. (2009). *Medicinal Natural Products: A Biosynthetic Approach*, 3rd Edition. West Sussex, UK: John Wiley & Sons, Ltd.
- Dhaniaputri, R., Suwono, H., Amin, M., & Lukiati, B. (2022, May). Introduction to Plant Metabolism, Secondary Metabolites Biosynthetic Pathway, and In-Silico Molecular Docking for Determination of Plant Medicinal Compounds: An Overview. In *7th International Conference on Biological Science (ICBS 2021)*. 373-382. Atlantis Press.
- Dias, M. I., Sousa, M. J., Alves, R. C., & Ferreira, I. C. (2016). Exploring Plant Tissue Culture to Improve the Production of Phenolic Compounds: A Review. *Industrial crops and products*, 82, 9-22. doi: <https://doi.org/10.1016/j.indcrop.2015.12.016>
- Dinesh Babu, K. S., Janakiraman, V., Palaniswamy, H., Kasirajan, L., Gomathi, R., & Ramkumar, T. R. (2022). A Short Review on Sugarcane: Its Domestication, Molecular Manipulations and Future Perspectives. *Genetic Resources and Crop Evolution*, 69(8), 2623-2643. doi: <https://doi.org/10.1007/s10722-022-01430-6>
- Dinurrosifa, R. S. (2022). Evaluasi Kadar Flavonoid Total pada Ekstrak Etanol Daun Keji Beling (*Strobilanthes crispus bi*) secara Spektrofotometri Visible. Repository Stifar.
- Dommels, Y. E., Haring, M. M., Keestra, N. G., Alink, G. M., van Bladeren, P. J., & van Ommen, B. (2003). The Role of Cyclooxygenase in n-6 and n-3 Polyunsaturated Fatty Acid Mediated Effects on Cell Proliferation, PGE 2 Synthesis and Cytotoxicity in Human Colorectal Carcinoma Cell Lines. *Carcinogenesis*, 24(3), 385-392.
- Efferth, T. (2019). Biotechnology Applications of Plant Callus Cultures. *Engineering*, 5(1), 50-59. <https://doi.org/10.1016/j.eng.2018.11.006>
- Elshafie, H. S., Camele, I., & Mohamed, A. A. (2023). A Comprehensive Review on the Biological, Agricultural And Pharmaceutical Properties of Secondary Metabolites Based-Plant Origin. *International Journal of Molecular Sciences*, 24(4), 3266. doi: <https://doi.org/10.3390/ijms24043266>
- Eramma, N. & Patil, S. J. (2023). Exploration of the Biomolecules in Roots of *Flacourtia indica* (Burm. F) Merr. Methanol Extract by Chromatography Approach. *Letters in Applied NanoBioscience*, 12(4), 166-177. doi: <http://dx.doi.org/10.33263/LIANBS124.166>

- Ergina., Nuryanti, S., & Pursitasari, D. (2014). Uji Kualitatif Senyawa Metabolit Sekunder Pada Daun Palado (*Agave angustifolia*) yang Diekstraksi dengan Pelarut Air dan Etanol. *Jurnal Akademika Kimia*, 3(3), 165-172.
- Espinosa-Leal, C. A., Puente-Garza, C. A., & García-Lara, S. (2018). In Vitro Plant Tissue Culture: Means for Production of Biological Active Compounds. *Planta*, 248, 1-18. doi: <https://doi.org/10.1007/s00425-018-2910-1>
- Fadzir, U. A., Mokhtar, K. I., Mustafa, B. E., & Darnis, D. S. (2018). Evaluation of Bioactive Compounds on Different Extracts of *Linum usitatissimum* and its antimicrobial properties against selected oral pathogens. *Makara Journal of Health Research*, 22(3), 3.
- Fajrih, N., Wiryawan, K. G., Sumiati, S., Syahpura, S. K., & Winarsih, W. (2022). Identification of Bioactive Compounds of Banana corm (*Musa paradisiaca*) using GC-MS and Its Inhibitory Effect against Pathogenic Bacteria. *Biodiversitas Journal of Biological Diversity*, 23(1). doi: [10.13057/biodiv/d230125](https://doi.org/10.13057/biodiv/d230125)
- Fakhrusy, F. (2020). Optimalisasi Metode Maserasi untuk Ekstraksi Tanin Rendemen Tinggi. *Menara Ilmu: Jurnal Penelitian dan Kajian Ilmiah*, 14(2).
- Fejér, J., Gruľová, D., De Feo, V., Üргеová, E., Obert, B., & Preťová, A. (2018). *Mentha* × *piperita* L. Nodal Segments Cultures and Their Essential Oil Production. *Industrial Crops and Products*, 112, 550-555. doi: <https://doi.org/10.1016/j.indcrop.2017.12.055>
- Fiehn, O. (2016). Metabolomics by Gas Chromatography–Mass Spectrometry: Combined Targeted and Untargeted Profiling. *Current protocols in molecular biology*, 114(1), 30-4. doi: <https://doi.org/10.1002/0471142727.mb3004s114>
- Food and Agriculture Organization. (2022). *Crops and Livestock Products*. [Online]. Diakses dari <https://www.fao.org/faostat/en/>.
- Ganesan, T., Subban, M., Christopher Leslee, D. B., Kuppannan, S. B., & Seedevis, P. (2024). Structural Characterization of n-hexadecanoic Acid from the Leaves of *Ipomoea eriocarpa* and Its Antioxidant and Antibacterial Activities. *Biomass Conversion and Biorefinery*, 14(13), 14547-14558. doi: <https://doi.org/10.1007/s13399-022-03576-w>
- Gardeazabal, L. & Izeta, A. (2024). Elastin and Collagen Fibres in Cutaneous Wound Healing. *Experimental Dermatology*, 33(3), e15052. doi: <https://doi.org/10.1111/exd.15052>
- Gawron-Skarbek, A., Guligowska, A., Prymont-Przyimińska, A., Nowak, D., & Kostka, T. (2023). The Anti-Inflammatory and Antioxidant Impact of Dietary Fatty Acids in Cardiovascular Protection in Older Adults may be Related to Vitamin C Intake. *Antioxidants*, 12(2), 267. doi: <https://doi.org/10.3390/antiox12020267>
- Ghaneian, M. T., Ehrampoush, M. H., Jebali, A., Hekmatimoghaddam, S., & Mahmoudi, M. (2015). Antimicrobial Activity, Toxicity and Stability of Phytol

- as A Novel Surface Disinfectant. *Environmental Health Engineering and Management Journal*, 2(1), 13-16.
- Goelzer Neto, C. F., do Nascimento, P., da Silveira, V. C., de Mattos, A. B. N., & Bertol, C. D. (2022). Natural Sources of Melanogenic Inhibitors: A Systematic Review. *International Journal of Cosmetic Science*, 44(2), 143-153. doi: <https://doi.org/10.1111/ics.12763>
- Gordon, M., Pokorny, J., & Yanishlieve. (2001). *Antioxidants in Food*. New York : CRC Press.
- Goto, T., Takahashi, N., Hirai, S., & Kawada, T. (2010). Various Terpenoids derived from Herbal and Dietary Plants Function as PPAR Modulators and Regulate Carbohydrate and Lipid Metabolism. *PPAR research*, 2010(1), 483958. doi: [10.1155/2010/483958](https://doi.org/10.1155/2010/483958)
- Gross, J.H. (2017). *Mass Spectrometry. A Textbook Third Edition*. New York : Springer.
- Gulcin, İ. & Alwasel, S. H. (2023). DPPH Radical Scavenging Assay. *Processes*, 11(8), 2248. doi: <https://doi.org/10.3390/pr11082248>
- Gultom, A. M. K., Yusa, N. M., & Wiadnyani, A. A. I. S. (2020). Pengaruh Jenis Pelarut Terhadap Aktivitas Antioksidan Ekstrak Daun Ubi Jalar Putih (*Ipomoea batatas* L) Menggunakan Metode Maserasi. *Jurnal Ilmu dan Teknologi Pangan (ITEPA)*, 9(4), 438-447. doi: <https://doi.org/10.24843/itepa.2020.v09.i04.p08>
- Guo, J., Cheng, J., North, B. J., & Wei, W. (2017). Functional Analyses of Major Cancer-Related Signaling Pathways in Alzheimer's Disease Etiology. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*, 1868(2), 341-358. doi: <https://doi.org/10.1016/j.bbcan.2017.07.001>
- Guo, Y., Ren, G., Zhang, K., Li, Z., Miao, Y., & Guo, H. (2021). Leaf Senescence: Progression, Regulation, and Application. *Molecular Horticulture*, 1, 1-25. doi: <https://doi.org/10.1186/s43897-021-00006-9>
- Gutbrod, K., Romer, J., & Dörmann, P. (2019). Phytol Metabolism in Plants. *Progress in lipid research*, 74, 1-17. doi: <https://doi.org/10.1016/j.plipres.2019.01.002>
- Gutbrod, P., Yang, W., Grujicic, G. V., Peisker, H., Gutbrod, K., Du, L. F., & Dörmann, P. (2021). Phytol Derived from Chlorophyll Hydrolysis in Plants is Metabolized Via Phytenal. *Journal of Biological Chemistry*, 296. doi: <https://doi.org/10.1016/j.jbc.2021.100530>
- Guzman, J. D. (2014). Natural Cinnamic Acids, Synthetic Derivatives and Hybrids With Antimicrobial Activity. *Molecules*, 19(12), 19292-19349. doi: [10.3390/molecules191219292](https://doi.org/10.3390/molecules191219292)
- Hakim, A. R. & Saputri, R. (2020). Narrative Review: Optimasi Etanol sebagai Pelarut Senyawa Flavonoid dan Fenolik: Narrative Review: Optimization of Ethanol as a Solvent for Flavonoids and Phenolic Compounds. *Jurnal Surya Medika (JSM)*, 6(1), 177-180.

- Handajani, F. (2019). Oksidan dan Antioksidan pada Beberapa Penyakit dan Proses Penuaan. Sidoarjo : Zifatama Jawa.
- Hapsoro, D. (2019). *Kultur In Vitro Tanaman Tebu dan Manfaatnya untuk Mutagenesis dengan Sinar Gamma*. Lampung : Cv. Anugrah Utama Raharja.
- Harahap, F. (2011). *Kultur Jaringan Tanaman*. Medan : Unimed Press
- Hartono, B., Chrisanto, C., & Farfar, I. O. (2019). Pengaruh Lama Penyimpanan terhadap Aktivitas Antioksidan Berbagai Macam Jus Buah Berdasarkan Metode DPPH. *Jurnal Kedokteran Meditek*, 25(2), 75-80. doi: <https://doi.org/10.36452/jkdoktmeditek.v25i2.1755>
- Hasanuddin, A. P., Yusran, Islawati, & Artati. (2023). Analisis Kadar Antioksidan Pada Ekstrak Daun Binahong Hijau *Anredera cordifolia* (Ten.) Steenis. *Bioma: Jurnal Biologi Makassar*, 8(2), 66-74.
- Hassanpour, S. H. & Doroudi, A. (2023). Review of the Antioxidant Potential of Flavonoids as a Subgroup of Polyphenols and Partial Substitute for Synthetic Antioxidants. *Avicenna journal of phytomedicine*, 13(4), 354.
- Hättenschwiler, S. & Vitousek, P. M. (2000). The Role of Polyphenols in Terrestrial Ecosystem Nutrient Cycling. *Trends in ecology & evolution*, 15(6), 238-243.
- He, M. & Ding, N. Z. (2020). Plant Unsaturated Fatty Acids: Multiple Roles in Stress Response. *Frontiers in plant science*, 11, 562785. doi: [10.3389/fpls.2020.562785](https://doi.org/10.3389/fpls.2020.562785)
- Hermanto. (2008). Aplikasi Alat HPTLC dan GC-MS. Jakarta: UI Press
- Hersila, N., Chatri, M., Vauzia, & Irdawati. (2023). Senyawa Metabolit Sekunder (Tanin) pada Tanaman sebagai Antifungi. *Jurnal Embrio*, 15(1), 16-22. doi: <https://doi.org/10.31317/embrio.v15i1.882>
- Heurteaux, C., Laigle, C., Blondeau, N., Jarretou, G., & Lazdunski, M. (2006). Alpha-Linolenic Acid and Riluzole Treatment Confer Cerebral Protection and Improve Survival after Focal Brain Ischemia. *Neuroscience*, 137(1), 241-251. doi: <https://doi.org/10.1016/j.neuroscience.2005.08.083>
- Honma, T. (2025). Discovery and Synthesis of Conjugated Fatty Acids from Natural Products. *Journal of Oleo Science*, 74(3), 241-249. doi: [10.5650/jos.ess24282](https://doi.org/10.5650/jos.ess24282)
- Horii, Y. & Kannan, K. (2008). Survey of Organosilicone Compounds, Including Cyclic and Linear Siloxanes, in Personal-Care and Household Products. *Archives of environmental contamination and toxicology*, 55, 701-710. doi: <https://doi.org/10.1007/s00244-008-9172-z>
- Hu, C., Zhao, H., Wang, W., Xu, M., Shi, J., Nie, X., & Yang, G. (2018). Identification of Conserved and Diverse Metabolic Shift of the Stylar, Intermediate and Peduncular Segments of Cucumber Fruit During Development. *International Journal of Molecular Sciences*, 19(1), 135. doi: [10.3390/ijms19010135](https://doi.org/10.3390/ijms19010135)

- Hussein, R. A. & El-Anssary, A. A. (2019). Plants Secondary Metabolites: The Key Drivers of the Pharmacological Actions of Medicinal Plants. *Herbal medicine*, 1(3), 11-30.
- Idu, M., Aihikhai, M. O., Imoni, C. A., Akokigho, C. E., & Olali, N. C. (2021). Gas Chromatography-Mass Spectrometry (GC-MS) Analysis and Phytochemical Screening of Polyherbal Aqueous Leaves Extract (PALE). *Journal of Complementary and Alternative Medical Research*, 14(2), 10-8.
- Ikhtimami, A. (2012). *Pengaruh Periode Subkultur Terhadap Kadar Saponin Akar Rambut Tanaman Gingseng Jawa (Talinum paniculatum Gaertn)*. (Skripsi). Universitas Airlangga, Surabaya.
- Imawati, M. F., Indriasari, C., & Azsrina, G. N. (2023). Studi Variasi Metode Pengeringan Terhadap Skrining Fitokimia Simplisia Krokot Magenta (*Portulaca grandiflora*). *Jurnal Mahasiswa Ilmu Kesehatan*, 1(3), 181-188. doi: <https://doi.org/10.59841/jumkes.v1i3>
- Indaryani, P. M. K., Ashari, S., & Mariana, B. D. (2019). Analisis Keragaman Genetik Tanaman Anggur (*Vitis vinifera* L.) Varietas BS 60 Hasil Perbanyakan Secara Kultur Jaringan Dengan Marka ISSR (Inter Simple Sequence Repeats). *Jurnal Produksi Tanaman*, 7(11), 2100-2106.
- Indrawanto, C., Purwono, S., Syakir, M., & Rumini, W. (2010). *Budidaya dan Pasca Panen Tebu*. Jakarta : ESKA media.
- Indriani, R., Prihastanti, E., Budihastuti, R., & Nurchayati, Y. (2020). Effect of Subculture Frequency Toward Growth and Carotenoid Content from Tomato (*Lycopersicon esculentum* Mill.) Callus. *Jurnal Biodjati*, 5(2), 303-315. doi: <https://doi.org/10.15575/biodjati.v5i2.5840>
- Indriani, S., Isdaryanti, M. A., Poleuleng, A. B., Syahra, N. J., & Prastiyo, Y. B. (2023). Analisis GC-MS (Gas Chromatography-Mass Spectrometry) terhadap Batang Kelapa Sawit (*Elaeis guineensis* Jacq.). *Jurnal Agropiantae*, 12(2), 147-155. doi: <https://doi.org/10.51978/agro.v12i2.527>
- Indriyah, S. N., Permatasari, D. A. I., & Pratama, K. J. (2023). Penetapan Kadar Fenolik Serta Uji Aktivitas Antioksidan Ekstrak dan Fraksi Batang Bajakah Kalalawit (*Uncaria Gambir* Roxb) Dengan Metode Frap. *Usada Nusantara: Jurnal Kesehatan Tradisional*, 1(2), 147-158. doi: <https://doi.org/10.47861/usd.v1i2>
- Ingrouille, M. & Eddie, B. (2006). *Plants: Diversity and Evolution*. Cambridge University Press.
- Iorizzo, M., Di Martino, C., Letizia, F., Crawford Jr, T. W., & Paventi, G. (2024). Production of Conjugated Linoleic Acid (CLA) by *Lactiplantibacillus plantarum*: A Review with Emphasis on Fermented Foods. *Foods*, 13(7), 975. doi: <https://doi.org/10.3390/foods13070975>

- Ironika, L. (2012). *Pengaruh Periode Subkultur Terhadap Kadar Saponin Akar Adventif Tanaman Gingseng Jawa (Talinum paniculatum Gaertn)*. (Skripsi). Universitas Airlangga, Surabaya.
- Ischebeck, T., Zbierzak, A. M., Kanwischer, M., & Dörmann, P. (2006). A Salvage Pathway for Phytol Metabolism in *Arabidopsis*. *Journal of Biological Chemistry*, 281(5), 2470-2477. doi: <https://doi.org/10.1074/jbc.M509222200>
- Islam, M. T., Ali, E. S., Uddin, S. J., Shaw, S., Islam, M. A., Ahmed, M. I., ... & Atanasov, A. G. (2018). Phytol: A Review of Biomedical Activities. *Food and chemical toxicology*, 121, 82-94. doi: <https://doi.org/10.1016/j.fct.2018.08.032>
- Islam, M. T., Rahman, M. A., Saeed, M., Ul-Haq, Z., Alam, M. J., Mondal, M., ... & Sharifi-Rad, J. (2020). Anti-Diarrheal Activities of Phytol Along with Its Possible Mechanism Of Action Through *In-Vivo* and *In-Silico Models*. *Cellular and Molecular Biology*, 66(4), 243-249. doi: <http://dx.doi.org/10.14715/cmb/2020.66.4.29>
- Javed, R., Yucesan, B., Zia, M., & Gurel, E. (2017). Differential Effects of Plant Growth Regulators on Physiology, Steviol Glycosides Content, and Antioxidant Capacity in Micropropagated Tissues of *Stevia rebaudiana*. *Biologia*, 72(10), 1156-1165. doi: [10.1515/biolog-2017-0133](https://doi.org/10.1515/biolog-2017-0133)
- Johannes, E., Litaay, M., & Syahribulan. (2016). The Bioactivity of Hexadecanoic Acid Compound Isolated from Hydroid *Aglaophenia cupressina* Lamoureux as Antibacterial Agent Against *Salmonella typhi*. *Int. J. Biol. Med. Res*, 7, 5469-5472.
- Julianto, T.S. (2019). *Fitokimia Tinjauan Metabolit Sekunder dan Skrining Fitokimia*. Malang : Penerbit Universitas Islam Indonesia.
- Junaidi, L. (2007). Antioksidan Alami: Sumber, Kimia, dan Teknologi Ekstraksi. *Warta Industri Hasil Pertanian*, 24(2), 52-69.
- Junairiah, J., Amalia, N. S., Manuhara, S. W., Matuzzahroh, N., & Sulistyorini, L. (2019). Pengaruh Variasi Zat Pengatur Tumbuh IAA, BAP, Kinetin Terhadap Metabolit Sekunder Kalus Sirih Hitam (*Piper betle* L. Var Nigra). *Jurnal Kimia Riset*, 4(2), 121-132.
- Kaban, S. M. P., Restiani, R., & Aditiyarini, D. (2024). Effect of Plant Growth Regulators (PGRs) on Biomass and Flavonoid Production of *Talinum paniculatum* Callus Culture. *Jurnal Biodjati*, 9(1), 11-25. doi: [10.15575/biodjati.v9i1.33788](https://doi.org/10.15575/biodjati.v9i1.33788)
- Kabera, J.N., Semana, E., Mussa, A.R., and He, X. (2014). Plant Secondary Metabolites: Biosynthesis, Classification, Function and Pharmacological Properties. *Journal of Pharmacy and Pharmacology*, 2, 377-392.
- Kadapi, M., Sunarso, C., Erizon, M. S., Maharani, N. D., Hakim, M. S., & Zahra, I. H. A. (2024). Teknologi Kultur *in vitro* untuk Meningkatkan Produksi Metabolit Sekunder pada Berbagai Tanaman Obat. *Jurnal Agrosains dan Teknologi*, 9(1), 18-29.

- Kanimozhi, S., Prithviraj, E., & Sumathy, G. (2021). Phytochemical and GC-MS Analysis of *Sphaeranthus amaranthoides* Burm. *Natural Volatiles & Essential Oils*, 8(5), 206-212.
- Kapitan, O.B., Kusumawardhani, D.T., & Nitti, F. (2023). Kandungan Senyawa Kimia Aktif Ekstrak Etanol dan Etil Asetat Daun Tembelekan (*Lantana camara*) asal pulau timor. *Seminar Nasioanal Politani Kupang Ke-6*. Kupang.
- Karthikeyan, J. & Samipillai, S. S. (2010). Sugarcane in Therapeutics. *Journal of Herbal Medicine and Toxicology*, 4(1), 9-14.
- Kartina, M. W. A. & Adiwena, M. (2019). Karakterisasi Kandungan Fitokimia Estrak Daun Karamunting (*Melastoma malabathricum* L.) menggunakan Metode Gas Chromatography Mass Spectrometry (GC-MS). *Biota: Jurnal Ilmiah Ilmu-Ilmu Hayati*, 16-23.
- Karunia, S. D., Supartono, M. A., & Sumarni, W. (2017). Analisis Sifat Antibakteri Ekstrak Biji Srikaya (*Annona squamosa* L.) dengan Pelarut Organik. *Indonesian Journal of Chemical Science*, 6(1), 56-60.
- Karyanti, K., Tajuddin, T., Khairiyah, H., Purwoko, D., Sukarnih, T., Rahmadara, G., ... & Saga, H. (2021). Proliferation of Oil Palm (*Elaeis Guineensis* Jacq.) Embryogenic Callus with Repeated Subcultures in Liquid Medium. *Jurnal Bioteknologi & Biosains Indonesia (JBBI)*, 8(1), 1-11. doi: [10.29122/jbbi.v8i1.4715](https://doi.org/10.29122/jbbi.v8i1.4715)
- Khafid, A., Wiraputra, M. D., Putra, A. C., Khoirunnisa, N., Putri, A. A. K., Suedy, S. W. A., & Nurchayati, Y. (2023). Uji Kualitatif Metabolit Sekunder pada Beberapa Tanaman yang Berkhasiat sebagai Obat Tradisional. *Buletin Anatomi Dan Fisiologi*, 8(1), 61-70. doi: <https://doi.org/10.14710/baf.8.1.2023.61-70>
- Khan, I. H. & Javaid, A. (2022). Hexane Soluble Bioactive Components of Leaf Extract of Quinoa. *Journal of Animal & Plant Sciences*, 32(2), 609-614. doi: <https://doi.org/10.36899/JAPS.2022.2.0461>
- Khan, I., Awan, S. A., Rizwan, M., Brestic, M., & Xie, W. (2023). Silicon: An Essential Element for Plant Nutrition and Phytohormones Signaling Mechanism Under Stresful Conditions. *Plant Growth Regulation*, 100(2), 301-319. doi: <https://doi.org/10.1007/s10725-022-00872-3>
- Klammer, H., Schlecht, C., Wuttke, W., Schmutzler, C., Gotthardt, I., Köhrle, J., & Jarry, H. (2007). Effects of a 5-day Treatment with the UV-filter octyl-methoxycinnamate (OMC) on the function of the Hypothalamo-Pituitary-Thyroid Function in Rats. *Toxicology*, 238(2-3), 192-199. doi: [10.1016/j.tox.2007.06.088](https://doi.org/10.1016/j.tox.2007.06.088)
- Ko, G. A. & Cho, S. K. (2018). Phytol Suppresses Melanogenesis Through Proteasomal Degradation of MITF via the ROS-ERK Signaling Pathway. *Chemico-biological interactions*, 286, 132-140. doi: <https://doi.org/10.1016/j.cbi.2018.02.033>
- Ko, G. A., Shrestha, S., & Kim Cho, S. (2018). *Sageretia thea* Fruit Extracts Rich in Methyl Linoleate and Methyl Linolenate Downregulate Melanogenesis Via

- The Akt/GSK3 β Signaling Pathway. *Nutrition research and practice*, 12(1), 3-12.
- Komunikasi Pribadi (Taslim, Pusat Laboratorium Forensik Badan Reserse Kriminal Kepolisian Negara Republik Indonesia, Sentul, Bogor, 2025)
- Konappa, N., Udayashankar, A. C., Krishnamurthy, S., Pradeep, C. K., Chowdappa, S., & Jogaiah, S. (2020). GC–MS Analysis of Phytoconstituents from *Amomum Nilgiricum* and Molecular Docking Interactions of Bioactive Serverogenin Acetate with Target Proteins. *Scientific reports*, 10(1), 16438. doi: <https://doi.org/10.1038/s41598-020-73442-0>
- Koo, I., Wei, X., & Zhang, X. (2014). Analysis of Metabolomic Profiling Data Acquired on GC–MS. In *Methods in Enzymology*, 543, 315-324. doi: <https://doi.org/10.1016/B978-0-12-801329-8.00016-7>
- Kumianjani, E., Damanik, R. I., & Siregar, L. A. M. (2015). Pengaruh Pemberian N 2, 4-D terhadap Pertumbuhan dan Metabolisme Kalus Kedelai pada Kondisi Hipoksida Secara In Vitro. *Jurnal Agroekoteknologi*, 4(1), 1673-1680.
- Kusuma, D. Y., Kristanti, A. N., Wibowo, A. T., Tan, B. C., & Manuhara, Y. S. W. (2023). Optimization of Biomass and Secondary Metabolite Production in *Gynura procumbens* (Lour.) Merr. Adventitious Roots Culture by Using the Method of Subculture and Fed-batch Cultivation in a Bioreactor. *HAYATI Journal of Biosciences*, 30(5), 797-807. doi: [10.4308/hjb.30.5.797-807](https://doi.org/10.4308/hjb.30.5.797-807)
- Kusumaningsih, T., Putra, A. A., & Aljunaid, M. (2021). Antibacterial Differences Effect Between Purple Leaves (*Graptophyllum pictum* (L) Griff.) 70% and 96% Ethanol Extract Against *Aggregatibacter actinomycetemcomittans* Bacteria. *Journal of International Dental and Medical Research*, 14(2), 519-524.
- Lathifah, U. (2020). *Identifikasi Senyawa Metabolit Sekunder dan Aktivitas Antioksidan Daun Tebu (Saccharum officinarum)*. (Skripsi). Universitas Negeri Semarang, Semarang.
- Lee, Y., Thompson, J. T., & Vanden Heuvel, J. P. (2009). 9E, 11E-conjugated Linoleic Acid Increases Expression of the Endogenous Antiinflammatory Factor, Interleukin-1 Receptor Antagonist, in RAW 264.7 Cells. *The Journal of nutrition*, 139(10), 1861-1866. doi: [10.3945/jn.108.100461](https://doi.org/10.3945/jn.108.100461)
- Letawe, Boone, & Piérard. (1998). Digital Image Analysis of the Effect of Topically Applied Linoleic Acid on Acne Microcomedones. *Clinical and experimental dermatology*, 23(2), 56-58. doi: <https://doi.org/10.1046/j.1365-2230.1998.00315.x>
- Leung, K. S., Oger, C., Guy, A., Bultel-Poncé, V., Vigor, C., Durand, T., ... & Lee, J. C. Y. (2022). Alpha-Linolenic Acid, Phytosteranes and Phytofurans in Plant, Algae and Food. *Advances in botanical research*, 101, 437-468. doi: <https://doi.org/10.1016/bs.abr.2021.09.005>
- Li, N., Xu, C., Li-Beisson, Y., & Philippar, K. (2016). Fatty Acid and Lipid Transport in Plant Cells. *Trends in plant science*, 21(2), 145-158.

- Li, P., Xu, G., Li, S. P., Wang, Y. T., Fan, T. P., Zhao, Q. S., & Zhang, Q. W. (2008). Optimizing Ultrapformance Liquid Chromatographic Analysis of 10 Diterpenoid Compounds in *Salvia Miltiorrhiza* Using Central Composite Design. *Journal of agricultural and food chemistry*, 56(4), 1164-1171. doi: <https://doi.org/10.1021/jf073020u>
- Lim, G. H., Singhal, R., Kachroo, A., & Kachroo, P. (2017). Fatty Acid–and Lipid-Mediated Signaling in Plant Defense. *Annual review of Phytopathology*, 55(1), 505-536.
- Liu, Y., Zhang, Y., Zhang, X., Xu, Q., Yang, X., & Xue, C. (2017). Medium-Chain Fatty Acids Reduce Serum Cholesterol by Regulating the Metabolism of Bile Acid in C57BL/6J Mice. *Food & function*, 8(1), 291-298. doi: [10.1039/C6FO01207H](https://doi.org/10.1039/C6FO01207H)
- Locatelli, C., Jardim, J. K. B., & Zancanaro, V. (2021). Role of Antioxidants in the Treatment of Hepatocellular Carcinoma: Integrative Review. *Research, Society and Development*, 10(1). doi: <https://doi.org/10.33448/rsd-v10i1.12028>
- Ma, K., Kou, J., Rahman, M. K. U., Du, W., Liang, X., Wu, F., ... & Pan, K. (2021). Palmitic Acid Mediated Change of Rhizosphere and Alleviation of Fusarium Wilt Disease in Watermelon. *Saudi Journal of Biological Sciences*, 28(6), 3616-3623. doi: <https://doi.org/10.1016/j.sjbs.2021.03.040>
- Mahawer, S. K., Karakoti, H., Kumar, R., Prakash, O., Kumar, S., Rawat, S., ... & Ren, Y. (2025). Assessment Of Antibacterial, Nematicidal and Herbicidal Activities of *Cautleya Spicata* (Sm.) Baker Extracts, With Chemical Profiling Using GC-MS. *Scientific reports*, 15(1), 7179. doi: <https://doi.org/10.1038/s41598-025-91661-1>
- Maji, S. R., Roy, C., & Sinha, S.K. (2023). Gas Chromatography–Mass Spectrometry (GC-MS): A Comprehensive Review of Synergistic Combinations and Their Applications in the Past Two Decades. *Journal of Analytical Sciences and Applied Biotechnology*, 5(2), 5-2.
- Malik, A., Ahmad, A. R., & Najib, A. (2017). Pengujian Aktivitas Antioksidan Ekstrak Terpurifikasi Daun Teh Hijau dan Jati Belanda. *Jurnal Fitofarmaka Indonesia*, 4(2), 238-240.
- Mamarimbing, M.S., Putra, I. G. N. A. D., & Setyawan, E. I. (2022). Aktivitas Antiinflamasi Ekstrak Etanol Tanaman Patah Tulang (*Euphorbia tirucalli* l.). Humantech: *Jurnal Ilmiah Multidisiplin Indonesia*, 2(3), 502-508.
- Marchev, A., Haas, C., Schulz, S., Georgiev, V., Steingroewer, J., Bley, T., & Pavlov, A. (2014). Sage In Vitro Cultures: A Promising Tool for the Production of Bioactive Terpenes and Phenolic Substances. *Biotechnology letters*, 36, 211-221. doi: <https://doi.org/10.1007/s10529-013-1350-z>
- Margareta, M. A. H. & Wonorahardjo, S. (2023). Optimasi Metode Penetapan Senyawa Eugenol dalam Minyak Cengkeh Menggunakan Gas Chromatography–Mass Spectrum dengan Variasi Suhu Injeksi. *Jurnal Sains*

- dan Edukasi Sains*, 6(2), 95-103. doi: <https://doi.org/10.24246/juses.v6i2p95-103>
- Martinez, R. A.S., Hernandez, J. M.P., Terol, G.L., Gallego-Jara, J., García-Marcos, L., Cánovas Díaz, M., & de Diego Puente, T. (2020). Data Preprocessing Workflow for Exhaled Breath Analysis by GC/MS Using Open Sources. *Scientific Reports*, 10(1), 22008. doi: <https://doi.org/10.1038/s41598-020-79014-6>
- Martins, J. L., Silva, D. M., Gomes, E. H., Fava, S. A., Carvalho, M. F., Macedo, I. Y., ... & Costa, E. A. (2022). Evaluation of Gastroprotective Activity of Linoleic Acid on Gastric Ulcer in A Mice Model. *Current Pharmaceutical Design*, 28(8), 655-660. doi: [10.2174/1381612826666200908144053](https://doi.org/10.2174/1381612826666200908144053)
- Maryam, F., Utami, Y. P., Mus, S., & Rohana, R. (2023). Perbandingan Beberapa Metode Ekstraksi Ekstrak Etanol Daun Sawo Duren (*Chrysophyllum cainito* L.) Terhadap Kadar Flavanoid Total Menggunakan Metode Spektrofotometri UV-VIS. *Jurnal Mandala Pharmacon Indonesia*, 9(1), 132-138. doi: [10.35311/jmpi.v9i1.336](https://doi.org/10.35311/jmpi.v9i1.336)
- Massoud, R., Jafari-Dastjerdeh, R., Naghavi, N., & Khosravi-Darani, K. (2022). All Aspects of Antioxidant Properties of Kombucha Drink. *Biointerface Res. Appl. Chem*, 12(3), 4018-4027.
- Mata, M. H. (2021). Akumulasi α Tokoferol pada Organ Tanaman dan Kultur Suspensi sel *Jatropha gossypifolia* Linn dari Nusa Tenggara Timur. *Jurnal Saintek Lahan Kering*, 4(1), 12-15. doi : <https://doi.org/10.32938/slk.v4i1.1411>
- Maulina, D. & Rochjana, A. U. H. (2023). Karakteristik Penggunaan Antioksidan Dibidang Dermatologi pada Pasien Rawat Jalan di Rumah Sakit X Periode Januari 2022-April 2023. *An-Najat*, 1(3), 213-237. doi: <https://doi.org/10.59841/an-najat.v1i3.168>
- Mazgaen, L. & Gurung, P. (2020). Recent Advances in Lipopolysaccharide Recognition Systems. *International journal of molecular sciences*, 21(2), 379. doi: [10.3390/ijms21020379](https://doi.org/10.3390/ijms21020379)
- Melani, A., Atikah, A., Robiah, R., & Khasanah, N. (2022). Kajian Pengaruh Variasi Pelarut, Kecepatan Pengadukan Dan Waktu Pada Proses Ekstraksi Kalium Dari Abu Kulit Buah Semangka (*Citrullus lanatus*). *Jurnal Distilasi*, 7(2), 29-36.
- Melariri, P., Campbell, W., Etusim, P., & Smith, P. (2012). In Vitro and In Vivo Antimalarial Activity of Linolenic and Linoleic Acids and their Methyl Esters. *Adv Stud Biol*, 4(7), 333-49.
- Mentari, D., Naima, M., Wulansari, R., Widada, J., Nuringtyas, T. R., Wibawa, T., & Wijayanti, N. (2019). Pengaruh Perbedaan Metode Ekstraksi Metabolit Sekunder *Streptomyces* sp. GMR22 terhadap Toksisitas pada Sel BHK-21. *Pharmacon: Jurnal Farmasi Indonesia*, 16(1), 1-10.

- Minarsih, H., Suharyo., Riyadi, I., & Ratnadewi, D. (2016). Pengaruh Jumlah Subkultur dan Media Sub-Optimal terhadap Pertumbuhan dan Kemampuan Regenerasi Kalus Tebu (*Saccharum officinarum* L.). *Menara Perkebunan*, 84(1), 28-40. doi : <http://dx.doi.org/10.22302/iribb.jur.mp.v84i1.219>
- Mishra, K., Ojha, H., & Chaudhury, N. K. (2012). Estimation of Antiradical Properties of Antioxidants Using DPPH Assay: A Critical Review and Results. *Food chemistry*, 130(4), 1036-1043. doi: [10.1016/j.foodchem.2011.07.127](https://doi.org/10.1016/j.foodchem.2011.07.127)
- Moglad, E. H., Alnoor, A. A., Eltayeb, N. M., Abdalkareem, E. A., Ali, A., Oraiby, M. E., ... & Abdalla, A. N. (2024). Therapeutic Potential of *Acanthospermum hispidum*: A Comprehensive Analysis of Its Antimicrobial, Antioxidant, and Anticancer Properties. *Journal of Spectroscopy*, 2024(1), 8733990. doi: <https://doi.org/10.1155/2024/8733990>
- Molyneux, P. (2004). The Use of the Stable Free Radical Diphenylpicrylhydrazyl (DPPH) for Estimating Antioxidant Activity. *Songklanakarin J. sci. technol*, 26(2), 211-219.
- Monteiro, M. S. S. B., Ozetti, R. O., Vergnanini, A. L., Brito-Gitirana, L., Volpato, N. M., Freitas, Z. M. F., Junior, E. R., & Santos, E. P. (2012). Evaluation of Octyl P-Methoxycinnamate Included in Liposomes and Cyclodextrins in Anti-Solar Preparations: Preparations, Characterizations And In Vitro Penetration Studies. *International journal of nanomedicine*, 7, 3045-3058. doi: <https://doi.org/10.2147/IJN.S28550>
- Mukhriani, T. (2014). Ekstraksi, Pemisahan Senyawa, dan Identifikasi Senyawa Aktif. *Jurnal Kesehatan*, 7(2).
- Mulyani, Y., Malonga, W. A. M., & Sandra, E. (2024). Teknik Subkultur dalam Kultur Jaringan Tanaman Anggrek Ki Aksara (*Macodes petola*) secara In Vitro. *Jurnal Satwa Tumbuhan Indonesia*, 1(1), 15-23.
- Muniyandi, K., George, E., Sathyanarayanan, S., George, B. P., Abrahamse, H., Thamburaj, S., & Thangaraj, P. (2019). Phenolics, Tannins, Flavonoids and Anthocyanins Contents Influenced Antioxidant And Anticancer Activities of Rubus Fruits from Western Ghats, India. *Food Science and Human Wellness*, 8(1), 73-81. doi: <https://doi.org/10.1016/j.fshw.2019.03.005>
- Murch, S. J. & Saxena, P. K. (2006). A Melatonin-Rich Germplasm Line of St John's wort (*Hypericum perforatum* L.). *Journal of pineal research*, 41(3), 284-287. doi: <https://doi.org/10.1111/j.1600-079X.2006.00367.x>
- Mutiara, E. V., Wildan, A., Advistasari, Y. D., & Indriyanti, E. (2022). Sintesis oktil sinamat dengan Metode Sonokimia dan Potensinya Sebagai Antikolesterol. *Jurnal Farmasi (Journal of Pharmacy)*, 11(1), 7-12.
- Nafiisah, A., Purnamasari, R., & Mudalianah, S. (2024). Identifikasi Senyawa Metabolit Sekunder pada Ekstrak Etanol Daun Binahong. *Jurnal sosial dan sains*, 4(11), 1093-1106.

- Nanlohy, F. N., Yalindua, A., & Kamagi, D. D. (2023). *Kultur Jaringan Tanaman*. Sleman : Bintang Semesta Media.
- Nasiruddin, Chen, G., Li, X., Minghui, J., Masood, T., Safir, W., ... & Zeb, S. (2022). Comparison of Phytochemical Constituents and Pharmacological Activities of Various Solvent Extracts Obtained from *Millettia Speciosa* Stem Powder. *BioMed Research International*, 2022(1), 2486979. doi: <https://doi.org/10.1155/2022/2486979>
- National Cancer Institute. (2018). Angiogenesis Inhibitors. [Online]. Diakses dari <https://pmc.ncbi.nlm.nih.gov/articles/PMC7190605/>.
- Ningsih, I. Y. (2014). Pengaruh Elisitor Biotik dan Abiotik pada Produksi Flavonoid Melalui Kultur Jaringan Tanaman. *PHARMACY: Jurnal Farmasi Indonesia (Pharmaceutical Journal of Indonesia)*, 11(2).
- Nordström, D. C. E., Friman, C., Konttinen, Y. T., Honkanen, V. E. A., Nasu, Y., & Antila, E. (1995). Alpha-Linolenic Acid in the Treatment of Rheumatoid Arthritis. A Double-Blind, Placebo-Controlled and Randomized Study: Flaxseed Vs. Safflower Seed. *Rheumatology international*, 14, 231-234.
- Nugroho, L. H., Sumardi, I., Wisnu, M., & Anggraeny, R. N. (2007). Distribusi dan Profil Kromatogram Minyak Atsiri pada Temulawak (*Curcuma xanthorrhiza* Roxb.) yang Ditumbuhkan Secara In Vitro Dan In Vivo. *Berkala Ilmiah Biologi*, 6(2), 87-95.
- Nuraini, S. (2022). Karakterisasi Perkecambahan Stek Satu Mata Tunas (*Bud Chips*) Tebu (*Saccharum officinarum* L.) Varietas Komersial Pada Kondisi Cekaman Kelebihan Air. (Skripsi). Universitas Lampung, Bandar Lampung.
- Nuriah, S., Putri, M. D., Rahayu, S., Advaita, C. V., Nurfadhila, L., & Utami, M. R. (2023). Analisis Kualitatif Senyawa Parasetamol Pada Sampel Biologis Menggunakan Metode *Gas Chromatography-Mass Spectrometry* (GC-MS). *Journal of Pharmaceutical and Sciences*, 795-803.
- Nury, T., Lizard, G., & Vejux, A. (2020). Lipids Nutrients in Parkinson and Alzheimer's diseases: Cell death and cytoprotection. *International Journal of Molecular Sciences*, 21(7), 2501. doi: <https://doi.org/10.3390/ijms21072501>
- Orcan, P. & Orcan, M. Y. (2024). Insights into Total Phenolic, Flavonoid, and Antioxidant Activity of Callus Subculture Frequency in Rare Endemic *Ajuga xylorrhiza* Kit Tan. *Scientific Reports*, 14(1), 31720. doi: <https://doi.org/10.1038/s41598-024-82824-7>
- Ozyigit, I. I., Dogan, I., Hocaoglu-Ozyigit, A., Yalcin, B., Erdogan, A., Yalcin, I. E., Cabi, E. & Kaya, Y. (2023). Production of Secondary Metabolites Using Tissue Culture-Based Biotechnological Applications. *Frontiers in Plant Science*, 14, 1132555. doi: [10.3389/fpls.2023.1132555](https://doi.org/10.3389/fpls.2023.1132555)
- Pandey, A. & Tripathi, S. (2014). Concept of Standardization, Extraction and Pre Phytochemical Screening Strategies for Herbal Drug. *Journal of Pharmacognosy and phytochemistry*, 2(5), 115-119.

- Papaccio, F., Caputo, S., & Bellei, B. (2022). Focus on the Contribution of Oxidative Stress In Skin Aging. *Antioxidants*, 11(6), 1121. doi: <https://doi.org/10.3390/antiox11061121>
- Pappelbaum, K. I., Virgilio, N., Epping, L., van der Steen, B., Jimenez, F., Funk, W., ... & Bertolini, M. (2024). Revealing Novel Insights on How Oral Supplementation with Collagen Peptides May Prevent Hair Loss: Lessons from The Human Hair Follicle Organ Culture. *Journal of Functional Foods*, 116, 106124. doi: <https://doi.org/10.1016/j.jff.2024.106124>
- Parawansah, N. I. & Qodri, U. L. (2023). Uji Aktivitas Antioksidan Ekstrak Tebu Merah dan Tebu Hijau (*Saccharum officinarum* L.) menggunakan Metode DPPH (1, 1-Difenil-2-Pikrilhidrazil). *Jurnal Farmasi Tinctura*, 4(2), 63-71.
- Pathak, V. & Tiwari, V. K. (2017). Phytochemical Screening of *Saccharum officinarum* (Linn.) Stem. *International Journal of Innovative Science and Research Technology*, 2(8), 291-305.
- Pauloi, L., Salim, R. M., Chiang, L. K., Sundang, M., Rusdi, N. A., Maid, M., ... & Moktar, J. (2024). GC-MS Analysis of Bioactive Compounds in Ethanolic Extract on Different Parts of *Ziziphus mauritiana*. *Biodiversitas Journal of Biological Diversity*, 25(10). doi: [10.13057/biodiv/d251006](https://doi.org/10.13057/biodiv/d251006)
- Peng, X., Zhang, T. T., & Zhang, J. (2015). Effect of Subculture Times on Genetic Fidelity, Endogenous Hormone Level and Pharmaceutical Potential of *Tetrastigma hemsleyanum* Callus. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 122, 67-77. doi: [10.1007/s11240-015-0750-2](https://doi.org/10.1007/s11240-015-0750-2)
- Perangin-Angin, Y., Purwaningrum, Y., Asbur, Y., Rahayu, M. S., & Nurhayati, N. (2019). Pemanfaatan Kandungan Metabolit Sekunder yang dihasilkan Tanaman pada Cekaman Biotik. *Agriland: Jurnal Ilmu Pertanian*, 7(1), 39-47.
- Pereira, D. M., Correia-da-Silva, G., Valentão, P., Teixeira, N., & Andrade, P. B. (2013). Palmitic Acid and Ergosta-7, 22-dien-3-ol Contribute to the Apoptotic Effect and Cell Cycle Arrest of an Extract From *Marthasterias glacialis* L. in Neuroblastoma Cells. *Marine drugs*, 12(1), 54-68. doi: [10.3390/md12010054](https://doi.org/10.3390/md12010054)
- Perez-Gonzalez, C., Serrano Vega, R., González-Chávez, M., Zavala Sanchez, M. A., & Pérez Gutiérrez, S. (2013). Anti-Inflammatory Activity and Composition of *Senecio salignus* Kunth. *BioMed research international*, 2013(1), 814693. doi: <http://dx.doi.org/10.1155/2013/814693>
- Pham, T. N., Lam, T. D., Nguyen, M. T., Le, X. T., Vo, D. V. N., Toan, T. Q., & Vo, T. S. (2019). Effect of Various Factors on Extraction Efficiency of Total Anthocyanins From Butterfly Pea (*Clitoria ternatea* L. Flowers) in Southern Vietnam. In *IOP Conference Series: Materials Science and Engineering*, 544(1), 012013. doi: [10.1088/1757-899X/544/1/012013](https://doi.org/10.1088/1757-899X/544/1/012013)
- Phillips, G. C., & Garda, M. (2019). Plant Tissue Culture Media and Practices: An Overview. In *In Vitro Cellular & Developmental Biology-Plant*, 55, 242-257. doi: <https://doi.org/10.1007/s11627-019-09983-5>

- Piechocka, J., Wieczorek, M., & Głowacki, R. (2020). Gas Chromatography–Mass Spectrometry Based Approach for the Determination of Methionine-Related Sulfur-Containing Compounds in Human Saliva. *International Journal of Molecular Sciences*, 21(23), 9252. doi: [10.3390/ijms21239252](https://doi.org/10.3390/ijms21239252)
- Pono, P., Restiani, R., & Adityarini, D. (2021). Elisitasi Saponin dalam Kultur Kalus Ginseng Jawa (*Talinum paniculatum Gaertn.*) menggunakan Asam Salisilat. *Sciscitatio*, 2(2), 45-53.
- Potenza, L., Minutelli, M., Stocchi, V., & Fraternale, D. (2020). Biological Potential of an Ethanolic Extract from “*Mela Rosa Marchigiana*” Pulp Callus Culture. *Journal of Functional Foods*, 75, 104269. doi: <https://doi.org/10.1016/j.jff.2020.104269>
- Pourebadi, N., Motafakkerzadeh, R., Kosari-Nasab, M., Farsad Akhtar, N., & Movafeghi, A. (2015). The influence of TDZ Concentrations On In Vitro Growth and Production of Secondary Metabolites by The Shoot and Callus Culture of *Lallemantia iberica*. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 122, 331-339. doi: <https://doi.org/10.1007/s11240-015-0769-4>
- Prasetyorini. (2019). *Buku Ajar Kultur Jaringan*. Bogor : Lembaga Penelitian dan Pengabdian Masyarakat Universitas Pakuan.
- Pretorius, C. J., Zeiss, D. R., & Dubery, I. A. (2021). The Presence of Oxygenated Lipids in Plant Defense in Response to Biotic Stress: A Metabolomics Appraisal. *Plant Signaling & Behavior*, 16(12), 1989215. doi: [10.1080/15592324.2021.1989215](https://doi.org/10.1080/15592324.2021.1989215)
- Prihantini, A. I., Sukito, A., & Tachibana, S. (2018). Production of Antioxidant Compounds from Tissue Culture of *Artemisia annua*. *Nusantara bioscience*, 10(4), 251-255. doi: [10.13057/nusbiosci/n100409](https://doi.org/10.13057/nusbiosci/n100409)
- Purushothaman, R., Vishnuram, G., & Ramanathan, T. (2025). Antiinflammatory Efficacy of n-hexadecanoic Acid from a Mangrove Plant *Excoecaria Agallocha* L. Through In Silico, In Vitro and In Vivo. *Pharmacological Research-Natural Products*, 7, 100203. doi: <https://doi.org/10.1016/j.prenap.2025.100203>
- Purushothaman, R., Vishnuram, M. G., & Ramanathan, D. T. (2024). Isolation and Identification of N-Hexadecanoic Acid from *Excoecaria Agallocha* L. and its Antibacterial and Antioxidant Activity. *Journal of emerging technologies and innovative research*, 11(1), 332-342.
- Purwakusumah, E. D., Royani, L., & Rafi, M. (2016). Evaluasi Aktivitas Antioksidan dan Perubahan Metabolit Sekunder Mayor Temulawak (*Curcuma xanthorrhiza*) pada Umur Rimpang Yang Berbeda. *Jurnal Jamu Indonesia*, 1(1), 10-17.
- Purwaningrum, Y. (2013). Kultur Kalus sebagai Penghasil Metabolit Sekunder Berupa Pigmen. *Agriland*, 2(2), 117-127.
- Purwanti, L., Dasuki, U. A., & Imawan, A. R. (2019). Perbandingan Aktivitas Antioksidan dari Seduhan 3 Merk Teh Hitam (*Camellia sinensis* (L.) kuntze)

- dengan Metode Seduhan berdasarkan SNI 01-1902-1995. *Jurnal Ilmiah Farmasi Farmasyifa*, 2(1), 19-25.
- Putra, I. G. A., Wrasianti, L. P., Ayu, D., & Yuarini, A. (2022). Identifikasi Senyawa Penyusun Minuman Herbal Serai-Gula Lontar Menggunakan Gas Chromatography-Mass Spectrometry Lemon Grass-Palm Sugar Herbal Drink Compounds Identification Using Gas Chromatography-Mass Spectrometry. *Jurnal Ilmu Dan Teknologi Pangan*, 11 (4), 593-600.
- Qidwai, A., Pandey, M., Pathak, S., Kumar, R., & Dikshit, A. (2017). The Emerging Principles for Acne Biogenesis: A Dermatological Problem of Puberty. *Human Microbiome Journal*, 4, 7-13. doi: <https://doi.org/10.1016/j.humic.2017.05.001>
- Raafat, K., El-Darra, N., & Saleh, F. A. (2020). Gastroprotective and Anti-Inflammatory Effects of *Prunus Cerasus* Phytochemicals and Their Possible Mechanisms of Action. *Journal of Traditional and Complementary Medicine*, 10(4), 345-353. doi: <https://doi.org/10.1016/j.jtcme.2019.06.001>
- Rao, A. S. (2015). Callus Induction and Organogenesis in Sugarcane (*Saccharum officinarum* L.) var 93v297. *International Letters of Natural Sciences*, (48). doi: [10.18052/www.scipress.com/ILNS.48.14](https://doi.org/10.18052/www.scipress.com/ILNS.48.14)
- Rao, D. A., Yerramneedi, R.B., & Padal, S.B. (2024). Phytochemical Screening, In Vitro Antimicrobial Potentiality, and GC-MS Activity of *Woodfordia fruticosa* L. Flowers. *High Technology Letters*, 30(6).
- Rao, T. R., Harshitha, S., & Lahari, K. L. (2023). Gas Chromatography-Mass Spectroscopy: An Overview. *European Journal of Biomedical*, 10(5), 83-89.
- Rarison, R. H., Truong, V. L., Yoon, B. H., Park, J. W., & Jeong, W. S. (2023). Antioxidant and Anti-Inflammatory Mechanisms of Lipophilic Fractions from *Polyscias Fruticosa* Leaves Based on Network Pharmacology, In Silico, And In Vitro Approaches. *Foods*, 12(19), 3643. doi: <https://doi.org/10.3390/foods12193643>
- Rasyid, A., Putra, M. Y. & Yasman (2023). Antibacterial and Antioxidant Activity of Sea Cucumber Extracts Collected from Lampung Waters, Indonesia. *Kuwait Journal of Science*, 50(4), 615-621. doi: <https://doi.org/10.1016/j.kjs.2023.03.012>
- Razack, S., Hemanth Kumar, K., Nallamuthu, I., Naika, M., & Khanum, F. (2015). Antioxidant, Biomolecule Oxidation Protective Activities of *Nardostachys jatamansi* DC And Its Phytochemical Analysis by RP-HPLC and GC-MS. *Antioxidants*, 4(1), 185-203. doi: [10.3390/antiox4010185](https://doi.org/10.3390/antiox4010185)
- Rivai, H., Nurdin, H., Suyani, H., & Bakhtiar, A. (2010). Pengaruh Cara Pengeringan Terhadap Perolehan Ekstraktif, Kadar Senyawa Fenolat dan Aktivitas Antioksidan dari Daun Dewa (*Gynura pseudochina* (L.) DC.). *Majalah Obat Tradisional*, 15(1), 26-33.
- Riwanti, P., Izazih, F., & Amaliyah, A. (2020). Pengaruh Perbedaan Konsentrasi Etanol pada Kadar Flavonoid Total Ekstrak Etanol 50, 70 dan 96% *Sargassum*

- polycystum* dari Madura. *Journal of Pharmaceutical Care Anwar Medika (J-PhAM)*, 2(2) : 82-95. doi: <http://dx.doi.org/10.36932/jpcam.v2i2.1>
- Roessner, U. & Beckles, D. M. (2009). Metabolite Measurements. *Plant Metabolic Networks*, 39-69. doi: https://doi.org/10.1007/978-0-387-78745-9_3
- Roeswitawati, D., Hussain, Z., Jan, A., Zekker, I., Mel, M., Setyobudi, R. H., ... & Hudin, D. (2022). The Evaluation of Secondary Metabolites in *Saccharum officinarum* L. and *Mimosa invisa* Mart. as Natural Herbicides. *Jordan Journal of Biological Sciences*, 15(1).1-6. doi: <https://doi.org/10.54319/jjbs/150101>
- Rojsanga, P., Bunsupa, S., Brantner, A. H., & Sithisarn, P. (2017). Comparative Phytochemical Profiling and *In Vitro* Antioxidant Activity of Extracts from Raw Materials, Tissue-Cultured Plants, and Callus of *Oroxylum indicum* (L.) Vent. *Evidence-Based Complementary and Alternative Medicine*, 2017(1), 6853212. doi: <https://doi.org/10.1155/2017/6853212>
- Romer, J., Gutbrod, K., Schuppener, A., Melzer, M., Müller-Schüssele, S. J., Meyer, A. J., & Dörmann, P. (2024). Tocopherol and Phylloquinone Biosynthesis in Chloroplasts Requires the Phytol Kinase VITAMIN E PATHWAY GENE5 (VTE5) and the Farnesol Kinase (FOLK). *The Plant Cell*, 36(4), 1140-1158. doi: <https://doi.org/10.1093/plcell/koad316>
- Rosa, G. P., Seca, A. M., Pinto, D. C., & Barreto, M. C. (2024). New Phytol Derivatives with Increased Cosmeceutical Potential. *Molecules*, 29(20), 4917. doi: <https://doi.org/10.3390/molecules29204917>
- Rouvier, F., Abou, L., Wafo, E., & Brunel, J. M. (2025). Linoleic Fatty Acid from Rwandan Propolis: A Potential Antimicrobial Agent Against *Cutibacterium acnes*. *Current Issues in Molecular Biology*, 47(3), 162. doi: <https://doi.org/10.3390/cimb47030162>
- Rudin, N. A. (2020). Pengaruh Cekaman abiotik Terhadap Ekspresi Gen dan Konsentrasi Metabolit Sekunder pada *Catharanthus roseus*. *Jurnal Pro-Life*, 7(3), 262-274.
- Rusandi & Rusli, M. (2021). Merancang Penelitian Kualitatif Dasar/Deskriptif dan Studi Kasus. *Al-Ubudiyah: Jurnal Pendidikan Dan Studi Islam*, 2(1), 48-60. doi: [10.55623/au.v2i1.18](https://doi.org/10.55623/au.v2i1.18)
- Sakakibara, H., Takei, K., & Hirose, N. (2006). Interactions between Nitrogen and Cytokinin in the Regulation of Metabolism and Development. *Trends in plant science*, 11(9), 440-448. doi: <https://doi.org/10.1016/j.tplants.2006.07.004>
- Salam, U., Ullah, S., Tang, Z. H., Elateeq, A. A., Khan, Y., Khan, J., Khan, A. & Ali, S. (2023). Plant Metabolomics: An Overview of the Role of Primary and Secondary Metabolites against Different Environmental Stres Factors. *Life*, 13(3), 706. doi: <https://doi.org/10.3390/life13030706>
- Salamah, N. & Widyasari, E. (2015). Aktivitas Antioksidan Ekstrak Metanol Daun Kelengkeng (*Euphoria longan* (L) Steud.) dengan metode Penangkapan

- Radikal 2,2'-difenil-1-pikrilhidrazil. *Pharmaciana*, 5(1), 25-34. doi: <https://doi.org/10.12928/pharmaciana.v5i1.2283>
- Salamah, N., Rozak, M., & Al Abror, M. (2017). Pengaruh Metode Penyarian terhadap Kadar Alkaloid Total Daun Jembirit (*Tabernaemontana sphaerocarpa*) dengan metode spektrofotometri visibel. *Pharmaciana*, 7(1), 113-122. doi: <https://doi.org/10.12928/pharmaciana.v7i1.6330>
- Samadd, M. A., Hossain, M. R., Taher, M. A., Hasan, M. M., Haque, S., Rafid, M. I., ... & Khan, M. (2024). Multifaceted Chemico-Pharmacological Insights into *Cynometra ramiflora* L.: Unveiling Its GC-MS, Cytotoxic, Thrombolytic, Anti-Inflammatory, Antioxidant, Anti-Diarrheal, Hypoglycemic, and Analgesic Potentials. *Natural Product Communications*, 19(5), 1934578X241257377.
- Sánchez-Ramos, M., Bahena, S. M., Romero-Estrada, A., Bernabé-Antonio, A., Cruz-Sosa, F., Gonzáles-Christen, J., ... & Alvarez, L. (2018). Establishment and Phytochemical Analysis of a Callus Culture from *Ageratina pichinchensis* (Asteraceae) and Its Anti-Inflammatory Activity. *Molecules*, 23(6), 1258. doi: <https://doi.org/10.3390/molecules23061258>
- Satria, A. W., Darmawan, A., & Sudarmanto, I. (2020). Analisis Senyawa Aktif dalam Cacing Spesies Lokal dan Efek Antiinflamasi. *BioWallacea : Jurnal Penelitian Biologi (Journal of Biological Research)*, 7(1), 1070-1077.
- Sayuti, K. & Yenrina, R. (2015). Antioksidan Alami dan Sintetik. Sumatera : Andalas University Press.
- Scalise, A., Bianchi, A., Tartaglione, C., Bolletta, E., Pierangeli, M., Torresetti, M., Marazzi, M., & Benedetto, G. D. (2015). Microenvironment and Microbiology of Skin Wounds: The Role of Bacterial Biofilms and Related Factors, *Seminars in Vascular Surgery*, 28(3-4), 151-159.
- Setiawati, T., Ayalla, A., Nurzaman, M., Kusumaningtyas, V., & Bari, I. (2020). Analisis Metabolit Sekunder Kultur Pucuk, Kalus, dan Tanaman Lapang *Chrysanthemum morifolium* Ramat. *Jurnal Ilmu Dasar*, 21(1), 1-10.
- Setyorini, S. D. & Yusnawan, E. (2016). Peningkatan Kandungan Metabolit Sekunder Tanaman Aneka Kacang Sebagai Respon Cekaman Biotik. *Iptek Tanaman Pangan*, 11(2).
- Sharafzadeh, S. & Zare, M. (2011). Influence of Growth Regulators on Growth and Secondary Metabolites of Some Medicinal Plants from Lamiaceae Family. *Adv. Environ. Biol*, 5(8), 2296-2302.
- Sharma, P. (2011). Cinnamic acid derivatives: A New Chapter of Various Pharmacological Activities. *Journal of Chemical and Pharmaceutical Research*, 3(2), 403.
- Shelke, V. & Bhot, M. (2019). GC-MS Analysis of Bio-active Compounds in Ethanolic Extract of Leaf and Stem of *Asclepias curassavica* L. *International Journal of Pharmaceutical Investigation*, 9(2). doi: [10.5330/ijpi.2019.2.13](https://doi.org/10.5330/ijpi.2019.2.13)

- Shofiyani, A. & Purnawanto, A. M. (2017). Pertumbuhan Kalus Kencur (*Kaemferia Galanga* L.) Pada Komposisi Media Dengan Perlakuan Sukrosa Dan Zat Pengatur Tumbuh (2, 4 D dan Benzil Aminopurin). *Agritech: Jurnal Fakultas Pertanian Universitas Muhammadiyah Purwokerto*, 19(1), 55-64.
- Shrivastava, V., Sharma, N., Shrivastava, V. & Dwidevi, K. (2023) In-Vitro Assessment of Antioxidant and Antimicrobial Activity of Field Grown and Tissue Cultured Fruit Callus of *Nothapodytes nimmoniana*. *Biomedical & Pharmacology Journal*, 16(4), 2099-2112. doi: <https://dx.doi.org/10.13005/bpj/2785>
- Singh, A., Lal, U. R., Mukhtar, H. M., Singh, P. S., Shah, G., & Dhawan, R. K. (2015). Phytochemical Profile of Sugarcane and Its Potential Health Aspects. *Pharmacognosy reviews*, 9(17), 45-54. doi: [10.4103/0973-7847.156340](https://doi.org/10.4103/0973-7847.156340)
- Sirivibulkovit, K., Nouanthavong, S., & Sameenoi, Y. (2018). Paper-based DPPH Assay for Antioxidant Activity Analysis. *Analytical Sciences*, 34, 795.
- Situmeang, P. H., Barus, A., & Irsal, I. (2015). Pengaruh Konsentrasi Zat Pengatur Tumbuh dan Sumber Bud Chips Terhadap Pertumbuhan Bibit Tebu (*Saccharum officinarum*) di Potray. *Jurnal Agroekoteknologi Universitas Sumatera Utara*, 3(3), 992-1004. doi: [10.32734/jaet.v3i3.10944](https://doi.org/10.32734/jaet.v3i3.10944)
- Smetanska, I. (2018). Sustainable Production of Polyphenols and Antioxidants by Plant In Vitro Cultures. *Bioprocessing of plant in vitro systems*, 225-269. doi: https://doi.org/10.1007/978-3-319-32004-5_2-1
- Soleh, M. A., Ariyanti, M., Dewi, I. R., & Kadapi, M. (2018). Chlorophyll Fluorescence and Stomatal Conductance of Ten Sugarcane Varieties Under Waterlogging and Fluctuation Light Intensity. *Emirates Journal of Food and Agriculture*, 30(11), 935-940. doi: [10.9755/ejfa.2018.v30.i11.1844](https://doi.org/10.9755/ejfa.2018.v30.i11.1844)
- Soleh, M. A., Rosniawaty, S., & Sofiani, E. F. (2019). Respons Pertumbuhan dan Fisiologi Beberapa Varietas Tebu (*Saccharum officinarum* L.) Asal Kultur Jaringan Yang Diberi Cekaman Genangan Air. *Jurnal Agrikultura*, 30(3), 117-124. doi: <https://doi.org/10.24198/agrikultura.v30i3.24976>
- Sukmadjaja, D., Supriyati, Y., & Pardal, S. J. (2014). Kultur Apeks untuk Penyediaan Bibit Unggul Tebu Varietas PS864 dan PS881. *Jurnal AgroBiogen*, 10(2), 45-52.
- Sukweenadhi, J., Setiawan, F., Yunita, O., Kartini, K., & Avanti, C. (2020). Antioxidant Activity Screening of Seven Indonesian Herbal Extract. *Biodiversitas*, 21(5), 2062-2067. doi: <https://doi.org/10.13057/biodiv/d210532>
- Suminar, E., Sumadi, S., Mubarak, S., Sunarto, T., & Rini, N. S. E. (2017). Percepatan Penyediaan Benih Sumber Kedelai Unggul Secara *In Vitro*. *Agrikultura*, 28(3).
- Supriyadi, S., Diana, N. E., & Djumali, D. (2018). Pertumbuhan Dan Produksi Tebu (*Saccharum Officinarum*; Poaceae) Pada Berbagai Paket Pemupukan di

- Lahan Kering Berpasir. *Berita Biologi*, 17(2), 147-156. doi: [10.14203/beritabiologi.v17i2.2287](https://doi.org/10.14203/beritabiologi.v17i2.2287)
- Surajo, N., Bem, A. A., & Musa, D. D. (2024). Phytochemical Analysis and Antifungal Activity of Leaves Extract of *Vachellia seyal* on *Phytophthora infestans* Isolated from rotten potatoes. *UMYU Journal of Microbiology Research*, 9(3), 79-88. doi: <https://doi.org/10.47430/ujmr.2493.011>
- Suryanto, E. (2018). Kimia Antioksidan. Bandung :Patra Media Gravindo.
- Sutha, S., Devi, K., & Mohan, V. R. (2011). GC-MS Determination of Bioactive Components of *Erythralum scandens* Bl., *Bijdr. Journal of Applied Pharmaceutical Science*, 1(09) : 170-173.
- Syabrina, A., Anjani, D., & Sarjani, T. M. (2023). Identification of Stomata Types in Plants of The Genus *Saccharum*. *Jurnal Biologi Tropis*, 23(2), 485-494. doi: <https://dx.doi.org/10.29303/jbt.v23i2.6315>
- Syamsul, E. S., Amanda, N. A., & Lestari, D. (2020). Perbandingan Ekstrak Lamur *Aquilaria malaccensis* dengan Metode Maserasi dan Refluks. *Jurnal Riset Kefarmasian Indonesia*, 2(2), 97-104.
- Taiz, L. & Zeiger, E. (2005). *Plant Physiology*, 3rd ed. Sinauer Association Inc.: Sunderland, CA, USA.
- Takada, K. (2023). Synthesis of Biobased Functional Materials Using Photoactive Cinnamate Derivatives. *Polymer Journal*, 55(10), 1023-1033. doi: <https://doi.org/10.1038/s41428-023-00804-6>
- Teresia, N., Zakiah, Z., & Turnip, M. (2024). Induksi Kalus dari Hipokotil Belimbing Merah (*Baccaurea angulata*) dengan Penambahan 2, 4-D (Dichlorophenoxy Acetic Acid) dan BAP (6-Benzyl Amino Purin). *Jurnal Biologi Tropis*, 24(1), 194-203. doi: <http://dx.doi.org/10.29303/jbt.v24i1.6387>
- Thejashree, A. B. & Naika, R. (2023). Identification of Bioactive Compounds in Acetone Leaf and Stem-Bark Extracts of *Psychotria dalzellii* Hook. f. by GC-MS Analysis and Evaluation of In Vitro Anti-Bacterial Properties. *Asian Journal of Biological and Life Sciences*, 12(3), 499.
- Tien, D. N., Oo, M. M., Soh, M. S., & Park, S. K. (2013). Bioengineering of Male Sterility in Rice (*Oryza sativa* L.). *Plant Breed. Biotechnol*, 1, 218-235. doi: <http://dx.doi.org/10.9787/PBB.2013.1.3.218>
- Tiwari, A.K., Gupta, M.K., Pandey, G., & Pandey, P.C. (2023). Siloxane-Silver Nanofluid as Potential Self-Assembling Disinfectant: A Preliminary Study on the Role of Functional Alkoxysilanes. *Nanoarchitectonics*, 4(1). doi: [10.37256/nat.4120231576](https://doi.org/10.37256/nat.4120231576)
- Tranggono, Firnanda, L.A., Nurfiyanti, W.P., Zahara, A. F. A., Angelina, V., & Nagara, N. P. (2023). Faktor–Faktor Yang Mempengaruhi Rendemen Tebu Yang Masih Belum Memenuhi Kebutuhan Gula Nasional. *Azzahra: Scientific Journal of Social Humanities*, 1, 63-72.

- Trejo-Tapia, G., Balcazar-Aguilar, J. B., Martínez-Bonfil, B., Salcedo-Morales, G., Jaramillo-Flores, M., Arenas-Ocampo, M. L., & Jiménez-Aparicio, A. (2008). Effect of Screening and Subculture on the Production of Betaxanthins in *Beta vulgaris* L. var. 'Dark Detroit' callus culture. *Innovative Food Science & Emerging Technologies*, 9(1), 32-36. doi: <https://doi.org/10.1016/j.ifset.2007.04.009>
- Triyanti, S. B., Lestari, F. P., Fitriana, P. A. N., Rostiana, H. R., Silalahi, D. D., Syalsabina, T. D., ... & Saputra, I. S. (2025). Pengaruh Metode Ekstraksi Maserasi, Sonikasi, dan Sokletasi Terhadap Nilai Rendemen Sampel Kulit Buah Naga (*Hylocereus polyrhizus*). *Jurnal Sains dan Edukasi Sains*, 8(1), 71-78. doi: <https://doi.org/10.24246/juses.v8i1p71-78>
- Twaij, B. M., Taha, A. J., Bhuiyan, F. H., & Hasan, M. N. (2022). Effect of Saccharides on Secondary Compounds Production from Stem Derived Callus of *Datura inoxia*. *Biotechnology Reports*, 33, e00701. doi: <https://doi.org/10.3390/ijpb13010003>
- Tyagi, T. & Agarwal, M. (2017). GC-MS Analysis of Invasive Aquatic Weed, *Pistia stratiotes* L. and *Eichhornia crassipes* (Mart.) Solms. *International Journal of Current Pharmaceutical Research*, 9(3), 111.
- Ucarli, C. (2022). Effects of Exogenous Linoleic Acid on Barley (*Hordeum vulgare* L.) Seedlings Under Salinity. *Journal of the Institute of Science and Technology*, 12(3), 1790-1800. doi: [10.21597/jist.1105133](https://doi.org/10.21597/jist.1105133)
- Ulia, R. V., Suryati., & Santoni, A. (2023). Cytotoxic Potential of Essential Oil Isolated from *Clibadium surinamense* L Leaves Against T47D Breast and HeLa Cervical Cancer Cells. *Molekul*, 18(2), 289-299.
- Ullah, Z., Ibrarullah, S. A. A., & Ozturk, M. (2022). Determination of Fatty Acids Profile, Antioxidant, and Anticholinesterase Inhibitory Activities of *Ganoderma lucidum* Collected From Swat. *Records Agric Food Chem*, 2(1), 41-49. doi: [10.25135/rfac.10.2202.2368](https://doi.org/10.25135/rfac.10.2202.2368)
- Valenzuela-Atondo, D. A., Delgado-Vargas, F., López-Angulo, G., Calderón Vázquez, C. L., Orozco-Cárdenas, M. L., & Cruz-Mendivil, A. (2020). Antioxidant activity of In Vitro Plantlets and Callus Cultures of *Randia Echinocarpa*, A Medicinal Plant From Northwestern Mexico. *In Vitro Cellular & Developmental Biology-Plant*, 56, 440-446. doi: <https://doi.org/10.1007/s11627-020-10062-3>
- Varshini, R., Manokiruthika, V., Jasayaprakash, S., Lakshmanan, H., & Muthusami, S., Sunganyadevi. P. (2024). Cycloheptasiloxane, Tetradecamethyl Identified from Petroleum Ether Fraction of *Sargassum Wightii* by Gas Chromatography–Mass Spectrometry Analysis. *Journal of Emerging Technologies and Innovative Research*, 11(8).
- Venkatramanan, M., Sankar Ganesh, P., Senthil, R., Akshay, J., Veera Ravi, A., Langeswaran, K., ... & Shankar, E. M. (2020). Inhibition of Quorum Sensing and Biofilm Formation in *Chromobacterium violaceum* by Fruit Extracts of

- Passiflora edulis*. *ACS omega*, 5(40), 25605-25616. doi: <https://dx.doi.org/10.1021/acsomega.0c02483>
- Verma, R., Kumar, A., Rani, L., & Singh, V. K. (2024). In Vitro Organogenesis of *Brassica juncea* (L.) from Callus as Explant from Stem and Leaf. *International Journal for Multidisciplinary Research*, 6(6).
- Wahyuni, A., Satria, B. & Zainal, A. (2020). Induksi Kalus Gaharu dengan NAA dan BAP Secara In Vitro. *Agrosains: Jurnal Penelitian Agronomi*, 22(1), 39-44. doi: <http://dx.doi.org/10.20961/agsjpa.v22i1.360071>
- Wahyuni, D. K., Huda, A., Faizah, S., Purnobasuki, H., & Wardoyo, B. P. E. (2020). Effects of Light, Sucrose Concentration and Repetitive Subculture on Callus Growth and Medically Important Production in *Justicia gendarussa* Burm. f. *Biotechnology Reports*, 27, e00473. doi: <https://doi.org/10.1016/j.btre.2020.e00473>
- Wang, X., Liu, B., Liu, Y., Wang, Y., Wang, Z., Song, Y., ... & Xue, C. (2023). Antioxidants Ameliorate Oxidative Stress in Alcoholic Liver Injury By Modulating Lipid Metabolism and Phospholipid Homeostasis. *Lipids*, 58(5), 229-240. doi: [10.1002/lipd.12377](https://doi.org/10.1002/lipd.12377)
- Wang, X., Zhang, C., & Bao, N. (2023). Molecular Mechanism of Palmitic Acid and Its Derivatives in Tumor Progression. *Frontiers in oncology*, 13, 1224125. doi: [10.3389/fonc.2023.1224125](https://doi.org/10.3389/fonc.2023.1224125)
- Wardani, D.P., Solichatun., & Setyawan, A.D. (2004). Pertumbuhan dan Produksi Saponin Kultur Kalus *Talinum paniculatum* Gaertn. Pada Variasi Penambahan Asam 2,4-Diklorofenoksi Asetat (2,4-D) dan Kinetin. *Biofarmasi*, 2(1), 35-43. doi: [10.13057/biofar/f020106](https://doi.org/10.13057/biofar/f020106)
- Wetter, L.R. & Constabel, F. (1991). *Metode Kultur Jaringan Tanaman*. Jakarta : ITB Press.
- Wibawa, I.P.A.H., Andila, P.S., Lugrayasa, I. N., & Sujarwo, W. (2021). Studi Potensi Tanaman Tebu Ireng (*Saccharum officinarum* L.) sebagai Antioksidan dan Antibakteri. *Jurnal Ilmu-Ilmu Hayati*, 20(1).
- Widiawati & Qodri, U. L. (2023). Analisis Fitokimia Dan Penentuan Kadar Fenolik Total Pada Ekstrak Etanol Tebu Merah Dan Tebu Hijau (*Saccharum officinarum* L.). *Jurnal Farmasi Tinctura*, 4(2), 91-102.
- Widiayani, N., Jasadina, I. M., & Nasruddin, N. (2024). Pengaruh Konsentrasi Auksin dan Sitokinin Terhadap Keberhasilan dan Pertumbuhan Stek Tanaman Kakao (*Theobroma cacao* L.). *Jurnal Agrivigor*, 40-59. doi: <https://doi.org/10.20956/ja.v15i1.43233>
- Widyastuti, N. & Deviyanti, J. (2024). *Kultur Jaringan—Teori dan Praktik Perbanyak Tanaman Secara In-Vitro*. Yogyakarta : Andi.
- Wijaya, D. C. & Linawati, N. M. (2024). The Role of Alpha Linolenic Acid on Neuroinflammation: A Systemic Review. *Jurnal Penelitian Pendidikan IPA*, 10(7), 3597-3604. doi: [10.29303/jppipa.v10i7.6173](https://doi.org/10.29303/jppipa.v10i7.6173)

- Wilson, I. D., Adlard, E. R., Cooke, M., & Poole, C. F. (2000). *Encyclopedia of separation science*. New York : Academic Press.
- Windiyani, I. P., Mahfut, M., Purnomo, P., & Daryono, B. S. (2022). Morphological Variations of Superior Sugarcane Cultivars (*Saccharum officinarum*) from Lampung, Indonesia. *Biodiversitas Journal of Biological Diversity*, 23(8).
- Wudali, S., Nagella, P., Al-Khayri, J. & Jain, S. M. (2022). *Role of Plant Tissue Culture Medium Components*. United Kingdom : Academic Press.
- Yukamgo, E., & Yuwono, W. (2007). Peran Silikon Sebagai Unsur Bermanfaat Pada Tanaman Tebu, *Jurnal Ilmu Tanah dan lingkungan*, 7(2), 103–116.
- Yunianto, M. F., Faisol, A., & Vendyansyah, N. (2021). Sistem Informasi Geografis Pemetaan Hasil Produksi Tebu Dengan Metode K-Means Di Kabupaten Malang. *JATI (Jurnal Mahasiswa Teknik Informatika)*, 5(2), 573-580.
- Yuniardi, F. (2019). Aplikasi Dimmer Switch pada Rak Kultur sebagai Pengatur Kebutuhan Intesitas Cahaya Optimum Bagi Tanaman *In Vitro*. *Indonesian Journal of Laboratory*, 1(4), 8-13.
- Zahrah, H., Mustika, A., & Debora, K. (2019). Aktivitas Antibakteri dan Perubahan Morfologi dari *Propionibacterium Acnes* setelah Pemberian Ekstrak *Curcuma xanthorrhiza*. *Jurnal Biosains Pascasarjana*, 20(3), 160. doi: <https://doi.org/10.20473/jbp.v20i3.2018.160-169>
- Zamir, R., Khalil, S. A., Shah, S. T., Ahmad, N., & Saima, S. (2012). Antioxidant Activity Influenced by in Vivo And In Vitro Mutagenesis in Sugarcane (*Saccharum officinarum* L.). *African Journal of Biotechnology*, 11(54), 11686-11692. doi: [10.5897/AJB11.3478](https://doi.org/10.5897/AJB11.3478)
- Zhang, Q. W., Lin, L. G., & Ye, W. C. (2018). Techniques for Extraction and Isolation Of Natural Products: A Comprehensive Review. *Chinese medicine*, 13, 1-26. doi: <https://doi.org/10.1186/s13020-018-0177-x>
- Zhang, S., Wu, S., Hou, Q., Zhao, J., Fang, C., An, X., & Wan, X. (2024). Fatty acid de novo biosynthesis in plastids: Key enzymes and their critical roles for male reproduction and other processes in plants. *Plant Physiology and Biochemistry*, 20, 108654. doi: <https://doi.org/10.1016/j.plaphy.2024.108654>
- Zhukov, A. V. (2015). Palmitic Acid and Its Role in the Structure and Functions of Plant Cell Membranes. *Russian journal of plant physiology*, 62, 706-713.
- Zi, X., Zhou, S., & Wu, B. (2022). Alpha-Linolenic Acid Mediates Diverse Drought Responses in Maize (*Zea mays* L.) at Seedling and Flowering Stages. *Molecules*, 27(3), 771. doi: <https://doi.org/10.3390/molecules27030771>
- Zulkarnain. (2009). *Kultur Jaringan Tanaman : Solusi Perbanyak Tanaman Budi Daya*. Jakarta: PT. Bumi Aksara.