

**RESPONS EKSPLAN DAUN *Nepenthes gymnamphora* PADA MEDIUM
½ MS DENGAN PENAMBAHAN 2,4-DIKLOROFENOKSIASETAT
DAN AIR KELAPA**

SKRIPSI

Diajukan untuk memenuhi sebagian syarat memperoleh gelar Sarjana Sains
Program Studi Biologi



oleh:

Salma Annisa Nur Alifah

NIM 2008156

PROGRAM STUDI BIOLOGI
FAKULTAS PENDIDIKAN MATEMATIKA
DAN ILMU PENGETAHUAN ALAM
UNIVERSITAS PENDIDIKAN INDONESIA

2024

Respons Eksplan Daun *Nepenthes gymnamphora* pada Medium $\frac{1}{2}$ MS dengan Penambahan 2,4-Diklorofenoksiasetat dan Air Kelapa

Oleh
Salma Annisa Nur Alifah

Sebuah skripsi yang diajukan untuk memenuhi salah satu syarat memperoleh gelar Sarjana Sains pada Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam

© Salma Annisa Nur Alifah 2024
Universitas Pendidikan Indonesia
Juli 2024

Hak Cipta dilindungi undang-undang.
Skripsi ini tidak boleh diperbanyak seluruhnya atau sebagian,
dengan dicetak ulang, difoto kopi, atau cara lainnya tanpa ijin dari penulis.

LEMBAR PENGESAHAN

SALMA ANNISA NUR ALIFAH

**RESPONS EKSPLAN DAUN *Nepenthes gymnamphora* PADA MEDIUM
½ MS DENGAN PENAMBAHAN 2,4-DIKLOROFENOKSIASETAT
DAN AIR KELAPA**

Disetujui dan disahkan oleh pembimbing:

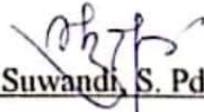
Pembimbing I,



Dr. rer. nat. Adi Rahmat, M.Si.

NIP. 196512301992021001

Pembimbing II,



Tri Suwandi, S. Pd., M.Sc.

NIP. 199005142018031001

Mengetahui,

Ketua Program Studi Biologi



Dr. Wahyu Surakusumah, M.T.

NIP. 197212301999031001

**RESPONS EKSPLAN DAUN *Nepenthes gymnamphora* PADA MEDIUM
½ MS DENGAN PENAMBAHAN 2,4-DIKLOROFENOKSIASETAT DAN
AIR KELAPA**

ABSTRAK

Nepenthes gymnamphora adalah salah satu species tumbuhan karnivora yang ditemukan di Pulau Jawa dan Sumatera. Tumbuhan ini mengalami kelangkaan akibat tingginya permintaan serta kerusakan habitat, sehingga upaya konservasi perlu segera dilakukan. Kesulitan dalam perbanyakan *Nepenthes gymnamphora* dapat diatasi dengan teknik kultur jaringan. Namun, penelitian mengenai perbanyakan kultur jaringan khususnya zat pengatur tumbuh yang diperlukan masih terbatas. Zat pengatur tumbuh seperti 2,4-D dan stimulator air kelapa sering digunakan untuk memaksimalkan hasil kultur jaringan. Penelitian ini bertujuan untuk menentukan kombinasi terbaik 2,4-D dan air kelapa dalam merangsang respons eksplan daun *Nepenthes gymnamphora* menuju pembentukan planlet. Media yang digunakan adalah ½ MS dengan variasi 2,4-D (0 ppm; 0,5 ppm; 1 ppm; 1,5 ppm; 2 ppm) dan air kelapa (0%, 10%, 15%, 20%, 25%). Hasil penelitian menunjukkan adanya perubahan morfologi berupa pelengkungan dan perubahan warna dari hijau menjadi kecoklatan. Konsentrasi 2,4-D memberikan pengaruh signifikan terhadap perubahan morfologi dan warna eksplan, sedangkan air kelapa tidak memberikan pengaruh signifikan. Konsentrasi 2,4-D yang paling mempengaruhi pelengkungan adalah 1,5 ppm, sedangkan tingkat *browning* paling dipengaruhi oleh konsentrasi 2 ppm.

Kata Kunci: 2,4-Diklorofenoksiasetat (2,4-D), Air Kelapa, Medium ½ MS, *Nepenthes gymnamphora*.

**RESPONSE OF *Nepenthes gymnamphora* LEAF EXPLANT ON ½ MS
MEDIUM WITH THE ADDITION OF 2,4-DICHLOROPHENOXYACETIC
ACID AND COCONUT WATER**

ABSTRACT

Nepenthes gymnamphora is one of the carnivorous plant species found in Java and Sumatra. This plant is experiencing scarcity due to high demand and habitat destruction, so immediate conservation efforts are needed. Difficulties in propagating *Nepenthes gymnamphora* can be overcome by tissue culture techniques. However, research on tissue culture propagation, especially the required growth regulators, is still limited. Growth regulators such as 2,4-D and coconut water stimulators are often used to maximize tissue culture results. This study aims to determine the best combination of 2,4-D and coconut water in stimulating the response of *Nepenthes gymnamphora* leaf explants towards plantlet formation. The media used was ½ MS with variations of 2,4-D (0 ppm; 0.5 ppm; 1 ppm; 1.5 ppm; 2 ppm) and coconut water (0%, 10%, 15%, 20%, 25%). The results showed morphological changes in the form of curvature and color changes from green to brownish. The concentration of 2,4-D gave a significant effect on the morphological and color changes of the explants, while coconut water did not give a significant effect. The concentration of 2,4-D that most affected the curvature was 1,5 ppm, while the browning level was most influenced by the concentration of 2 ppm.

Keywords: 2,4-Dichlorophenoxyacetic acid, Coconut Water, ½ MS Medium, *Nepenthes gymnamphora*.

DAFTAR ISI

LEMBAR PENGESAHAN.....	iii
LEMBAR PERNYATAAN	iv
UCAPAN TERIMA KASIH.....	v
ABSTRAK	vii
ABSTRACT	viii
DAFTAR ISI.....	ix
DAFTAR TABEL	xi
DAFTAR GAMBAR	xii
DAFTAR LAMPIRAN	xiii
BAB I PENDAHULUAN	14
1.1 Latar Belakang Penelitian	14
1.2 Rumusan Masalah	17
1.3 Pertanyaan Penelitian	17
1.4 Batasan Masalah.....	18
1.5 Tujuan Penelitian.....	18
1.6 Manfaat Penelitian.....	18
1.7 Asumsi.....	19
1.8 Hipotesis.....	19
1.9 Struktur Organisasi Skripsi	19
BAB II KULTUR JARINGAN <i>Nepenthes gymnamphora</i>	21
2.1 Tinjauan Umum Kantong Semar <i>Nepenthes gymnamphora</i>	21
2.2 Kultur Jaringan Tumbuhan.....	23
2.2.1 Media Kultur	25
2.2.2 Zat Pengatur Tumbuh.....	27
2.3 Respons Eksplan.....	30
2.4 <i>Browning</i>	32
2.5 Kontaminasi.....	33
BAB III METODE PENELITIAN.....	35
3.1 Jenis Penelitian	35
3.2 Desain Penelitian.....	35
3.3 Waktu dan Lokasi Penelitian.....	37

3.4	Alat dan Bahan	37
3.5	Populasi dan Sampel	37
3.6	Prosedur Penelitian.....	37
3.6.1	Tahap Persiapan Penelitian	37
3.6.2	Tahap Pelaksanaan	42
3.6.3	Tahap Pengumpulan Data	45
3.6.4	Analisis Data	45
3.7	Alur Penelitian.....	46
BAB IV TEMUAN DAN PEMBAHASAN		47
4.1	Respons Perubahan Morfologi Eksplan	47
4.1.1	Interaksi 2,4-D dan Air Kelapa terhadap Perubahan Morfologi Eksplan	49
4.1.2	Efek 2,4-D terhadap Perubahan Morfologi Eksplan	51
4.1.3	Efek Air Kelapa terhadap Perubahan Morfologi Eksplan	54
4.2	Respons Perubahan Warna Eksplan	55
4.2.1	Interaksi 2,4-D dan Air Kelapa terhadap Perubahan Warna Eksplan.....	59
4.2.2	Efek 2,4-D terhadap Perubahan Warna Eksplan.....	60
4.2.3	Efek Air Kelapa terhadap Perubahan Warna Eksplan	63
4.3	Faktor Penghambat Pertumbuhan dan Perkembangan Eksplan	64
BAB V SIMPULAN, IMPLIKASI, DAN REKOMENDASI		67
5.1	Simpulan.....	67
5.2	Implikasi	67
5.3	Rekomendasi	67
DAFTAR PUSTAKA		69
LAMPIRAN		80
RIWAYAT HIDUP		98

DAFTAR TABEL

Tabel 3.1 Kombinasi Media dengan Air Kelapa dan 2,4-D.....	36
Tabel 4.1 Hasil Uji ANOVA RAL Faktorial pada Eksplan Memberikan Respon	50
Tabel 4.2 Hasil Uji Statistik DMRT 2,4-D pada Eksplan Memberikan Respons.	50
Tabel 4.3 Rata-Rata Eksplan Memberikan Respons pada Konsentrasi 2,4-D.....	51
Tabel 4.4 Hasil Uji ANOVA RAL Faktorial pada Eksplan Berubah Warna.....	59
Tabel 4.5 Hasil Uji Statistik DMRT 2,4-D pada Eksplan Mengalami <i>Browning</i> .	60
Tabel 4.6 Rata-Rata Eksplan Mengalami <i>Browning</i> pada Konsentrasi 2,4-D.....	61

DAFTAR GAMBAR

Gambar 2.1 <i>Nepenthes gymnamphora</i>	22
Gambar 2.2 Jalur regenerasi eksplan.....	31
Gambar 3.1 Proses Pembuatan Larutan Stok	38
Gambar 3.2 Proses Pembuatan Media Kultur	40
Gambar 3.3 Proses Sterilisasi Alat dan Akuades	41
Gambar 3.4 Proses Pengambilan Eksplan.....	42
Gambar 3.5 Proses Sterilisasi Eksplan.....	43
Gambar 3.6 Proses Penanaman Eksplan	45
Gambar 3.7 Diagram Alur Penelitian.....	46
Gambar 4.1 Grafik Persentase Eksplan Memberikan Respons.....	48
Gambar 4.2 Eksplan yang Melengkung	49
Gambar 4.3 Grafik Polinomial Pengaruh 2,4-D terhadap Eksplan Memberikan Respons	52
Gambar 4.4 Grafik Persentase Eksplan yang Bertahan Hijau.....	55
Gambar 4.5 Eksplan yang Bertahan Hijau	56
Gambar 4.6 Grafik Persentase Eksplan Mengalami Browning.....	57
Gambar 4.7 Eksplan yang Mengalami Browning	58
Gambar 4.8 Grafik Polinomial Pengaruh 2,4-D terhadap Eksplan Browning	62
Gambar 4.9 Kontaminasi pada Kultur.....	65

DAFTAR LAMPIRAN

Lampiran 1. Alat dan Bahan Penelitian.....	80
Lampiran 2. Komposisi Media Murashige-Skoog	82
Lampiran 3. Pendataan Eksplan Daun <i>Nepenthes gymnamphora</i>	84
Lampiran 4. Dokumentasi Kegiatan.....	86
Lampiran 5. Hasil Olah Data Statistika.....	93
Lampiran 6. Surat Izin Pengambilan Eksplan	93

DAFTAR PUSTAKA

- Adamczuk, A., Siegień, I., & Ciereszko, I. (2012). *Morphogenesis of plants in vitro under stress conditions*. Polish Botanical Society.
- Ahmadi-Sakha, S., Sharifi, M., Niknam, V., & Zali, H. (2022). Production of phenylethanoid glycosides under PEG-induced osmotic stress in *Scrophularia Striata* Boiss. Cell culture in bioreactor. *Industrial Crops and Products*, 181, 114843. <https://doi.org/10.1016/j.indcrop.2022.114843>
- Akhiriana, E., Samanhudi, S., & Yunus, A. (2019). Coconut Water and IAA Effect on the *In vitro* Growth of *Tribulus terrestris* L. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*, 67(1), 9–18. <https://doi.org/10.11118/actaun201967010009>
- Al-Khayri, J. M. (2010). Somatic Embryogenesis of Date Palm (*Phoenix dactylifera* L.) Improved by Coconut Water. *Biotechnology(Faisalabad)*, 9(4), 477–484. <https://doi.org/10.3923/biotech.2010.477.484>
- Al-Khayri, J. M., Huang, F. H., Morelock, T. E., & Busharar, T. A. (2019). Spinach Tissue Culture Improved with Coconut Water. *HortScience*, 27(4), 357–358. <https://doi.org/10.21273/HORTSCI.27.4.357>
- Aliyu, & Mashood, O. (2005). Application of tissue culture to cashew (*Anacardium occidentale* L.) breeding: An appraisal. *African Journal of Biotechnology*, Vol. 4.
- Amente, G., & Chimdessa, E. (2021). Control of *browning* in plant tissue culture: A review. *Journal of Scientific Agriculture*, 67–71. <https://doi.org/10.25081/jsa.2021.v5.7266>
- Arsela, P. (2022). *The Effect of Ms Media Concentration and Coconut Water Addition on Pitcher Plant (Nepenthes mirabilis) In vitro Germination*. 15(7).
- Asmono, S. L., Rahmawati, & Sjamsijah, N. (2021). The effect of murashige and skoog (MS) modified medium and several types of auxins on the growth of stevia (*Stevia rebaudiana bertonii*) *in vitro*. *IOP Conference Series: Earth and Environmental Science*, 672(1), 012001. <https://doi.org/10.1088/1755-1315/672/1/012001>
- Awua, A. K., Doe, E. D., & Agyare, R. (2011). Exploring the influence of sterilisation and storage on some physicochemical properties of coconut (*Cocos nucifera* L.) water. *BMC Research Notes*, 4(1), 451. <https://doi.org/10.1186/1756-0500-4-451>
- Bahadur, V., Kirad, K. S., Mathew, A., & Singh, D. B. (2008). TISSUE CULTURE STUDIES IN NEPENTHES KHASIANA. *Acta Horticulturae*, 786, 287–293. <https://doi.org/10.17660/ActaHortic.2008.786.35>
- Baharan, E., Pour Mohammadi, P., Ehsan, S., & Hosseini, Z. (2015). Effects of some plant growth regulators and light on callus induction and explants browning in date palm (*Phoenix dactylifera* L.) *in vitro* leaves culture. *Iranian Journal of Plant Physiology*, 5, 1473–1481.
- Ball, S. T., Zhou, H., & Konzak, C. F. (1993). Influence of 2,4-D, IAA, and duration of callus induction in anther cultures of spring wheat. *Plant Science*, 90(2), 195–200. [https://doi.org/10.1016/0168-9452\(93\)90240-Z](https://doi.org/10.1016/0168-9452(93)90240-Z)
- Bhojwani, S. S., & Razdan, M. K. (1996). Plant tissue culture: Theory and Practice, a Revised Edition. Dalam *Studies in Plant Science* (Vol. 5, hlm. ix). Elsevier. [https://doi.org/10.1016/S0928-3420\(96\)80002-4](https://doi.org/10.1016/S0928-3420(96)80002-4)

- Bednarek, P. T., Orłowska, R., Koebner, R. M., & Zimny, J. (2007). Quantification of the tissue-culture induced variation in barley (*Hordeum vulgare* L.). *BMC Plant Biology*, 7(1), 10. <https://doi.org/10.1186/1471-2229-7-10>
- Budisantoso, I., Indriani, M., & Kamsinah, K. (2018). Effect of BAP (6-Benzyl Amino Purine) Concentration on Growth Micro Cutting of *Nepenthes ampullaria*. *Biosaintifika: Journal of Biology & Biology Education*, 10(3), Article 3. <https://doi.org/10.15294/biosaintifika.v10i3.15718>
- Bunn, E., & Tan, B. (2004). Microbial Contaminants in Plant Tissue Culture Propagation. Dalam K. Sivasithamparama, K. W. Dixon, & R. L. Barrett (Ed.), *Microorganisms in Plant Conservation and Biodiversity* (hlm. 307–335). Kluwer Academic Publishers. https://doi.org/10.1007/0-306-48099-9_12
- Chua, L. S. L., & Henshaw, G. (1999). *IN VITRO* PROPAGATION OF *NEPENTHES MACFARLANEI*. *Journal of Tropical Forest Science*, 11(3), 631–638.
- Clarke, C. (1997). *Nepenthes of Borneo*. Kota Kinabalu: Natural History Publications in association with Science and Technology Unit, Sabah. <http://archive.org/details/nepenthesofborne0000clar>
- Clarke, C., & Moran, J. A. (2016). Climate, soils and vicariance—Their roles in shaping the diversity and distribution of *Nepenthes* in Southeast Asia. *Plant and Soil*, 403(1–2), 37–51. <https://doi.org/10.1007/s11104-015-2696-x>
- Clarke, C., Schlauer, J., Moran, J., & Robinson, A. (2018). *Systematics and evolution of Nepenthes* (Vol. 1). Oxford University Press. <https://doi.org/10.1093/oso/9780198779841.003.0005>
- Dangariya, M., Khandhar, D., Monpara, J., Chudasama, K., & Thaker, V. (2020). Detection and identification of microbial contaminants from plant tissue culture. *Tropical Plant Research*, 7(2), 388–395. <https://doi.org/10.22271/tpr.2020.v7.i2.045>
- Dewanti, P., Maryam, S. A., Widuri, L. I., & Okviandari, P. (2020). PERFORMANCE OF SOMATIC EMBRYOGENESIS DEVELOPMENT UNDER DIFFERENT 2,4-D AND COCONUT WATER CONCENTRATION IN SUGARCANE VAR. BULULAWANG. *BIOVALENTIA: Biological Research Journal*, 6(1), Article 1. <https://doi.org/10.24233/BIOV.6.1.2020.155>
- Dinarti, D., Sayekti, U., & Alitalia, Y. (2010). Kultur Jaringan Kantong Semar (*Nepenthes mirabilis*). *Jurnal Hortikultura Indonesia*, 1(2), Article 2. <https://doi.org/10.29244/jhi.1.2.59-65>
- Dong, Y., Fu, C., Su, P., Xu, X., Yuan, J., Wang, S., Zhang, M., Zhao, C., & Yu, L. (2016). Mechanisms and effective control of physiological *browning* phenomena in plant cell cultures. *Physiologia Plantarum*, 156(1), 13–28. <https://doi.org/10.1111/pp1.12382>
- Duan, C. (2004). Advances of explant *browning* in plant tissue culture. *Letters in Biotechnology*. <https://www.aminer.org/pub/622996c95aee126c0f0ada96/advances-of-explant-browning-in-plant-tissue-culture>
- El Amine, B., Mosseddaq, F., Naciri, R., & Oukarroum, A. (2023). Interactive effect of Fe and Mn deficiencies on physiological, biochemical, nutritional and growth status of soybean. *Plant Physiology and Biochemistry*, 199,

107718. <https://doi.org/10.1016/j.plaphy.2023.107718>
- Erniwitama, E., Taryono, T., & Murti, R. H. (2018). Effect of Explant Types and Benzyl Amino Purine Concentrations on the *In vitro* Regeneration of Several Local Eggplant Cultivars. *Agrotechnology Innovation (Agrinova)*, 1(1), Article 1. <https://doi.org/10.22146/agrinova.41750>
- Estévez, E. R. (2021). *Why Coconut Water is both a Biostimulant and an Anti-cancer Agent*. Department of Biosystems and Technology.
- Fatmawati, A. (2008). *Kajian Konsentrasi BAP dan 2,4-D Terhadap Induksi Kalus Tanaman Artemisia annua L. Secara In vitro*. Fakultas Pertanian, Universitas Sebelas Maret.
- Feng, F., Li, H., & Xie, J. (2002, Januari 1). *Rapid propagation of nepenthes mirabilis by tissue cultur*. Chinese Journal of Tropical Crops.
- Fitriani, Y., Wijana, G., & Darmawati, I. A. P. (2019). TEKNIK STERILISASI DAN EFEKTIVITAS 2,4-D TERHADAP PEMBENTUKAN KALUS EKSPAN DAUN NILAM (Pogostemon cablin Benth) *IN VITRO*. *Jurnal of Agricultural Science and Biotechnology*, 8(1). <https://jurnal.harianregional.com/jasb/id-54630>
- Guimarães, J. de J., Sousa, F. G. G. de, Román, R. M. S., Dal Pai, A., Rodrigues, S. A., & Sarnighausen, V. C. R. (2021). Effect of irrigation water pH on the agronomic development of hops in protected cultivation. *Agricultural Water Management*, 253, 106924. <https://doi.org/10.1016/j.agwat.2021.106924>
- George, E. F., Hall, M. A., & Klerk, G.-J. de. (2008a). *Plant propagation by tissue culture* (3rd ed). Springer.
- George, E. F., Hall, M. A., & Klerk, G.-J. D. (2008b). The Components of Plant Tissue Culture Media II: Organic Additions, Osmotic and pH Effects, and Support Systems. Dalam E. F. George, M. A. Hall, & G.-J. D. Klerk (Ed.), *Plant Propagation by Tissue Culture: Volume 1. The Background* (hlm. 115–173). Springer Netherlands. https://doi.org/10.1007/978-1-4020-5005-3_4
- Gomes, G. L. B., & Scortecci, K. C. (2021). Auxin and its role in plant development: Structure, signalling, regulation and response mechanisms. *Plant Biology*, 23(6), 894–904. <https://doi.org/10.1111/plb.13303>
- Grafi, G., Florentin, A., Ransbotyn, V., & Morgenstern, Y. (2011). The Stem Cell State in Plant Development and in Response to Stress. *Frontiers in Plant Science*, 2. <https://doi.org/10.3389/fpls.2011.00053>
- Gulzar, B., Mujib, A., Qadir Malik, M., Mamgain, J., Syeed, R., & Zafar, N. (2020). Plant tissue culture: Agriculture and industrial applications. Dalam *Transgenic Technology Based Value Addition in Plant Biotechnology* (hlm. 25–49). Elsevier. <https://doi.org/10.1016/B978-0-12-818632-9.00002-2>
- Gupta, S., Singh, A., Yadav, K., Pandey, N., & Kumar, S. (2022). Chapter 2—Micropropagation for multiplication of disease-free and genetically uniform sugarcane plantlets. Dalam A. Chandra Rai, A. Kumar, A. Modi, & M. Singh (Ed.), *Advances in Plant Tissue Culture* (hlm. 31–49). Academic Press. <https://doi.org/10.1016/B978-0-323-90795-8.00015-1>
- Hai-bo, H. (2006). Main Problems and Their Countermeasures in Plant Tissue Culture. *Journal of Anhui Agricultural Sciences*. <https://www.semanticscholar.org/paper/Main-Problems-and-Their-Countermeasures-in-Plant-Hai->

[bo/34a2bb1cd26bf17f49d0746ea9d7762b608c3bb9](https://doi.org/10.34a2bb1cd26bf17f49d0746ea9d7762b608c3bb9)

- Handayani, T. (2021). PERANAN TANAMAN KANTONG SEMAR (*Nepenthes spp*) DALAM KEHIDUPAN MANUSIA DAN LINGKUNGANNYA. *Gunung Djati Conference Series*, 6, 11–18.
- Haque, M. I., Singh, P. K., Ghuge, S., Kumar, A., Chandra Rai, A., Kumar, A., & Modi, A. (2022). Chapter 1 - A general introduction to and background of plant tissue culture: Past, current, and future aspects. Dalam A. Chandra Rai, A. Kumar, A. Modi, & M. Singh (Ed.), *Advances in Plant Tissue Culture* (hlm. 1–30). Academic Press. <https://doi.org/10.1016/B978-0-323-90795-8.00019-9>
- Hayashi, K., Fujita, Y., Ashizawa, T., Suzuki, F., Nagamura, Y., & Hayano-Saito, Y. (2016). Serotonin attenuates biotic stress and leads to lesion *browning* caused by a hypersensitive response to *Magnaporthe oryzae* penetration in rice. *The Plant Journal*, 85(1), 46–56. <https://doi.org/10.1111/tpj.13083>
- Herman, E. B. (2017). Plant tissue culture contamination: Challenges and opportunities. *Acta Horticulturae*, 1155, 231–238. <https://doi.org/10.17660/ActaHortic.2017.1155.33>
- Hettiarachchi, H. D. B. K., Vidhanaarachchi, V. R. M., Jayarathna, S. P. N. C., & Dinum, P. (2022). Effect of exogenous polyamines on coconut (*Cocos nucifera* L.) embryogenic callus multiplication. *COCOS*, 23(1), 47–56. <https://doi.org/10.4038/cocos.v23i1.5823>
- Hidayat, A. (2013, Januari 18). *Pengertian Dan Jenis Transformasi Data*. Uji Statistik. <https://www.statistikian.com/2013/01/transformasi-data.html>
- Hu, W., Fagundez, S., Katin-Grazzini, L., Li, Y., Li, W., Chen, Y., Wang, X., Deng, Z., Xie, S., McAvoy, R. J., & Li, Y. (2017). Endogenous auxin and its manipulation influence in vitro shoot organogenesis of citrus epicotyl explants. *Horticulture Research*, 4, 17071. <https://doi.org/10.1038/hortres.2017.71>
- Hunter, P. J., Atkinson, L. D., Vickers, L., Lignou, S., Oruna-Concha, M. J., Pink, D., Hand, P., Barker, G., Wagstaff, C., & Monaghan, J. M. (2017). Oxidative discolouration in whole-head and cut lettuce: Biochemical and environmental influences on a complex phenotype and potential breeding strategies to improve shelf-life. *Euphytica*, 213(8), 180. <https://doi.org/10.1007/s10681-017-1964-7>
- Huq, E. (2006). Degradation of negative regulators: A common theme in hormone and light signaling networks? *Trends in Plant Science*, 11(1), 4–7. <https://doi.org/10.1016/j.tplants.2005.11.005>
- Illahi, A. K., Ratnasari, E., & Dewi, S. K. (2022). Pengaruh 2,4-D terhadap Pertumbuhan Kalus Daun *Diospyros discolor* Willd pada Media MS secara *in vitro*. *LenteraBio: Berkala Ilmiah Biologi*, 11(3), 369–377. <https://doi.org/10.26740/lenterabio.v11n3.p369-377>
- Ingram, P. A., & Malamy, J. E. (2010). Chapter 2—Root System Architecture. Dalam J.-C. Kader & M. Delseny (Ed.), *Advances in Botanical Research* (Vol. 55, hlm. 75–117). Academic Press. <https://doi.org/10.1016/B978-0-12-380868-4.00002-8>
- Insani, F. (2013). *Induksi Kantong dengan Perlakuan Berbagai Konsentrasi Media Murashige & Skoog pada Beberapa Ukuran Eksplan Kantong Semar (Nepenthes gracilis Korth.) secara In vitro*.

<http://repository.ipb.ac.id/handle/123456789/63369>

- Isnaini, Y., Handayani, I., Novitasari, Y., Febrianto, Y., Erwansyah, D., Rukmanto, H., Setiabudi, M., & Asikin, D. (2021). *AKLIMATISASI DAN DISEMINASI BIBIT KANTONG SEMAR (Nepenthes spp.) HASIL KULTUR JARINGAN DI KEBUN RAYA BOGOR*. 19, 14–23.
- Integrated Taxonomic Information System (ITIS). (2023). *Nepenthes gymnamphora* Nees. [Online]. Diakses melalui: https://www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=896552#null
- Jain, S. M., & Nakhooda, M. (2017). Clonal and Micropropagation. Dalam *Encyclopedia of Applied Plant Sciences* (hlm. 428–432). Elsevier. <https://doi.org/10.1016/B978-0-12-394807-6.00144-1>
- Jeffri, W., Rafdinal, & Turnip, M. (2017). Keanekaragaman Jenis Kantong Semar (*Nepenthes* spp.) di Kawasan Pelestarian Plasma Nutfah (KPPN) PT. Mura Sungai Landak Kabupaten Mempawah. *Jurnal Protobiont*, 6(2). <https://doi.org/10.26418/protobiont.v6i2.20802>
- Jones, A. M. P., & Saxena, P. K. (2013). Inhibition of Phenylpropanoid Biosynthesis in *Artemisia annua* L.: A Novel Approach to Reduce Oxidative Browning in Plant Tissue Culture. *PLOS ONE*, 8(10), e76802. <https://doi.org/10.1371/journal.pone.0076802>
- Joshaghani, M. S., Ghasemnezhad, A., & Alizadeh, M. (2014). Effect of explants types, culture media and concentrations of plant growth regulator on callus induction rate in Artichoke (*Cynara scolymus* L.). *International Journal of Biotechnology Research*, 2(6), 070–074.
- Joshi, B., Panwar, G. S., & Singh, S. K. (2022). *In vitro* propagation of insectivorous plant *Nepenthes khasiana* Hook. f.- an endangered ornamental and ethnomedicinal species. *Vegetos*, 35(2), 534–539. <https://doi.org/10.1007/s42535-021-00310-1>
- Kassanuk, T., Selakorn, O., Phasinam, K., & Sutaphan, S. (2020). INFLUENCE OF ORGANIC ADDITIVES ON MULTIPLE SHOOT FORMATION OF MUSA (AA GROUP) ‘KLUAI NAM THAI’ *IN VITRO*. *Life Sciences and Environment Journal*, 21(2), Article 2.
- Khan, E. A., Yadav, M., Yadav, S., & Ahmed, H. M. I. (2023). Chapter 3—Role of auxins in regulating physiological and molecular aspects of plants under abiotic stress. Dalam A. Sharma, S. Pandey, R. Bhardwaj, B. Zheng, & D. K. Tripathi (Ed.), *The Role of Growth Regulators and Phytohormones in Overcoming Environmental Stress* (hlm. 39–65). Academic Press. <https://doi.org/10.1016/B978-0-323-98332-7.00008-1>
- Krishna, H., Sairam, R. K., Singh, S. K., Patel, V. B., Sharma, R. R., Grover, M., Nain, L., & Sachdev, A. (2008). Mango explant *browning*: Effect of ontogenic age, mycorrhization and pre-treatments. *Scientia Horticulturae*, 118(2), 132–138. <https://doi.org/10.1016/j.scienta.2008.05.040>
- Kulus, D., & Tymoszuk, A. (2020). Induction of Callogenesis, Organogenesis, and Embryogenesis in Non-Meristematic Explants of Bleeding Heart and Evaluation of Chemical Diversity of Key Metabolites from Callus. *International Journal of Molecular Sciences*, 21(16), Article 16. <https://doi.org/10.3390/ijms21165826>
- Kumar, P. P., & Loh, C. S. (2012). Plant tissue culture for biotechnology. Dalam

- Plant Biotechnology and Agriculture* (hlm. 131–138). Elsevier. <https://doi.org/10.1016/B978-0-12-381466-1.00009-2>
- Kunita, L. Y., Susiyanti, Isminingsih, S., & Isnaini, Y. (2011). *PERTUMBUHAN TANAMAN KANTONG SEMAR (Nepenthes rafflesiana Jack.) DENGAN MODIFIKASI KONSENTRASI MEDIA DAN pH SECARA IN VITRO*.
- Kuswandi, P. C., Prasetya, F., & Rahayu, R. S. (2023). The effects of 2,4-Dichlorophenoxyacetic acid and leaf surface orientation on callus induction of black betel (*Piper betle* L. var. *Nigra*). *Biogenesis: Jurnal Ilmiah Biologi*, *11*(1), 114–122. <https://doi.org/10.24252/bio.v11i1.35796>
- Laukkanen, H., Rautiainen, L., Taulavuori, E., & Hohtola, A. (2000). Changes in cellular structures and enzymatic activities during *browning* of Scots pine callus derived from mature buds. *Tree Physiology*, *20*(7), 467–475. <https://doi.org/10.1093/treephys/20.7.467>
- Lazar, T. (2003). Taiz, L. and Zeiger, E. Plant physiology. 3rd edn. *Annals of Botany*, *91*(6), 750–751. <https://doi.org/10.1093/aob/mcg079>
- Leifert, C., & Cassells, A. C. (2001). Microbial hazards in plant tissue and cell cultures. *In vitro Cellular & Developmental Biology - Plant*, *37*(2), 133–138. <https://doi.org/10.1007/s11627-001-0025-y>
- Li, Y., He, N., Hou, J., Xu, L., Liu, C., Zhang, J., Wang, Q., Zhang, X., & Wu, X. (2018). Factors Influencing Leaf Chlorophyll Content in Natural Forests at the Biome Scale. *Frontiers in Ecology and Evolution*, *6*. <https://doi.org/10.3389/fevo.2018.00064>
- Long, Y., Yang, Y., Pan, G., & Shen, Y. (2022). New Insights Into Tissue Culture Plant-Regeneration Mechanisms. *Frontiers in Plant Science*, *13*. <https://doi.org/10.3389/fpls.2022.926752>
- Mansur, M. (2002). *Nepenthes gymnamphora* Nees. *DI TAMAN NASIONAL GUNUNG HALIMUN DAN PENYEBARANNYA DI INDONESIA*.
- Mansur, M. (2007). KEANEKARAGAMAN JENIS *Nepenthes* (KANTONG SEMAR) DATARAN RENDAH DI KALIMANTAN TENGAH. *BERITA BIOLOGI*, *8*(5), Article 5. <https://doi.org/10.14203/beritabiologi.v8i5.1898>
- Mansur, M. (2013). TINJAUAN ULANG (REVIEW) TINJAUAN TENTANG *Nepenthes* (NEPENTHACEAE) DI INDONESIA [A Review of *Nepenthes* (*Nepenthaceae*) in Indonesia]. *BERITA BIOLOGI*, *12*(1), Article 1. <https://doi.org/10.14203/beritabiologi.v12i1.512>
- Mardhikasari, S., Yunus, A., & Samanhudi, S. (2019). Modification of Media for Banana *In vitro* Propagation with Foliar Fertilizer and Coconut Water in cv. Rajabulu. *Caraka Tani: Journal of Sustainable Agriculture*, *35*(1), Article 1. <https://doi.org/10.20961/carakatani.v35i1.27756>
- Meilani, V. (2014). *PENGARUH STERILISASI DAN KOMBINASI HORMON PADA EKSPAN KANTONG SEMAR (Nepenthes mirabilis Lour.) SECARA IN VITRO*. Institut Pertanian Bogor.
- Meinaswati, F. S., Setiari, N., Nurchayati, Y., & Suedy, S. W. A. (2022). RESPONSE OF SEED GERMINATION AND GROWTH OF *Nepenthes gymnamphora* Nees *IN VITRO* TO THE CONCENTRATION OF MS MINERAL SALT, PEPTONE AND THIDIAZURON. *Jurnal Bioteknologi & Biosains Indonesia (JBBI)*, *9*(1), Article 1. <https://doi.org/10.29122/jbbi.v9i1.5049>
- Miguel, S., Hehn, A., & Bourgaud, F. (2018). *Nepenthes*: State of the art of an

- inspiring plant for biotechnologists. *Journal of Biotechnology*, 265, 109–115. <https://doi.org/10.1016/j.jbiotec.2017.11.014>
- Miguel, S., Michel, C., Biteau, F., Hehn, A., & Bourgaud, F. (2020). *In vitro* plant regeneration and Agrobacterium-mediated genetic transformation of a carnivorous plant, *Nepenthes mirabilis*. *Scientific Reports*, 10(1), Article 1. <https://doi.org/10.1038/s41598-020-74108-7>
- Mohd Lazim, M. I., & Badruzaman, N. A. (2015). Quantification of Cytokinins in Coconut Water from Different Maturation Stages of Malaysiaâs Coconut (*Cocos nucifera* L.) Varieties. *Journal of Food Processing & Technology*, 6(11). <https://doi.org/10.4172/2157-7110.1000515>
- Muthi'ah, A., Sakya, A. T., Setyawati, A., Samanhudi, & Rahayu, M. (2023). *Callus induction of Calotropis gigantea using BAP and 2,4-D in vitro*. IOP Conference Series: Earth and Environmental Science. <https://doi.org/10.1088/1755-1315/1177/1/012021>
- Neumann, K.-H., Kumar, A., & Imani, J. (2009). *Plant cell and tissue culture: A tool in biotechnology ; basics and application*. Springer.
- Ng, Z. C., Tan, S. H., Mahmud, S. H. R. S., & Ma, N. L. (2020). Preliminary Study on Micropropagation of *Hylocereus polyrhizus* with Waste Coconut Water and Sucrose. *Materials Science Forum*, 981, 316–321. <https://doi.org/10.4028/www.scientific.net/MSF.981.316>
- Okoroafor, U. E. (2022). Microbial Contamination in Plant Tissue Culture and Elimination Strategies. *NIGERIAN AGRICULTURAL JOURNAL*, 53(2), 348–355.
- Novitasari, Y., & Isnaini, Y. (2021). PROPAGATION OF PITCHER PLANTS (*Nepenthes gracilis* KORTH. AND *Nepenthes reinwardtiana* MIQ.) THROUGH CALLUS INDUCTION. *Agric*, 33(2), Article 2. <https://doi.org/10.24246/agric.2021.v33.i2.p81-92>
- Nurokhman, A., Faizah, H., Utami, E. S. W., & Manuhara, Y. S. W. (2019). Effect of Plant Growth Regulator and Explant Types on *in vitro* Callus Induction of *Gynura procumbens* (Lour.) Merr. *Research Journal of Biotechnology*, 14.
- Pan, Z., Zhu, S., Guan, R., & Deng, X. (2010). Identification of 2,4-D-responsive proteins in embryogenic callus of Valencia sweet orange (*Citrus sinensis* Osbeck) following osmotic stress. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 103(2), 145–153. <https://doi.org/10.1007/s11240-010-9762-0>
- Pant, M., & Husen, A. (2022). Chapter 19—Micropropagation in mature trees by manipulation of phase change, stress, and culture environment. Dalam A. Husen (Ed.), *Environmental, Physiological and Chemical Controls of Adventitious Rooting in Cuttings* (hlm. 421–437). Academic Press. <https://doi.org/10.1016/B978-0-323-90636-4.00002-7>
- Park, S. (2021). *Plant tissue culture: Techniques and experiments* (4th ed). Academic press.
- Pasternak, T. P., & Steinmacher, D. (2024). Plant Growth Regulation in Cell and Tissue Culture *In vitro*. *Plants*, 13(2), Article 2. <https://doi.org/10.3390/plants13020327>
- Permadi, N., Nurzaman, M., Alhasnawi, A. N., Doni, F., & Julaeha, E. (2023). Managing Lethal *Browning* and Microbial Contamination in *Musa* spp. Tissue Culture: Synthesis and Perspectives. *Horticulturae*, 9(4), Article 4.

<https://doi.org/10.3390/horticulturae9040453>

- Pernisová, M., Klíma, P., Horák, J., Válková, M., Malbeck, J., Souček, P., Reichman, P., Hoyerová, K., Dubová, J., Friml, J., Zažímalová, E., & Hejátko, J. (2009). Cytokinins modulate auxin-induced organogenesis in plants via regulation of the auxin efflux. *Proceedings of the National Academy of Sciences of the United States of America*, 106(9), 3609–3614. <https://doi.org/10.1073/pnas.0811539106>
- Previaningrum, H., Qadir, A., & Isnaini, Y. (2021). Konservasi *In vitro* Kantong Semar (*Nepenthes rafflesiana* Jack.) dengan Metode Slow Growth. *Jurnal Jejaring Matematika dan Sains*, 3(1), Article 1. <https://doi.org/10.36873/jjms.2021.v3.i1.502>
- Purba, R. V., Yuswanti, H., & Astawa, I. N. G. (2017). Induksi Kalus Eksplan daun Tanaman Anggur (*Vitis vinifera* L.) dengan Aplikasi 2,4-D Secara *in vitro*. *Agroekoteknologi Tropika*, 6(2).
- Rahmadi, A., Wicaksana, N., Nurhadi, B., Suminar, E., Pakki, S. R. T., & Mubarak, S. (2020). Induksi Kalus pada Eksplan Daun Muda Tanaman Durian (*Durio zibethinus* Murr.) Klon Baru Kamajaya dengan Kombinasi Zat Pengatur Tumbuh 2,4-D dan Kinetin Secara *In Vitro*. *Agrikultura*, 31(3), Article 3. <https://doi.org/10.24198/agrikultura.v31i3.29388>
- Rahmy, N., Thomy, Z., Yunita, Y., & Harnelly, E. (2019). THE EFFECT OF SOME OF COCONUT WATER CONCENTRATION IN ARTIFICIAL MEDIA TO CHRYSANTHEMUM GROWTH (*Dendranthema grandiflora*) BY *IN VITRO*. *Jurnal Natural*. <https://doi.org/10.24815/JN.V19I2.11541>.
- Rai, A. C. (2022). *Advances in Plant Tissue Culture: Current Developments and Future Trends*. Academic Press.
- Ratmingsih, N. M. (2010). PENELITIAN EKSPERIMENTAL DALAM PEMBELAJARAN BAHASA KEDUA. *PRASI*, 6(11).
- Reis, R. S., Vale, E. M., Sousa, K. R., Santa-Catarina, C., & Silveira, V. (2021). Pretreatment free of 2,4-dichlorophenoxyacetic acid improves the differentiation of sugarcane somatic embryos by affecting the hormonal balance and the accumulation of reserves. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 145(1), 101–115. <https://doi.org/10.1007/s11240-020-01995-z>
- Ristiawan, H., & Hikmat, A. (2022). The PREY COMPOSITION OF *Nepenthes gymnamphora* Reinw. Ex Nees AT MOUNT BISMO, DERODUWUR HIKING TRAIL, WONOSOBO, CENTRAL JAVA. *Media Konservasi*, 27(3), 116–120. <https://doi.org/10.29244/medkon.27.3.116-120>
- Roy, J., & Banerjee, N. (2003). Induction of callus and plant regeneration from shoot-tip explants of *Dendrobium fimbriatum* Lindl. Var. *Oculatum* Hk. F. *Scientia Horticulturae*, 97, 333–340. [https://doi.org/10.1016/S0304-4238\(02\)00156-5](https://doi.org/10.1016/S0304-4238(02)00156-5)
- Safitri, S. K., Siregar, L. A. M., & Lubis, K. (2017). Induksi Kalus Tanaman Rosella (*Hibiscus sabdariffa* Linn.) pada Jenis Eksplan dan Konsentrasi Auksin yang Berbeda. *Jurnal Agroekoteknologi FP USU*, 5(3).
- Sathyanarayana, B. N., & Varghese, D. B. (2007). *Plant Tissue Culture: Practices and New Experimental Protocols*. I. K. International Pvt Ltd.
- Schwarz, O. J., & Beaty, R. M. (2000). Organogenesis. Dalam *Plant Tissue Culture*

Concepts and Laboratory Exercises (2 ed.). Routledge.

- Sehgal, H., & Joshi, M. (2022). Chapter 4—The journey and new breakthroughs of plant growth regulators in tissue culture. Dalam A. Chandra Rai, A. Kumar, A. Modi, & M. Singh (Ed.), *Advances in Plant Tissue Culture* (hlm. 85–108). Academic Press. <https://doi.org/10.1016/B978-0-323-90795-8.00002-3>
- Sivanesan, I., & Park, S. W. (2014). The role of silicon in plant tissue culture. *Frontiers in Plant Science*, 5. <https://doi.org/10.3389/fpls.2014.00571>
- Skirvin, R. M., Chu, M. C., Mann, M. L., Young, H., Sullivan, J., & Fermanian, T. (1986). Stability of tissue culture medium pH as a function of autoclaving, time, and cultured plant material. *Plant Cell Reports*, 5(4), 292–294. <https://doi.org/10.1007/BF00269825>
- Smith, R. H. (2013). *Plant tissue culture: Techniques and experiments* (Third edition). Academic Press.
- Sosnowski, J., Truba, M., & Vasileva, V. (2023). The Impact of Auxin and Cytokinin on the Growth and Development of Selected Crops. *Agriculture*, 13(3), Article 3. <https://doi.org/10.3390/agriculture13030724>
- Su, Y.-H., Liu, Y.-B., & Zhang, X.-S. (2011). Auxin–Cytokinin Interaction Regulates Meristem Development. *Molecular Plant*, 4(4), 616–625. <https://doi.org/10.1093/mp/ssr007>
- Sudheer, W. N., Praveen, N., Al-Khayri, J. M., & Jain, S. M. (2022). Chapter 3—Role of plant tissue culture medium components. Dalam A. Chandra Rai, A. Kumar, A. Modi, & M. Singh (Ed.), *Advances in Plant Tissue Culture* (hlm. 51–83). Academic Press. <https://doi.org/10.1016/B978-0-323-90795-8.00012-6>
- Sukamto, L. A. (2016). PENGARUH EKSPLOAN DAN ZPT TERHADAP PERTUMBUHAN NEPENTHES ALBOMARGINATA SECARA *In vitro*. *Jurnal Teknologi Lingkungan*, 12(1), 103. <https://doi.org/10.29122/jtl.v12i1.1267>
- Sukamto, L. A., & Henuhili, V. (2011). Shoot Tip Culture of *Nepenthes albomarginata* Lobb ex Lindl. *In vitro*. *Jurnal Biologi Indonesia*, 7(2), 251–261.
- Supriyanto, & Prakasa, K. E. (2011). Pengaruh Zat Pengatur Tumbuh Rootone-F Terhadap Pertumbuhan Stek Duabanga mollucana. Blume. *Jurnal Silvikultur Tropika*, 3(1).
- Tang, W., & Newton, R. J. (2004). Increase of polyphenol oxidase and decrease of polyamines correlate with tissue *browning* in Virginia pine (*Pinus virginiana* Mill.). *Plant Science*, 167(3), 621–628. <https://doi.org/10.1016/j.plantsci.2004.05.024>
- Tao, F., Zhang, Z., Zhou, J., Yao, N., & Wang, D. (2007). Contamination and *browning* in tissue culture of *Platanus occidentalis* L. *Forestry Studies in China*, 9(4), 279–282. <https://doi.org/10.1007/s11632-007-0044-9>
- Teixeira, M. C., Telo, J. P., Duarte, N. F., & Sá-Correia, I. (2004). The herbicide 2,4-dichlorophenoxyacetic acid induces the generation of free-radicals and associated oxidative stress responses in yeast. *Biochemical and Biophysical Research Communications*, 324(3), 1101–1107. <https://doi.org/10.1016/j.bbrc.2004.09.158>
- Trigiano, R. N., & Gray, D. J. (Ed.). (2005). *Plant development and biotechnology*.

CRC Press.

- Van Winkle, S. C., & Pullman, G. S. (2003). The combined impact of pH and activated carbon on the elemental composition of a liquid conifer embryogenic tissue initiation medium. *Plant Cell Reports*, 22(5), 303–311. <https://doi.org/10.1007/s00299-003-0686-6>
- Vanegas-Espinoza, P. E., Bazaldúa, C., Ríos-Salomé, L. B., Paredes-López, O., Camelo-Méndez, G. A., & Del Villar-Martínez, A. A. (2020). Cellular and morphological changes during leaf explant dedifferentiation and plant regeneration of *Tagetes erecta*. *Horticulture, Environment, and Biotechnology*, 61(2), 407–414. <https://doi.org/10.1007/s13580-019-00210-z>
- Wahyuni, D. K., Prasetyo, D., & Hariyanto, S. (2014). Perkembangan Kultur Daun *Aglaonema* sp. Dengan Perlakuan Kombinasi Zat Pengatur Tumbuh NAA dan 2,4-D dengan BAP. *BIOSLOGOS*, 4(1).
- Wang, Q., Yang, S., Wan, S., & Li, X. (2019). The Significance of Calcium in Photosynthesis. *International Journal of Molecular Sciences*, 20(6), 1353. <https://doi.org/10.3390/ijms20061353>
- Widiani, E., Perdana, R., Fakhri, M. A., Muhammad, F., Visda, D., Harahap, N., Andarini, F. M., Aziz, M. A., Aqsa, H., Sandra, I. E., & Si, M. (2019). *Keanekaragaman Jenis Kantong Semar dan Penyebarannya di Taman Nasional Bukit Barisan Selatan, Bengkulu*.
- Windiyani, I. P., Handayani, T. T., Zulkifli, Z., & Irawan, B. (2020). The Effect of Coconut Water (*Cocos nucifera* L.) and Atonik to The Growth of Tomato Plant (*Lycopersicum esculentum* Mill.). *Jurnal Ilmiah Biologi Eksperimen Dan Keanekaragaman Hayati (J-BEKH)*, 7(1), Article 1. <https://doi.org/10.23960/jbekh.v7i1.9>
- Wongsa, T., Inthima, P., Nakkuntod, M., Premjet, D., & Kongbangkerd, A. (2018). Effects of Cytokinin and Auxin on *In vitro* Organ Development and Plumbagin Content of *Drosera peltata* Thunb. *AGRIVITA Journal of Agricultural Science*, 40(3). <https://doi.org/10.17503/agrivita.v40i0.1276>
- Xu, C., Ru, Z., Zeng, B., Huang, J., Huang, W., & Hu, O. (2015). The Effects of Polyphenol Oxidase and Cycloheximide on the Early Stage of *Browning* in *Phalaenopsis* Explants. *Horticultural Plant Journal*, 1. <https://doi.org/10.16420/j.issn.2095-9885.2015-0030>
- Yamauchi, N. (2015). Postharvest Chlorophyll Degradation and Oxidative Stress. Dalam Y. Kanayama & A. Kochetov (Ed.), *Abiotic Stress Biology in Horticultural Plants* (hlm. 101–113). Springer Japan. https://doi.org/10.1007/978-4-431-55251-2_8
- Ye, X.-G., She, M.-Y., Wang, K., Du, L.-P., & Xu, H.-J. (2011). Identification, Cloning, and Potential Application of Genes Related to Somatic Embryogenesis in Plant Tissue Culture. *Acta Agron Sin*, 38(02), 191–201. <https://doi.org/10.3724/SP.J.1006.2012.00191>
- Yelli, F. (2020). INDUKSI PEMBENTUKAN KANTONG DAN PERTUMBUHAN DUA SPECIES TANAMAN KANTONG SEMAR (*Nepenthes* spp.) PADA BERBAGAI KONSENTRASI MEDIA MS SECARA *IN VITRO*. *JURNAL AGROTROPIKA*, 18(2), Article 2. <https://doi.org/10.23960/ja.v18i2.4297>
- Yong, J. W. H., Ge, L., Ng, Y. F., & Tan, S. N. (2009). The Chemical Composition

and Biological Properties of Coconut (*Cocos nucifera* L.) Water. *Molecules*,
14(12), Article 12. <https://doi.org/10.3390/molecules14125144>
Yuliarti, N. (2010). *Kultur Jaringan Tanaman Skala Rumah Tangga*. Penerbit Andi.