

**ANALISIS FILOGENETIK LAMIACEAE BERDASARKAN
BARKODE *ITS*, *matK*, DAN *rbcL* SECARA *IN SILICO***

SKRIPSI

Diajukan sebagai salah satu syarat untuk memperoleh
gelar Sarjana Sains pada Program Studi Biologi



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FAKULTAS PENDIDIKAN MATEMATIKA DAN ILMU PENGETAHUAN ALAM
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Skripsi yang diajukan untuk memenuhi salah satu syarat memperoleh gelar Sarjana Sains pada Program Studi Biologi Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam

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Dengan ini saya menyatakan bahwa skripsi dengan judul "Analisis Filogenetik Lamiaceae Berdasarkan Barkode *ITS*, *matK*, dan *rbcL* secara *In Silico*" ini beserta seluruh isinya adalah benar-benar karya saya sendiri. Saya tidak melakukan penjiplakan atau pengutipan dengan cara-cara yang tidak sesuai dengan etika ilmu yang berlaku dalam masyarakat keilmuan. Atas pernyataan ini, saya siap menanggung risiko/sanksi apabila di kemudian hari ditemukan adanya pelanggaran etika keilmuan atau ada klaim dari pihak lain terhadap keaslian karya saya ini.

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Yang membuat pernyataan,

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Dalam proses penulisan skripsi ini, penulis menyadari bahwa masih terdapat ruang untuk perbaikan. Oleh karena itu, saran dan kritik yang membangun sangat diharapkan agar skripsi ini dapat menjadi referensi yang lebih baik di masa mendatang. Semoga hasil dari skripsi ini dapat memberikan kontribusi positif dalam pengembangan ilmu pengetahuan bagi penulis dan pembaca yang berkepentingan.

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ANALISIS FILOGENETIK LAMIACEAE BERDASARKAN BARKODE *ITS*, *matK*, DAN *rbcL* SECARA *IN SILICO*

ABSTRAK

Lamiaceae kerap dimanfaatkan dalam pengobatan tradisional dan modern tetapi rentan terhadap adulterasi dan eksploitasi. Pendekatan filogenetik mampu menjadi solusi alternatif untuk mencegah adulterasi dan menjaga biodiversitas Lamiaceae. Namun, rekonstruksi filogenetik Lamiaceae terhambat karena kemiripan morfologi, diskrepansi filogenetik berbasis morfologi dan DNA, serta kurangnya studi filogenetik genus tertentu. Studi ini bertujuan untuk mendapatkan informasi filogenetik Lamiaceae berdasarkan barkode DNA inti (*ITS*) dan kloroplas (*matK* dan *rbcL*) parsial untuk mencegah adulterasi dan mendukung konservasi Lamiaceae. Daftar genus dicari di *Barcode of Life Data System*. Sekuens DNA parsial 52 spesies perwakilan 11 genus, dan *Spathodea campanulata* sebagai *outgroup*, didapat dari *GenBank NCBI*. Pensejajaran dan pemotongan DNA dilakukan sebelum empat pohon filogenetik *maximum parsimony* direkonstruksi (tiga berbasis satu barkode dan satu gabungan ketiganya) dengan MEGA 11. Analisis pohon meliputi prinsip *parsimony*, *bootstrapping*, indeks konsistensi (CI), dan indeks retensi (RI). Rerata sekuens terpanjang diperoleh *matK* (785.6 pb), tingkat homologi tertinggi *rbcL* (83.5%), dan jumlah situs *parsimony-informative* terbanyak *ITS* (40.3%). Setiap pohon mempunyai CI rendah (~0.5) dan RI tinggi (>0.5). Setiap barkode secara terpisah menunjukkan pola pengelompokan yang serupa pada genus yang berkerabat dekat tetapi memisahkan beberapa spesies dari genusnya. Gabungan ketiga barkode mengoreksi posisi spesies yang terpisah dan mengelompokkan Lamiaceae ke dalam enam klad monofiletik (*bootstrap* >70%): *Callicarpa*; *Scutellaria*; *Clerodendrum*, *Lamium*, dan *Stachys*; *Salvia*; *Thymus*, *Origanum*, dan *Mentha*; *Orthosiphon* dan *Ocimum*. Pohon filogenetik gabungan mampu mencegah adulterasi dan mendukung konservasi Lamiaceae karena memberikan informasi mengenai kekerabatan spesies target adulterasi yang bermanfaat untuk mengidentifikasi spesies adulteran dan menemukan spesies alternatif dengan sifat minyak esensial yang serupa.

Kata kunci: adulterasi, barkode DNA, filogenetik, Lamiaceae.

***IN SILICO PHYLOGENETIC ANALYSIS OF LAMIACEAE
BASED ON ITS, matK, AND rbcL DNA BARCODES***

ABSTRACT

*Lamiaceae, often utilized in traditional and modern medicine, are vulnerable to adulteration and exploitation. A phylogenetic approach can prevent these issues and conserve Lamiaceae biodiversity. However, reconstructing the phylogeny of Lamiaceae is hindered by morphological similarities, discrepancies between morphological and DNA data, and limited phylogenetic studies on certain genera. This study aimed to obtain phylogenetic information of Lamiaceae based on partial nuclear (ITS) and chloroplast (matK and rbcL) DNA barcodes to prevent adulteration and promote the conservation of Lamiaceae. Genera were obtained from the Barcode of Life Data System. Partial DNA sequences from the NCBI GenBank for 52 species across 11 genera (with *Spathodea campanulata* as the outgroup) were used. Sequences were aligned and trimmed to reconstruct four maximum parsimony trees using MEGA 11 (three based on one barcode and one combined). Tree analysis included parsimony principle, bootstrapping, consistency index (CI), and retention index (RI). The longest average sequence was found in matK (785.6 bp), highest homology in rbcL (83.5%), and most parsimony-informative sites in ITS (40.3%). Trees had low CI (~0.5) and high RI (>0.5). While individual barcodes showed similar genus groupings, they separated some species from their genera. Combining the barcodes corrected these positions, grouping Lamiaceae into six monophyletic clades (bootstrap >70%): *Callicarpa*; *Scutellaria*; *Clerodendrum*, *Lamium*, and *Stachys*; *Salvia*; *Thymus*, *Origanum*, and *Mentha*; *Orthosiphon* and *Ocimum*. The combined tree aids in preventing adulteration and conserving Lamiaceae by providing phylogenetic information on adulterated species, which is useful in identifying adulterants and finding alternative species with similar essential oil properties.*

Keywords: adulteration, DNA barcodes, phylogenetics, Lamiaceae.

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DAFTAR PUSTAKA

- Adhikari, B., Marasini, B. P., Rayamajhee, B., Bhattarai, B. R., Lamichhane, G., Khadayat, K., Khanal, S., & Parajuli, N. (2020). Potential roles of medicinal plants for the treatment of viral diseases focusing on COVID-19: A review. *Phytotherapy Research*, 35(3), 1298–1312. <https://doi.org/10.1002/ptr.6893>
- Afonso, A. F., Pereira, O. R., Fernandes, Â., Calhelha, R. C., Silva, A. M. S., Ferreira, I. C. F. R., & Cardoso, S. M. (2019). Phytochemical composition and bioactive effects of *Salvia africana*, *Salvia officinalis* “Icterina” and *Salvia mexicana* aqueous extracts. *Molecules*, 24(23), 4327. <https://doi.org/10.3390/molecules24234327>
- Ahmed, S. M. (2018). Molecular identification of *Lavendula dentata* L., *Mentha longifolia* (L.) Huds. and *Mentha × piperita* L. by DNA barcodes. *Bangladesh Journal of Plant Taxonomy*, 25(2), 149–157. <https://doi.org/10.3329/bjpt.v25i2.39519>
- Ajawatanawong, P. (2016). Molecular phylogenetics: Concepts for a newcomer. In I. Nookae (Ed.), *Advances in Biochemical Engineering/Biotechnology* (Vol 160, pp. 185-196). Springer.
- Al-Juhani, W. S., & Khalik, K. N. A. (2021). Identification and molecular study of medicinal *Plectranthus* species (Lamiaceae) from Saudi Arabia using plastid DNA regions and ITS2 of the nrDNA gene. *Journal of King Saud University - Science*, 33(5), 101452. <https://doi.org/10.1016/j.jksus.2021.101452>
- Alizadeh, F., Ramezani, M., & Piravar, Z. (2020). Effects of *Stachys sylvatica* hydroalcoholic extract on the ovary and hypophysis-gonadal axis in a rat with polycystic ovary syndrome. *Middle East Fertility Society Journal*, 25(1). <https://doi.org/10.1186/s43043-020-0015-9>
- Allkin, B. (2017). *Useful plants – medicines: At least 28,187 plant species are currently recorded as being of medicinal use*. In K. J. Willis (Ed.), *State of the World's Plants 2017*. London: Royal Botanic Gardens, Kew.
- Álvarez, I., & Wendel, J. F. (2003). Ribosomal ITS sequences and plant phylogenetic inference. *Molecular Phylogenetics and Evolution*, 29(3), 417–434. [https://doi.org/10.1016/s1055-7903\(03\)00208-2](https://doi.org/10.1016/s1055-7903(03)00208-2)
- Aminian, A. R., Mohebbati, R., & Boskabady, M. H. (2022). The effect of *Ocimum basilicum* L. and its main ingredients on respiratory disorders: an experimental, preclinical, and clinical review. *Frontiers in Pharmacology*, 12, 805391. <https://doi.org/10.3389/fphar.2021.805391>
- Aneva, I., Zhelev, P., Bonchev, G., Boycheva, I., Simeonova, S., & Kancheva, D. (2022). DNA barcoding study of representative *Thymus* species in bulgaria. *Plants*, 11(3), 270. <https://doi.org/10.3390/plants11030270>

- Arabaci, T., Çelenk, S., Özcan, T., Martin, E., Yazici, T., Açıar, M., Üzel, D., & Dirmenci, T. (2020). Homoploid hybrids of *Origanum* (Lamiaceae) in Turkey: morphological and molecular evidence for a new hybrid. *Plant Biosystems - an International Journal Dealing with All Aspects of Plant Biology*, 155(3), 1–13. <https://doi.org/10.1080/11263504.2020.1762777>
- Ashraf, K., Sultan, S., & Adam, A. (2018). *Orthosiphon stamineus* Benth. is an outstanding food medicine: Review of phytochemical and pharmacological activities. *Journal of Pharmacy and Bioallied Sciences*, 10(3), 109. https://doi.org/10.4103/jpbs.jpbs_253_17
- Assis, L. C. S., & Rieppel, O. (2011). Are monophyly and synapomorphy the same or different? Revisiting the role of morphology in phylogenetics. *Cladistics*, 27(1), 94–102. <https://doi.org/10.1111/j.1096-0031.2010.00317.x>
- Bahmani, M., & Zargaran, A. (2015). Ethno-botanical medicines used for urinary stones in the Urmia, Northwest Iran. *European Journal of Integrative Medicine*, 7(6), 657–662. <https://doi.org/10.1016/j.eujim.2015.09.006>
- Bai, C., Xu, J., Cao, B., Li, X., & Li, G. (2018). Transcriptomic analysis and dynamic expression of genes reveal flavonoid synthesis in *Scutellaria viscidula*. *Acta Physiologiae Plantarum*, 40, 161. <https://doi.org/10.1007/s11738-018-2733-5>
- Baldwin, B. G., Sanderson, M. J., Porter, J. M., Wojciechowski, M. F., Campbell, C. S., & Donoghue, M. J. (1995). The ITS region of nuclear ribosomal DNA: A valuable source of evidence on angiosperm phylogeny. *Annals of the Missouri Botanical Garden*, 82(2), 247–277. <https://doi.org/10.2307/2399880>
- Bast, F., Rani, P., & Meena, D. (2014). Chloroplast DNA phylogeography of holy basil (*Ocimum tenuiflorum*) in Indian subcontinent. *The Scientific World Journal*, 2014, 1–6. <https://doi.org/10.1155/2014/847482>
- Bayat, A. (2002). Science, medicine, and the future: Bioinformatics. *BMJ*, 324(7344), 1018–1022. <https://doi.org/10.1136/bmj.324.7344.1018>
- Begnini, K. R., Nedel, F., Lund, R. G., Carvalho, P. H. de A., Rodrigues, M. R. A., Beira, F. T. A., & Del-Pino, F. A. B. (2014). Composition and antiproliferative effect of essential oil of *Origanum vulgare* against tumor cell lines. *Journal of Medicinal Food*, 17(10), 1129–1133. <https://doi.org/10.1089/jmf.2013.0063>
- Bejar, E. (2019). *Adulteration of oregano herb and essential oil*. Botanical Adulterants Prevention Bulletin; American Botanical Council. <http://cms.herbalgram.org/BAP/BAB/OreganoHerbandEOBulletin.html?ts=1608160808&signature=a474e37d3291079d017ae38fc0825ff7>
- Bendiksby, M., Brysting, A. K., Thorbek, L., Gussarova, G., & Ryding, O. (2011a). Molecular phylogeny and taxonomy of the genus *Lamium* L. (Lamiaceae): Disentangling origins of presumed allotetraploids. *Taxon*, 60(4), 986–1000. <https://www.jstor.org/stable/41317321?seq=1>

- Bendiksby, M., Thorbek, L., Scheen, A.-C., Lindqvist, C., & Ryding, O. (2011b). An updated phylogeny and classification of Lamiaceae subfamily Lamioideae. *TAXON*, 60(2), 471–484. <https://doi.org/10.1002/tax.602015>
- Birkett, M. A., Hassanali, A., Hoglund, S., Pettersson, J., & Pickett, J. A. (2011). Repellent activity of catmint, *Nepeta cataria*, and iridoid nepetalactone isomers against Afro-tropical mosquitoes, ixodid ticks and red poultry mites. *Phytochemistry*, 72(1), 109–114. <https://doi.org/10.1016/j.phytochem.2010.09.016>
- Bozin, B., Mimica-Dukic, N., Simin, N., & Anackov, G. (2006). Characterization of the volatile composition of essential oils of some Lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *Journal of Agricultural and Food Chemistry*, 54(5), 1822–1828. <https://doi.org/10.1021/jf051922u>
- Bouyahya, A., Chamkhi, I., Benali, T., Guaouguau, F-E., Balahbib, A., El Omari, N., Taha, D., Belmehdi, O., Ghokhan, Z., & El Meniy, N. (2021). Traditional use, phytochemistry, toxicology, and pharmacology of *Origanum majorana* L. *Journal of Ethnopharmacology*, 265, 113318. <https://doi.org/10.1016/j.jep.2020.113318>
- Brahmi, F., Khodir, M., Mohamed, C., & Pierre, D. (2017). Chemical composition and biological activities of *Mentha* species. In H. El-Shemy (Ed.), *Aromatic and Medicinal Plants - Back to Nature*. London: IntechOpen.
- Bräuchler, C., Meimberg, H., & Heubl, G. (2010). Molecular phylogeny of Menthinae (Lamiaceae, Nepetoideae, Mentheae) – Taxonomy, biogeography and conflicts. *Molecular Phylogenetics and Evolution*, 55(2), 501–523. <https://doi.org/10.1016/j.ympev.2010.01.016>
- Bruno, W. J., Socci, N. D., & Halpern, A. L. (2000). Weighted neighbor joining: A likelihood-based approach to distance-based phylogeny reconstruction. *Molecular Biology and Evolution*, 17(1), 189–197. <https://doi.org/10.1093/oxfordjournals.molbev.a026231>
- Can, T. (2014). Introduction to bioinformatics. In M. Yousef & J. Allmer (Eds.), *miRNomics: MicroRNA Biology and Computational Analysis. Methods in Molecular Biology* (vol 1107). Humana Press.
- Cantino, P. D. (1992). Evidence for a polyphyletic origin of the labiateae. *Annals of the Missouri Botanical Garden*, 79(2), 361-379. <https://doi.org/10.2307/2399774>
- Cantino, P. D., & Sanders, R. W. (1986). Subfamilial classification of Labiateae. *Systematic Botany*, 11(1), 163. <https://doi.org/10.2307/2418955>
- Carmel, A., Musa-Lempel, N., Tsur, D., & Ziv-Ukelson, M. (2014). The worst case complexity of maximum parsimony. *Journal of Computational Biology*, 21(11), 799–808. <https://doi.org/10.1089/cmb.2014.0128>

- Caspermeyer J. (2018). MEGA software celebrates silver anniversary. *Molecular Biology and Evolution*, 35(6), 1558–1560. <https://doi.org/10.1093/molbev/msy098>
- CBOL Plant Working Group. (2009). A DNA barcode for land plants. *Proceedings of the National Academy of Sciences*, 106(31), 12794–12797. <https://doi.org/10.1073/pnas.0905845106>
- Chase, M. W., Salamin, N., Wilkinson, M., Dunwell, J. M., Kesanakurthi, R. P., Haider, N., & Savolainen, V. (2005). Land plants and DNA barcodes: short-term and long-term goals. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences*, 360(1462), 1889–1895. <https://doi.org/10.1098/rstb.2005.1720>
- Chatzou, M., Magis, C., Chang, J.-M., Kemen, C., Bussotti, G., Erb, I., & Notredame, C. (2015). Multiple sequence alignment modeling: methods and applications. *Briefings in Bioinformatics*, 17(6), 1009–1023. <https://doi.org/10.1093/bib/bbv099>
- Chenni, M., El Abed, D., Rakotomanomana, N., Fernandez, X., & Chemat, F. (2016). Comparative study of essential oils extracted from egyptian basil leaves (*Ocimum basilicum* L.) using hydro-distillation and solvent-free microwave extraction. *Molecules*, 21(1), 113. <https://doi.org/10.3390/molecules21010113>
- Chethana, G. S., Hari, V. K. R., & Gopinath, S. M. (2013). Review on *Clerodendrum inerme*. *Journal of Pharmaceutical and Scientific Innovation*, 2, 38–40.
- Chiang, K., Cheng, T., Kan, W., Wang, H.-Y., Li, J.-C., Cai, Y., Cheng, C.-H., Liu, Y., Chia Yu Chang, & Jiunn-Jye Chuu. (2024). *Orthosiphon aristatus* (Blume) Miq. extracts attenuate alzheimer-like pathology through anti-inflammatory, anti-oxidative, and β-amyloid inhibitory activities. *Journal of Ethnopharmacology*, 320, 117132–117132. <https://doi.org/10.1016/j.jep.2023.117132>
- Chua, L. S., Lau, C. H., Chew, C. Y., Ismail, N. I. M., & Soontorngun, N. (2018). Phytochemical profile of *Orthosiphon aristatus* extracts after storage: Rosmarinic acid and other caffeic acid derivatives. *Phytomedicine*, 39, 49–55. <https://doi.org/10.1016/j.phymed.2017.12.015>
- Cirlini, M., Mena, P., Tassotti, M., Herrlinger, K., Nieman, K., Dall'Asta, C., & Del Rio, D. (2016). Phenolic and volatile composition of a dry spearmint (*Mentha spicata* L.) extract. *Molecules*, 21(8), 1007. <https://doi.org/10.3390/molecules21081007>
- Czerwińska, M., Świerczewska, A., Woźniak, M., & Kiss, A. (2017). Bioassay-guided isolation of iridoids and phenylpropanoids from aerial parts of *Lamium album* and their anti-inflammatory activity in human neutrophils. *Planta Medica*, 83(12/13), 1011–1019. <https://doi.org/10.1055/s-0043-107031>

- Drew, B. T. (2020). Evolution, pollination biology, and species richness of *Salvia* (Lamiaceae). *International Journal of Plant Sciences*, 181(8), 767769. <https://doi.org/10.1086/710711>
- Drew, B. T., González-Gallegos, J. G., Xiang, C.-L., Kriebel, R., Drummond, C. P., Walker, J. B., & Sytsma, K. J. (2017). *Salvia* united: The greatest good for the greatest number. *Taxon*, 66(1), 133–145. <https://doi.org/10.12705/661.7>
- Drew, B. T., & Sytsma, K. J. (2012). Phylogenetics, biogeography, and staminal evolution in the tribe Mentheae (Lamiaceae). *American Journal of Botany*, 99(5), 933–953. <https://doi.org/10.3732/ajb.1100549>
- Edwards, A. F., & Cavalli-Sforza, L. L. (1963). The reconstruction of evolution. *Annals of Human Genetics*, 27, 105.
- EghbaliFeriz, S., Taleghani, A., & Tayarani-Najaran, Z. (2018). *Scutellaria*: Debates on the anticancer property. *Biomedicine & Pharmacotherapy*, 105, 1299–1310. <https://doi.org/10.1016/j.bioph.2018.06.107>
- Ekor M. (2014). The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*, 4, 177. <https://doi.org/10.3389/fphar.2013.00177>
- El Meniyi, N., Mrabti, H. N., El Omari, N., Bakili, A. E., Bakrim, S., Mekkaoui, M., Balahbib, A., Amiri-Ardekani, E., Ullah, R., Alqahtani, A. S., Shahat, A. A., & Bouyahya, A. (2022). Medicinal uses, phytochemistry, pharmacology, and toxicology of *Mentha spicata*. *Evidence-based Complementary and Alternative Medicine: eCAM*, 2022, 7990508. <https://doi.org/10.1155/2022/7990508>
- Erickson, D. L., & Driskell, A. C. (2012). Construction and analysis of phylogenetic trees using DNA barcode data. In W. Kress & D. Erickson (Eds.), *DNA Barcodes. Methods in Molecular Biology* (Vol. 858, pp. 395–408). New Jersey: Humana Press.
- Erickson, D. L., Spouge, J., Resch, A., Weigt, L. A., & Kress, W. J. (2008). DNA barcoding in land plants: Developing standards to quantify and maximize success. *Taxon*, 57(4), 1304–1316. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2749701/>
- Farris, J. S. (1966). Estimation of conservatism of characters by constancy within biological populations. *Evolution*, 20(4), 587–591. <https://doi.org/10.1111/j.1558-5646.1966.tb03389.x>
- Federici, S., Galimberti, A., Bartolucci, F., Bruni, I., De mattia, F., Cortis, P., & Labra, M. (2013). DNA barcoding to analyse taxonomically complex groups in plants: the case of *Thymus* (Lamiaceae). *Botanical Journal of the Linnean Society*, 171(4), 687–699. <https://doi.org/10.1111/boj.12034>

- Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, 39(4), 783–791. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>
- Fikry, S., Khalil, N., & Salama, O. (2019). Chemical profiling, biostatic and biocidal dynamics of *Origanum vulgare* L. essential oil. *AMB Express*, 9(1), 41. <https://doi.org/10.1186/s13568-019-0764-y>
- Filip, S. (2017). Basil (*Ocimum basilicum* L.) a source of valuable phytonutrients. *International Journal of Clinical Nutrition & Dietetics*, 3(1), 118. <https://doi.org/10.15344/2456-8171/2017/118>
- Flamini, G., Cioni, P. L., & Morelli, I. (2005). Composition of the essential oils and in vivo emission of volatiles of four *Lamium* species from Italy: *L. purpureum*, *L. hybridum*, *L. bifidum* and *L. amplexicaule*. *Food Chemistry*, 91, 63–68. <https://doi.org/10.1016/j.foodchem.2004.05.047>
- Fragman-Sapir, O. (t.t.). *Lamium album*. Plants of the World Online. Diakses dari: <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:448792-1/images>
- Francisco, O. L. (t.t.). *Clerodendrum bungei*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:861960-1/images>
- Fu, Z., Wang, H., Hu, X., Sun, Z., & Han, C. (2013). The pharmacological properties of *Salvia* essential oils. *Journal of Applied Pharmaceutical Science*, 3(07), 122–127. <https://doi.org/10.7324/JAPS.2013.3723>
- Fuchs, L. K., Holland, A. H., Ludlow, R. A., Coates, R. J., Armstrong, H., Pickett, J. A., Harwood, J. L., & Scofield, S. (2022). Genetic manipulation of biosynthetic pathways in mint. *Frontiers in Plant Science*, 13. <https://doi.org/10.3389/fpls.2022.928178>
- Gerbi, S. A. (1986). The evolution of eukaryotic ribosomal DNA. *Bio Systems*, 19(4), 247–258. [https://doi.org/10.1016/0303-2647\(86\)90001-8](https://doi.org/10.1016/0303-2647(86)90001-8)
- Govaerts, R. (2018). *Clerodendrum bungei*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:861960-1/images>
- Hać-Szymańczuk, E., Cegielka, A., Karkos, M., Gniewosz, M., & Piwowarek, K. (2018). Evaluation of antioxidant and antimicrobial activity of oregano (*Origanum vulgare* L.) preparations during storage of low-pressure mechanically separated meat (BAADER meat) from chickens. *Food Science and Biotechnology*, 28(2), 449–457. <https://doi.org/10.1007/s10068-018-0491-1>
- Hajibabaei, M., Singer, G. A. C., Hebert, P. D. N., & Hickey, D. A. (2007). DNA barcoding: How it complements taxonomy, molecular phylogenetics and population genetics. *Trends in Genetics*, 23(4), 167–172. <https://doi.org/10.1016/j.tig.2007.02.001>

- Halmschlag, C. B., Carneiro de Melo Maura, C., Brambach, F., Siregar, I. Z., & Gailing, O. (2022). Molecular and morphological survey of Lamiaceae species in converted landscapes in Sumatra. *PLOS ONE*, 17(12), e0277749–e0277749. <https://doi.org/10.1371/journal.pone.0277749>
- Harley, R. M., Atkins, S., Budantsev, A. L., Cantino, P. D., Conn, B. J., Grayer, R., Harley, M. M., De Kok, R., Krestovskaja, T., Morales, R., Paton, A. J., Ryding, O., & Upson, T. (2004). Labiateae. In Kadereit, J. W. (Ed.), *Families and Genera of Vascular Plants* (Vol. 7, pp. 167–275). Berlin: Springer.
- Hasebe, M., Wolf, P. G., Pryer, K. M., Ueda, K., Ito, M., Sano, R., Gastony, G. J., Yokoyama, J., Manhart, J. R., Murakami, N., Crane, E. H., Haufler, C. H., & Hauk, W. D. (1995). Fern phylogeny based on *rbcL* nucleotide sequences. *American Fern Journal*, 85(4), 134. <https://doi.org/10.2307/1547807>
- Hebert, P. D. N., Cywinska, A., Ball, S. L., & deWaard, J. R. (2003). Biological identifications through DNA barcodes. *Proceedings. Biological Sciences*, 270(1512), 313–321. <https://doi.org/10.1098/rspb.2002.2218>
- Hennig, W. (1999). *Phylogenetic systematics*. Illinois: University Of Illinois Press.
- Heylen, O. C. G., Debortoli, N., Marescaux, J., & Olofsson, J. K. (2021). A revised phylogeny of the *Mentha spicata* clade reveals cryptic species. *Plants*, 10(4), 819. <https://doi.org/10.3390/plants10040819>
- Hillis, D. M., & Bull, J. J. (1993). An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Systematic Biology*, 42(2), 182. <https://doi.org/10.2307/2992540>
- Hilu, K. W., & Liang, H. gping. (1997). The *matK* gene: Sequence variation and application in plant systematics. *American Journal of Botany*, 84(6), 830–839. <https://doi.org/10.2307/2445819>
- Ho, V. T., Tran, T. K. P., Vu, T. T. T., & Widiarsih, S. (2021). Comparison of *matK* and *rbcL* DNA barcodes for genetic classification of jewel orchid accessions in Vietnam. *Journal of Genetic Engineering and Biotechnology*, 19(1), 93. <https://doi.org/10.1186/s43141-021-00188-1>
- Hollingsworth , M., Clark, A. A., Forrest, L. L., Richardson, J., Pennington, R. T., Long, D. G., Cowan, R., Chase, M., Gaudeul, M., & Hollingsworth , P. M. (2009). Selecting barcoding loci for plants: Evaluation of seven candidate loci with species-level sampling in three divergent groups of land plants. *Molecular Ecology Resources*, 9(2), 439–457. <https://doi.org/10.1111/j.1755-0998.2008.02439.x>
- Hudz, N., Kobylinska, L., Pokajewicz, K., Horčinová Sedláčková, V., Fedin, R., Voloshyn, M., Myskiv, I., Brindza, J., Wieczorek, P. P., & Lipok, J. (2023). *Mentha piperita*: Essential oil and extracts, their biological activities, and perspectives on the development of new medicinal and cosmetic products. *Molecules*, 28(21), 7444. <https://doi.org/10.3390/molecules28217444>

- Huxley, J. (1957). The three types of evolutionary process. *Nature*, 180(4584), 454–455. <https://doi.org/10.1038/180454a0>
- Huynh, D. L., Sharma, N., Kumar Singh, A., Singh Sodhi, S., ZHANG, J.-J., Mongre, R. K., Ghosh, M., Kim, N., Ho Park, Y., & Kee Jeong, D. (2017). Anti-tumor activity of wogonin, an extract from *Scutellaria baicalensis*, through regulating different signaling pathways. *Chinese Journal of Natural Medicines*, 15(1), 15–40. [https://doi.org/10.1016/s1875-5364\(17\)30005-5](https://doi.org/10.1016/s1875-5364(17)30005-5)
- Hyun, T. K., Kim, H. C., & Kim, J. S. (2014). Antioxidant and antidiabetic activity of *Thymus quinquecostatus* Celak. *Industrial Crops and Products*, 52, 611–616. <https://doi.org/10.1016/j.indcrop.2013.11.039>
- India Biodiversity Portal.* (2014). *Orthosiphon aristatus* (Blume) Miq. <https://indiabiodiversity.org/observation/show/377543>
- Isaac, S., & Michael, W. B. (1979). *Handbook in Research and Evaluation*. Edits Publishers.
- Jabeen, A., Guo, B., Abbasi, B. H., Shinwari, Z. K., & Mahmood, T. (2012). Phylogenetics of selected *Mentha* species on the basis of *rps8*, *rps11* and *rps14* chloroplast genes. *Journal of Medicinal Plants Research*, 6(1), 30–36. <https://doi.org/10.5897/jmpr11.658>
- Jahani, R., Khaledyan, D., Jahani, A., Jamshidi, E., Kamalinejad, M., Khoramjouy, M., & Faizi, M. (2019). Evaluation and comparison of the antidepressant-like activity of *Artemisia dracunculus* and *Stachys lavandulifolia* ethanolic extracts: an *in vivo* study. *Research in Pharmaceutical Sciences*, 14(6), 544. <https://doi.org/10.4103/1735-5362.272563>
- Jiang, D., Zhao, Z., Zhang, T., Zhong, W., Liu, C., Yuan, Q., & Huang, L. (2017). The chloroplast genome sequence of *Scutellaria baicalensis* provides insight into intraspecific and interspecific chloroplast genome diversity in *Scutellaria*. *Genes*, 8(9), 227. <https://doi.org/10.3390/genes8090227>
- Jin, G., Nakhleh, L., Snir, S., & Tuller, T. (2007). Inferring phylogenetic networks by the maximum parsimony criterion: A case study. *Molecular Biology and Evolution*, 24(1), 324–337. <https://doi.org/10.1093/molbev/msl163>
- Jirovetz, L., Buchbauer, G., Shafi, M. P., & Kaniampady, M. M. (2003). Chemotaxonomical analysis of the essential oil aroma compounds of four different *Ocimum* species from southern India. *European Food Research & Technology*, 217(2), 120–124. <https://doi.org/10.1007/s00217-003-0708-1>
- Jobes, D. V., & Thien, L. B. (1997). A conserved motif in the 5.8s ribosomal RNA (*rRNA*) gene is a useful diagnostic marker for plant internal transcribed spacer (*ITS*) sequences. *Plant Molecular Biology Reporter*, 15(4), 326–334. <https://doi.org/10.1023/a:1007462330699>

- Jones, W., & Kinghorn, A. (2008). Biologically active natural products of the genus *Callicarpa*. *Current Bioactive Compounds*, 4(1), 15–32. <https://doi.org/10.2174/157340708784533393>
- Kang, C.-H., Molagoda, I. M. N., Choi, Y. H., Park, C., Moon, D.-O., & Kim, G.-Y. (2018). Apigenin promotes TRAIL-mediated apoptosis regardless of ROS generation. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 111, 623–630. <https://doi.org/10.1016/j.fct.2017.12.018>
- Kannan, L., & Wheeler, W. C. (2012). Maximum parsimony on phylogenetic networks. *Algorithms for Molecular Biology*, 7(1). <https://doi.org/10.1186/1748-7188-7-9>
- Kapli, P., Flouri, T., & Telford, M. J. (2021). Systematic errors in phylogenetic trees. *Current Biology*, 31(2), R59–R64. <https://doi.org/10.1016/j.cub.2020.11.043>
- Karioti, A., Bolognesi, L., Vincieri, F. F., & Bilia, A. R. (2010). Analysis of the constituents of aqueous preparations of *Stachys recta* by HPLC–DAD and HPLC–ESI-MS. *Journal of Pharmaceutical and Biomedical Analysis*, 53(1), 15–23. <https://doi.org/10.1016/j.jpba.2010.03.002>
- Kellogg, E., & Juliano, N. (1997). The structure and function of RuBisCO and their implications for systematic studies. *American Journal of Botany*, 84(3), 413–428.
- Keng, C. L., & Siong, L. P. (2006). Morphological similarities and differences between the two varieties of cat's whiskers (*Orthosiphon stamineus* Benth.) grown in Malaysia. *International Journal of Botany*, 2(1), 1–6. <https://doi.org/10.3923/ijb.2006.1.6>
- Kim, M., Moon, J.-C., Kim, S., & Sowndhararajan, K. (2020). Morphological, chemical, and genetic characteristics of korean native thyme *bak-ri-hyang* (*Thymus quinquecostatus* Celak.). *Antibiotics*, 9(6), 289. <https://doi.org/10.3390/antibiotics9060289>
- Klingenberg, C. P., & Gidaszewski, N. A. (2010). Testing and quantifying phylogenetic signals and homoplasy in morphometric data. *Systematic Biology*, 59(3), 245–261. <https://doi.org/10.1093/sysbio/syp106>
- Kress, W. J., & Erickson, D. L. (2012). *DNA barcodes: Methods and protocols* (Vol. 858). Humana Press.
- Kress, W. J., Wurdack, K. J., Zimmer, E. A., Weigt, L. A., & Janzen, D. H. (2005). Use of DNA barcodes to identify flowering plants. *Proceedings of the National Academy of Sciences of the United States of America*, 102(23), 8369–8374. <https://doi.org/10.1073/pnas.0503123102>

- Krogsgaard, E. (2020a). *Origanum vulgare*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:453395-1/images>
- Krogsgaard, E. (2020b). *Stachys sylvatica*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:460020-1/images>
- Krogsgaard, E. (2021). *Thymus vulgaris*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:461765-1/images>
- Lahaye, R., van der Bank, M., Bogarin, D., Warner, J., Pupulin, F., Gigot, G., Maurin, O., Duthoit, S., Barraclough, T. G., & Savolainen, V. (2008). DNA barcoding the floras of biodiversity hotspots. *Proceedings of the National Academy of Sciences*, 105(8), 2923–2928. <https://doi.org/10.1073/pnas.0709936105>
- Laitinen, R. A. E., & Nikoloski, Z. (2018). Genetic basis of plasticity in plants. *Journal of Experimental Botany*, 70(3), 739–745. <https://doi.org/10.1093/jxb/ery404>
- Leyva-López, N., Gutiérrez-Grijalva, E. P., Vazquez-Olivo, G., & Heredia, J. B. (2017). Essential oils of oregano: biological activity beyond their antimicrobial properties. *Molecules*, 22(6), 989. <https://doi.org/10.3390/molecules22060989>
- Li, B., Cantino, P. D., Olmstead, R. G., Bramley, G. L. C., Xiang, C.-L., Ma, Z.-H., Tan, Y.-H., & Zhang, D.-X. (2016). A large-scale chloroplast phylogeny of the Lamiaceae sheds new light on its subfamilial classification. *Scientific Reports*, 6(1), 34343. <https://doi.org/10.1038/srep34343>
- Li, B., & Olmstead, R. G. (2017). Two new subfamilies in Lamiaceae. *Phytotaxa*, 313(2), 222–226. <https://doi.org/10.111646/phytotaxa.313.2.9>
- Li, B., Xu, W. X., Tu, T., Wang, Z., Olmstead, R., Peng, H., Francisco-Ortega, J., Cantino, P. D., & Zhang, D. (2012). Phylogenetic position of *Wenchengia* (Lamiaceae): A taxonomically enigmatic and critically endangered genus. *Taxon*, 61(2), 392–401. <http://dx.doi.org/10.1002/tax.612010>
- Li, P., Qi, Z.-C., Liu, L.-X., Ohi-Toma, T., Lee, J., Hsieh, T.-H., Fu, C.-X., Cameron, K. M., & Qiu, Y.-X. (2017). Molecular phylogenetics and biogeography of the mint tribe Elsholtzieae (Nepetoideae, Lamiaceae), with an emphasis on its diversification in East Asia. *Scientific Reports*, 7(1), 2057. <https://doi.org/10.1038/s41598-017-02157-6>
- Li, Z., Duan, B., Zhou, Z., Fang, H., Yang, M., Xia, C., Zhou, Y., & Wang, J. (2024). Comparative analysis of medicinal plants *Scutellaria baicalensis* and common adulterants based on chloroplast genome sequencing. *BMC Genomics*, 25(1). <https://doi.org/10.1186/s12864-023-09920-2>
- Lindqvist, C., & Albert, V. A. (2002). Origin of the Hawaiian endemic mints within North American *Stachys* (Lamiaceae). *American Journal of Botany*, 89(10), 1709–1724. <https://doi.org/10.3732/ajb.89.10.1709>

- Liu, B. (t.t.-a). *Mentha canadensis*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:30016176-2/images>
- Liu, B. (t.t.-b). *Callicarpa dichotoma*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:60475856-2/images>
- Liu, B. (t.t.-c). *Lamium album*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:448792-1/images>
- Liu, B. (t.t.-d). *Scutellaria baicalensis*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:458155-1/images>
- Liu, Y., Guo, S., Bei, J., & Wu, W. (2022). The complete chloroplast genome of *Callicarpa macrophylla* Vahl.. *Mitochondrial DNA. Part B. Resources*, 7(10), 1845–1847. <https://doi.org/10.1080/23802359.2022.2134749>
- Lombrea, A., Antal, D., Ardelean, F., Avram, S., Pavel, I. Z., Vlaia, L., Mut, A.-M., Diaconeasa, Z., Dehelean, C. A., Soica, C., & Danciu, C. (2020). A recent insight regarding the phytochemistry and bioactivity of *Origanum vulgare* L. Essential Oil. *International Journal of Molecular Sciences*, 21(24), E9653. <https://doi.org/10.3390/ijms21249653>
- Lukas, B., & Novak, J. (2013). The complete chloroplast genome of *Origanum vulgare* L. (Lamiaceae). *Gene*, 528(2), 163–169. <https://doi.org/10.1016/j.gene.2013.07.026>
- Lukas, B., Samuel, R., Mader, E., Can, H., Duman, H., & Novak, J. (2013). Complex evolutionary relationships in *Origanum* section *Majorana* (Lamiaceae). *Botanical Journal of the Linnean Society*, 171(4), 667–686. <https://doi.org/10.1111/boj.12022>
- Lumbsch, H. T., & Leavitt, S. D. (2011). Goodbye morphology? A paradigm shift in the delimitation of species in lichenized fungi. *Fungal Diversity*, 50(1), 59–72. <https://doi.org/10.1007/s13225-011-0123-z>
- Mabberley, D. J. (2008). *Mabberley's Plant-book* (3rd ed.). Cambridge University Press.
- Mahajan, N., Rawal, S., Verma, M., Poddar, M., & Alok, S. (2013). A phytopharmacological overview on *Ocimum* species with special emphasis on *Ocimum sanctum*. *Biomedicine & Preventive Nutrition*, 3(2), 185–192. <https://doi.org/10.1016/j.bionut.2012.08.002>
- Mahomoodally, F., Suroowan, S., & Sreekeessoon, U. (2018). Adverse reactions of herbal medicine—A quantitative assessment of severity in Mauritius. *Journal of Herbal Medicine*, 12, 49-65. <https://doi.org/10.1016/j.hermed.2018.01.006>

- Marieschi, M., Torelli, A., Bianchi, A., & Bruni, R. (2011). Detecting *Satureja montana* L. and *Origanum majorana* L. by means of SCAR-PCR in commercial samples of Mediterranean oregano. *Food Control*, 22(3-4), 542–548. <https://doi.org/10.1016/j.foodcont.2010.10.001>
- Masters, J. C., & Pozzi, L. (2017). Phylogenetic inference. In A. Fuentes (Ed.), *The International Encyclopedia of Primatology*. New Jersey: John Wiley & Sons, Inc.
- Masyita, A., Mustika Sari, R., Dwi Astuti, A., Yasir, B., Rahma Rumata, N., Emran, T. B., Nainu, F., & Simal-Gandara, J. (2022). Terpenes and terpenoids as main bioactive compounds of essential oils, their roles in human health and potential application as natural food preservatives. *Food Chemistry: X*, 13, 100217. <https://doi.org/10.1016/j.fochx.2022.100217>
- McCune, A. R., & Schimenti, J. C. (2012). Using genetic networks and homology to understand the evolution of phenotypic traits. *Current Genomics*, 13(1), 74–84. <https://doi.org/10.2174/138920212799034785>
- Mennema, J. (1989). *A taxonomic revision of Lamium (Lamiaceae)*. Leiden: BRILL.
- Mickevich, M. F., & Lipscomb, D. (1991). Parsimony and the choice between different transformations for the same character set. *Cladistics*, 7(2), 111–139. <https://doi.org/10.1111/j.1096-0031.1991.tb00028.x>
- Mint Evolutionary Genomics Consortium. (2018). Phylogenomic mining of the mints reveals multiple mechanisms contributing to the evolution of chemical diversity in Lamiaceae. *Molecular Plant*, 11(8), 1084–1096. <https://doi.org/10.1016/j.molp.2018.06.002>
- Mishra, P., Kumar, A., Nagireddy, A., Mani, D. N., Shukla, A. K., Tiwari, R., & Sundaresan, V. (2016). DNA barcoding: an efficient tool to overcome authentication challenges in the herbal market. *Plant Biotechnology Journal*, 14(1), 8–21. <https://doi.org/10.1111/pbi.12419>
- Moon, H.-K., Smets, E., & Huysmans, S. (2010). Phylogeny of tribe Mentheae (Lamiaceae): The story of molecules and micromorphological characters. *TAXON*, 59(4), 1065–1076. <https://doi.org/10.1002/tax.594007>
- Munjal, G., Hanmandlu, M., & Srivastava, S. (2019). phylogenetics algorithms and applications. *Advances in Intelligent Systems and Computing*, 904, 187–194. https://doi.org/10.1007/978-981-13-5934-7_17
- Naghibi, F., Mosaddegh, M., Motamed, S. M., & Ghorbani, A. (2005). Labiate family in folk medicine in Iran: from ethnobotany to pharmacology. *Iranian Journal of Pharmaceutical Research*, 4(2), 63–79. <https://doi.org/10.22037/ijpr.2010.619>

- Nazar, N., Howard, C., Slater, A., & Sgamma, T. (2022). Challenges in medicinal and aromatic plants DNA barcoding—Lessons from the Lamiaceae. *Plants*, 11(1), 137. <https://doi.org/10.3390/plants11010137>
- Nickavar, B., Alinaghi, A., & Kamalinejad, M. (2008). Evaluation of the antioxidant properties of five *Mentha* Species. *Iranian Journal of Pharmaceutical Research*, 7(3), 203–209. <https://doi.org/10.22037/ijpr.2010.766>
- Niu, Y., Qin, Q., Dong, Y., Wang, X., Zhang, S., & Mu, Z. (2023). Chloroplast genome structure and phylogenetic analysis of 13 Lamiaceae plants in Tibet. *Frontiers in Bioscience (Landmark Edition)*, 28(6), 110. <https://doi.org/10.31083/j.fbl2806110>
- Nurhasanah, Sundari, & Papuangan, N. (2019). Amplification and analysis of *rbcL* gene (*ribulose-1,5-bisphosphate carboxylase*) of clove in Ternate Island. *IOP Conf. Ser.: Earth Environ. Sci.*, 276. <https://doi.org/10.1088/1755-1315%2F276%2F1%2F012061>
- Oakeley, H. (t.t.). *Scutellaria baicalensis*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:458155-1/images>
- Ojha, K. K., Mishra, S., & Singh, V. K. (2022). Computational molecular phylogeny: concepts and applications. In D. B. Singh & R. K. Pathak (Eds.), *Bioinformatics: Methods and Applications* (pp. 67–89). Academic Press.
- Panda, S. P., Van Puyvelde, L., Mukazayire, M.-J., & Gazim, Z. C. (2022). Editorial: Ethnopharmacology of the Lamiaceae: Opportunities and challenges for developing new medicines. *Frontiers in Pharmacology*, 13, 961486. <https://doi.org/10.3389/fphar.2022.961486>
- Pathak, R. K., Singh, D. B., & Singh, R. (2022). Introduction to basics of bioinformatics. In D. B. Singh & R. K. Pathak (Eds.), *Bioinformatics Methods and Applications* (pp. 1–15). Academic Press.
- Paton, A. (1990). A global taxonomic investigation of *Scutellaria* (Labiatae). *Kew Bulletin*, 45(3), 399. <https://doi.org/10.2307/4110512>
- Paton, A. J., Springate, D., Suddee, S., Otieno, D., Grayer, R. J., Harley, M. M., Willis, F., Simmonds, M. S. J., Powell, M. P., & Savolainen, V. (2004). Phylogeny and evolution of basils and allies (Ocimeae, Labiatae) based on three plastid DNA regions. *Molecular Phylogenetics and Evolution*, 31(1), 277–299. <https://doi.org/10.1016/j.ymp.2003.08.002>
- Prokhozhy, V. (t.t.-a). *Ocimum basilicum*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:452874-1/images>
- Prokhozhy, V. (t.t.-b). *Mentha arvensis*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:30016176-2/images>
- Chayra Endessa, 2024
ANALISIS FILOGENETIK LAMIACEAE BERDASARKAN BARKODE ITS, matK, DAN rbcL SECARA IN SILICO
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- Prokhozhy, V. (t.t.-c). *Salvia splendens*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:60466516-2/images>
- Ranganathan, P., & Aggarwal, R. (2018). Study designs: Part 1 - an overview and classification. *Perspectives in Clinical Research*, 9(4), 184–186. https://doi.org/10.4103/picr.PICR_124_18
- Rattray, R. D., & Van Wyk, B.-E. (2021). The botanical, chemical and ethnobotanical diversity of Southern African Lamiaceae. *Molecules*, 26(12), 3712. <https://doi.org/10.3390/molecules26123712>
- Ravikumar, K., Begum, S. N., Ved, D. K., Bhatt, J. R., & Goraya, G. S. (2018). *Compendium of traded medicinal plants*. Bengaluru, India: Foundation for Revitalization of Local Health Traditions.
- Rokas, A., & Carroll, S. B. (2005). More genes or more taxa? The relative contribution of gene number and taxon number to phylogenetic accuracy. *Molecular Biology and Evolution*, 22(5), 1337–1344. <https://doi.org/10.1093/molbev/msi121>
- Rose, J. P., Kriebel, R., Kahan, L., DiNicola, A., González-Gallegos, J. G., Celep, F., Lemmon, E. M., Lemmon, A. R., Sytsma, K. J., & Drew, B. T. (2021). Sage insights into the phylogeny of *Salvia*: Dealing with sources of discordance within and across genomes. *Frontiers in Plant Science*, 12. <https://doi.org/10.3389/fpls.2021.767478>
- Roy, S., Tyagi, A., Shukla, V., Kumar, A., Singh, U. M., Chaudhary, L. B., Datt, B., Bag, S. K., Singh, P. K., Nair, N. K., Husain, T., & Tuli, R. (2010). Universal plant DNA barcode loci may not work in complex groups: A case study with Indian berberis species. *PLoS ONE*, 5(10), e13674. <https://doi.org/10.1371/journal.pone.0013674>
- Runyoro, D., Ngassapa, O., Vagionas, K., Aligiannis, N., Graikou, K., & Chinou, I. (2010). Chemical composition and antimicrobial activity of the essential oils of four *Ocimum* species growing in Tanzania. *Food Chemistry*, 119(1), 311–316. <https://doi.org/10.1016/j.foodchem.2009.06.028>
- Russo, C. A. de M., & Selvatti, A. P. (2018). Bootstrap and rogue identification tests for phylogenetic analyses. *Molecular Biology and Evolution*, 35(9), 2327–2333. <https://doi.org/10.1093/molbev/msy118>
- Ryding, O. (2007). Amount of calyx fibres in Lamiaceae, relation to calyx structure, phylogeny and ecology. *Plant Systematics and Evolution*, 268(1-4), 45–58. <https://doi.org/10.1007/s00606-007-0537-y>
- Sadeghi, H., Mansourian, M., Panahi Kokhdan, E., Salehpour, Z., Sadati, I., Abbaszadeh-Goudarzi, K., Asfaram, A., & Dousttimotagh, A. H. (2020). Antioxidant and protective effect of *Stachys pilifera* Benth against

- nephrotoxicity induced by cisplatin in rats. *Journal of Food Biochemistry*, 44(5), e13190. <https://doi.org/10.1111/jfbc.13190>
- Safikhani, K., Jamzad, Z., & Saeidi, H. (2018). Phylogenetic relationships in Iranian *Scutellaria* (Lamiaceae) based on nuclear ribosomal ITS and chloroplast trnLF DNA data. *Plant Systematics and Evolution*, 304(9), 1077–1089. <https://doi.org/10.1007/s00606-018-1533-0>
- Saha, O., Hossain, Md. S., & Rahaman, Md. M. (2020). Genomic exploration light on multiple origin with potential parsimony-informative sites of the severe acute respiratory syndrome coronavirus 2 in Bangladesh. *Gene Reports*, 21, 100951. <https://doi.org/10.1016/j.genrep.2020.100951>
- Salehi, B., Armstrong, L., Rescigno, A., Yeskaliyeva, B., Seitimova, G., Beyatli, A., Sharmin, J., Mahomoodally, M. F., Sharopov, F., Durazzo, A., Lucarini, M., Santini, A., Abenavoli, L., Capasso, R., & Sharifi-Rad, J. (2019). *Lamium* Plants—A comprehensive review on health benefits and biological activities. *Molecules*, 24(10), 1913–1913. <https://doi.org/10.3390/molecules24101913>
- Salimov, R. A., Parolly, G., & Borsch, T. (2021). Overall phylogenetic relationships of *Scutellaria* (Lamiaceae) shed light on the origin of the predominantly Caucasian and Irano-Turanian *S. orientalis* group. *Willdenowia*, 51(3), 395–427. <https://doi.org/10.3372/wi.51.51307>
- Salmaki, Y., Heubl, G., & Weigend, M. (2019). Towards a new classification of tribe Stachydeae (Lamiaceae): naming clades using molecular evidence. *Botanical Journal of the Linnean Society*, 190(4), 345–358. <https://doi.org/10.1093/botlinnean/boz021>
- Salmaki, Y., Zarre, S., Ryding, O., Lindqvist, C., Bräuchler, C., Heubl, G., Barber, J., & Bendiksby, M. (2013). Molecular phylogeny of tribe Stachydeae (Lamiaceae subfamily Lamioideae). *Molecular Phylogenetics and Evolution*, 69(3), 535–551. <https://doi.org/10.1016/j.ympev.2013.07.024>
- Saric-Kundalic B., Fialova S., Dobes C., Olzant S., Tekelova D., Granca D., Reznicek G., Saukel J. (2009). Multivariate numerical taxonomy of *Mentha* species, hybrids, varieties and cultivars. *Scientia Pharmaceutica*, 77(4), 851–8766. <https://doi.org/10.3797/scipharm.0905-10>
- Satthaphorn, J., Paton, A. J., Zuntini, A. R., Cowan, R. S., & Charan Leeratiwong. (2023). Phylogeny and infrageneric classification of *Clerodendrum* (Lamiaceae). *Botanical Journal of the Linnean Society*, 204(2), 103–136. <https://doi.org/10.1093/botlinnean/boad045>
- Savolainen, V., Cowan, R. S., Vogler, A. P., Roderick, G. K., & Lane, R. (2005). Towards writing the encyclopaedia of life: an introduction to DNA barcoding. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 360(1462), 1805–1811. <https://doi.org/10.1098/rstb.2005.1730>

- Scheen, A., Bendiksby, M., Ryding, O., Mathiesen, C., Albert, V. A., & Lindqvist, C. (2010). Molecular phylogenetics, character evolution, and suprageneric classification of Lamioideae (Lamiaceae). *Annals of the Missouri Botanical Garden*, 97(2), 191–217. <https://doi.org/10.3417/2007174>
- Schoch, C. L., Seifert, K. A., Huhndorf, S., & Schindel, D. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for *Fungi*. *Proceedings of the National Academy of Sciences*, 109(16), 6241–6246. <https://doi.org/10.1073/pnas.1117018109>
- Seethapathy, G. S., Balasubramani, S. P., & Venkatasubramanian, P. (2014). nrDNA ITS sequence based SCAR marker to authenticate *Aconitum heterophyllum* and *Cyperus rotundus* in Ayurvedic raw drug source and prepared herbal products. *Food Chemistry*, 145, 1015–1020. <https://doi.org/10.1016/j.foodchem.2013.09.027>
- Selvaraj, D., Sarma, R. K., & Sathishkumar, R. (2008). Phylogenetic analysis of chloroplast *matK* gene from Zingiberaceae for plant DNA barcoding. *selvarajBioinformation*, 3(1), 24–27. <https://doi.org/10.6026/97320630003024>
- Shah, M., Mubin, S., Hassan, S. S. ul, Tagde, P., Ullah, O., Rahman, Md. H., Al-Harrasi, A., Rehman, N. U., & Murad, W. (2022a). Phytochemical profiling and bio-potentiality of genus *Scutellaria*: Biomedical approach. *Biomolecules*, 12(7), 936. <https://doi.org/10.3390/biom12070936>
- Shah, M., Rahman, H., Khan, A., Bibi, S., Ullah, O., Ullah, S., Ur Rehman, N., Murad, W., & Al-Harrasi, A. (2022b). Identification of α-glucosidase inhibitors from *Scutellaria edelbergii*: ESI-LC-MS and computational approach. *Molecules*, 27(4), 1322. <https://doi.org/10.3390/molecules27041322>
- Shahrajabian, M. H., Sun, W., & Cheng, Q. (2020). Chemical components and pharmacological benefits of Basil (*Ocimum basilicum*): A review. *International Journal of Food Properties*, 23(1), 1961–1970. <https://doi.org/10.1080/10942912.2020.1828456>
- Sharifi-Rad, M., Ozcelik, B., Altin, G., Daskaya-Dikmen, C., Martorell, M., Ramirez-Alarcon, K., Alarcon-Zapata, P., Morais-Braga, M. F. B., Carneiro, J. N. P., Leal, A., Coutinho, H. D. M., Gyawali, R., Tahergorabi, R., Ibrahim, S. A., Sahrifi-Rad, R., Sharopov, F., Salehi, B., Contreras, M. D., Segura-Carretero, A., ... Sharifi-Rad, J. (2018). *Salvia* spp. plants-from farm to food applications and phytopharmacotherapy. *Trends in Food Science & Technology*, 80, 242–263. <http://eprints.zbmu.ac.ir/3812/>
- Shen, J., Li, P., Liu, S., Liu, Q., Li, Y., Sun, Y., He, C., & Xiao, P. (2020). Traditional uses, clinical studies, and ten-years research progress in phytochemistry and pharmacology of the genus *Scutellaria*. *Journal of Ethnopharmacology*, 265, 113198. <https://doi.org/10.1016/j.jep.2020.113198>

- Sheremetev, I. (2011). *Salvia splendens*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:60466516-2/images>
- Sheremetev, I. (2013). *Stachys sylvatica*. Plants of the World Online. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:460020-1/images>
- Shrivastava , N., & Patel , T. (2007). *Clerodendrum* and heathcare: An overview. *Medicinal and Aromatic Plant Science and Biotechnology*, 1, 209–223.
- Simpson, M. G. (2010). *Plant Systematics* (2nd ed.). Cambridge: Academic Press.
- Soilhi, Z., Rhimi, A., Heuskin, S., Fauconnier, M. L., & Mekki, M. (2019). Essential oil chemical diversity of Tunisian *Mentha* spp. collection. *Industrial Crops and Products*, 131, 330-340. <https://doi.org/10.1016/j.indcrop.2019.01.041>
- Sonmezdag, A., Kelebek, H., & Sellı, S. (2017). Identification of aroma compounds of Lamiaceae Species in Turkey using the purge and trap technique. *Foods*, 6(2), 10. <https://doi.org/10.3390/foods6020010>
- Srivastava, A., Chaurasia, J., Khan, R., Dhand, C., & Verma, S. (2020). role of medicinal plants of traditional use in recuperating devastating COVID-19 situation. *Medicinal and Aromatic Plants*, 9(5), 1–16. <https://doi.org/10.35248/2167-0412.20.9.359>
- Stamatakis, A. (2015). Using RAxML to infer phylogenies. *Current Protocols in Bioinformatics*, 51(1), 1–14. <https://doi.org/10.1002/0471250953.bi0614s51>
- Stern, K., Bidlack, J., & Jansky, S. (2000). *Stern's Introductory Plant Biology*. McGraw Hill .
- Su, Y., Li, B., Liang, W., Wen, H., & Wei, W. (2024). The complete chloroplast genome of *Orthosiphon aristatus* (Blume) Miq. (Lamiaceae). *Mitochondrial DNA. Part B, Resources*, 9(1), 79–82. <https://doi.org/10.1080/23802359.2023.2301012>
- Sudarmono, Kim, S. Y., & Paik , J. H. (2020). Contradictory between morphology and phylogenetic trees of *Orthosiphon* spp. (Lamiaceae) from Indonesia. *IOP Conference Series*, 457(1), 012030–012030. <https://doi.org/10.1088/1755-1315/457/1/012030>
- Tamura, K., Stecher, G., & Kumar, S. (2021). MEGA11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology and Evolution*, 38(7), 3022–3027. <https://doi.org/10.1093/molbev/msab120>
- Tarhan, S., Telci, I., Tuncay, M. T., & Polatci, H. (2010). Product quality and energy consumption when drying peppermint by rotary drum dryer. *Industrial Crops and Products*, 32(3), 420-427. <https://doi.org/10.1016/j.indcrop.2010.06.003>

- Tassy, P., & Fischer, M. S. (2021). “Cladus” and clade: A taxonomic odyssey. *Theory in Biosciences*, 140(1), 77–85. <https://doi.org/10.1007/s12064-020-00326-2>
- Thakur, V. V., Tripathi, N., & Tiwari, S. (2021). DNA barcoding of some medicinally important plant species of Lamiaceae family in India. *Molecular Biology Reports*, 48(4), 3097–3106. <https://doi.org/10.1007/s11033-021-06356-3>
- The Angiosperm Phylogeny Group IV. (2016). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Botanical Journal of the Linnean Society*, 181(1), 1–20. <https://doi.org/10.1111/boj.12385>
- Tomas-Barberan, F. A., & Gil, M. L. (1992). *Chemistry and natural distribution of flavonoids in the Labiateae*. London: Royal Botanical Gardens.
- Tomou, E.-M., Barda, C., & Skaltsa, H. (2020). Genus *Stachys*: A review of traditional uses, phytochemistry and bioactivity. *Medicines*, 7(10), 63. <https://doi.org/10.3390/medicines7100063>
- Tu, Y., Sun, L., Guo, M., & Chen, W. (2013). The medicinal uses of *Callicarpa* L. in traditional Chinese medicine: An ethnopharmacological, phytochemical and pharmacological review. *Journal of Ethnopharmacology*, 146(2), 465–481. <https://doi.org/10.1016/j.jep.2012.12.051>
- Tundis, R., Peruzzi, L., & Menichini, F. (2014). Phytochemical and biological studies of *Stachys* species in relation to chemotaxonomy: A review. *Phytochemistry*, 102, 7–39. <https://doi.org/10.1016/j.phytochem.2014.01.023>
- Upton, R., David, B., Gafner, S., & Glasl, S. (2020). Botanical ingredient identification and quality assessment: strengths and limitations of analytical techniques. *Phytochemistry Reviews*, 19(5), 1157–1177. <https://doi.org/10.1007/s11101-019-09625-z>
- Upton, R., & DAyu, R. H. (2012). Skullcap *Scutellaria lateriflora* L.: An American nervine. *Journal of Herbal Medicine*, 2(3), 76–96. <https://doi.org/10.1016/j.hermed.2012.06.004>
- Uritu, C. M., Mihai, C. T., Stanciu, G.-D., Dodi, G., Alexa-Stratulat, T., Luca, A., Leon-Constantin, M.-M., Stefanescu, R., Bild, V., Melnic, S., & Tamba, B. I. (2018). Medicinal plants of the family Lamiaceae in pain therapy: A review. *Pain Research and Management*, 2018, 1–44. <https://doi.org/10.1155/2018/7801543>
- Van Iersel, L., Jones, M., & Scornavacca, C. (2017). Improved maximum parsimony models for phylogenetic networks. *Systematic Biology*, 67(3), 518–542. <https://doi.org/10.1093/sysbio/syx094>

- Vineesh, S., Balaji, R., Tanuja, & Parani, M. (2023). The complete chloroplast genome of *Ocimum americanum* Linnaeus 1755 and phylogenetic analysis among the Lamiaceae family. *Mitochondrial DNA. Part B. Resources*, 8(10), 1077–1081. <https://doi.org/10.1080/23802359.2023.2264545>
- Vundać, V. B. (2019). Taxonomical and phytochemical characterisation of 10 *Stachys* taxa recorded in the Balkan Peninsula Flora: A review. *Plants*, 8(2), 32. <https://doi.org/10.3390/plants8020032>
- Walker, J. B., & Sytsma, K. J. (2007). Staminal evolution in the genus *Salvia* (Lamiaceae): Molecular phylogenetic evidence for multiple origins of the staminal lever. *Annals of Botany*, 100(2), 375–391. <https://doi.org/10.1093/aob/mcl176>
- Wang, J. (2022). Editorial: Methods and applications in molecular phylogenetics. *Frontiers in Genetics*, 13. <https://doi.org/10.3389/fgene.2022.923409>
- Wang, J.-H., Luan, F., He, X.-D., Wang, Y., & Li, M.-X. (2018). Traditional uses and pharmacological properties of *Clerodendrum* phytochemicals. *Journal of Traditional and Complementary Medicine*, 8(1), 24–38. <https://doi.org/10.1016/j.jtcme.2017.04.001>
- Wang, Q., Wang, J., Li, N., Liu, J., Zhou, J., Zhuang, P., & Chen, H. (2022). A systematic review of *Orthosiphon stamineus* Benth. in the treatment of diabetes and its complications. *Molecules*, 27(2), 444. <https://doi.org/10.3390/molecules27020444>
- Webb, C. O. (2000). Exploring the phylogenetic structure of ecological communities: An example for rain forest trees. *The American Naturalist*, 156(2), 145–155. <https://doi.org/10.1086/303378>
- Wiens, J. J., & Tiu, J. (2012). Highly incomplete taxa can rescue phylogenetic analyses from the negative impacts of limited taxon sampling. *PLoS ONE*, 7(8), e42925. <https://doi.org/10.1371/journal.pone.0042925>
- Winkelman, R. (2013). *Cat's whiskers flowering stalk*. ClipPix ETC: Educational Photos for Students and Teachers. <https://etc.usf.edu/clippix/picture/cats-whiskers-flowering-stalk.html>
- Xiang, C.-L., Zhao, F., Cantino, P. D., Drew, B. T., Li, B., Liu, E.-D., Soltis, D. E., Soltis, P. S., & Peng, H. (2018). Molecular systematics of *Caryopteris* (Lamiaceae) and its allies with reference to the molecular phylogeny of subfamily Ajugoideae. *Taxon*, 67(2), 376–394. <https://doi.org/10.12705/672.7>
- Xu, J.-H., Liu, Q., Hu, W., Wang, T., Xue, Q., & Messing, J. (2015). Dynamics of chloroplast genomes in green plants. *Genomics*, 106(4), 221–231. <https://doi.org/10.1016/j.genbo.2015.07.004>

- Yalcin, F. N., & Kaya, D. (2006). Ethnobotany, pharmacology and phytochemistry of the genus *Lamium* (Lamiaceae). *Fabab Journal of Pharmaceutical Sciences*, 31, 43–52.
- Yao, X., Tan, Y., Yang, J., Wang, Y., Corlett, R. T., & Manen, J.-F. (2019). Exceptionally high rates of positive selection on the *rbcL* gene in the genus *Ilex* (Aquifoliaceae). *BMC Evolutionary Biology*, 19(1), 192. <https://doi.org/10.1186/s12862-019-1521-1>
- Yu, D., Pei, Y., Cui, N., Zhao, G., Hou, M., Chen, Y., Chen, J., & Li, X. (2023). Comparative and phylogenetic analysis of complete chloroplast genome sequences of *Salvia* regarding its worldwide distribution. *Scientific Reports*, 13(1), 14268. <https://doi.org/10.1038/s41598-023-41198-y>
- Yu, J., Lei, J., Yu, H., Cai, X., & Zou, G. (2004). Chemical composition and antimicrobial activity of the essential oil of *Scutellaria barbata*. *Phytochemistry*, 65(7), 881–884. <https://doi.org/10.1016/j.phytochem.2004.02.005>
- Yu, J., Xue, J.-H., & Zhou, S.-L. (2011). New universal *matK* primers for DNA barcoding angiosperms. *Journal of Systematics and Evolution*, 49(3), 176–181. <https://doi.org/10.1111/j.1759-6831.2011.00134.x>
- Yuan, Y.-W., Mabberley, D. J., Steane, D. A., & Olmstead, R. G. (2010). Further disintegration and redefinition of *Clerodendrum* (Lamiaceae): Implications for the understanding of the evolution of an intriguing breeding strategy. *Taxon*, 59(1), 125–133. <https://www.jstor.org/stable/27757057>
- Zhang, T.-T., Yang, Y., Song, X.-Y., Gao, X.-Y., Zhang, X.-L., Zhao, J.-J., Zhou, K.-H., Zhao, C.-B., Li, W., Yang, D.-G., Ma, X.-F., & Li, Z.-H. (2021). Novel structural variation and evolutionary characteristics of chloroplast tRNA in *Gossypium* plants. *Genes*, 12(6), 822. <https://doi.org/10.3390/genes12060822>
- Zhao, F., Chen, Y.-P., Salmaki, Y., Drew, B. T., Wilson, T. C., Scheen, A.-C., Celep, F., Bräuchler, C., Bendiksby, M., Wang, Q., Min, D.-Z., Peng, H., Olmstead, R. G., Li, B., & Xiang, C.-L. (2021). An updated tribal classification of Lamiaceae based on plastome phylogenomics. *BMC Biology*, 19(1), 1–27. <https://doi.org/10.1186/s12915-020-00931-z>
- Zhao, F., Drew, B. T., Chen, Y.-P., Hu, G.-X., Li, B., & Xiang, C.-L. (2020a). The chloroplast genome of *Salvia*: Genomic characterization and phylogenetic analysis. *International Journal of Plant Sciences*, 181(8), 812–830. <https://doi.org/10.1086/710083>
- Zhao, F., Li, B., Drew, B. T., Chen, Y. P., Wang, Q., Yu, W. B., Liu, E. D., Salmaki, Y., Peng, H., & Xiang, C. L. (2020b). Leveraging plastomes for comparative analysis and phylogenomic inference within Scutellarioideae (Lamiaceae). *PloS one*, 15(5), e0232602. <https://doi.org/10.1371/journal.pone.0232602>

- Zhao, F., Liu, E.-D., Peng, H., & Xiang, C.-L. (2017). A new species of *Scutellaria* (Scutellarioideae, Lamiaceae) from Sichuan Province in southwest China. *PeerJ*, 5, e3624. <https://doi.org/10.7717/peerj.3624>
- Zhu, B., Qian, F., Hou, Y., Yang, W., Cai, M., & Wu, X. (2021). Complete chloroplast genome features and phylogenetic analysis of *Eruca sativa* (Brassicaceae). *PLOS ONE*, 16(3), e0248556. <https://doi.org/10.1371/journal.pone.0248556>