

**PROFIL METABOLIT SERTA AKTIVITAS ANTIOKSIDAN PADA
AKAR DAN DAUN TANAMAN HANJELI (*Coix lacryma-jobi* L.) DESA
WISATA HANJELI SUKABUMI**

SKRIPSI

Diajukan untuk memenuhi sebagian syarat memperoleh gelar Sarjana Sains
Program Studi Biologi



Oleh
Zahra Apriyani Pratiwi
NIM 1902025

**PROGRAM STUDI BIOLOGI
FAKULTAS PENDIDIKAN MATEMATIKA DAN ILMU PENGETAHUAN ALAM
UNIVERSITAS PENDIDIKAN INDONESIA
2023**

**PROFIL METABOLIT SERTA AKTIVITAS ANTIOKSIDAN PADA
AKAR DAN DAUN TANAMAN HANJELI (*Coix lacryma-jobi* L.) DESA
WISATA HANJELI SUKABUMI**

Oleh
Zahra Apriyani Pratiwi

Skripsi yang diajukan untuk memenuhi sebagian syarat memperoleh gelar Sarjana
Sains pada Program Studi Biologi Fakultas Pendidikan Matematika dan Ilmu
Pengetahuan Alam

©Zahra Apriyani Pratiwi 2023
Universitas Pendidikan Indonesia 2023
Agustus 2023

Hak Cipta dilindungi Undang-Undang
Skripsi ini tidak boleh diperbanyak seluruhnya ataupun sebagian,
dengan dicetak ulang, difotokopi, atau cara lainnya tanpa izin dari penulis.

LEMBAR PENGESAHAN

ZAHRA APRIYANI PRATIWI
PROFIL METABOLIT SERTA AKTIVITAS ANTIOKSIDAN PADA AKAR
DAN DAUN TANAMAN HANJELI (*Coix lacryma-jobi* L.) DESA WISATA
HANJELI SUKABUMI

Disetujui dan disahkan oleh:

Pembimbing I,

Dr. R. Kusdianti, M.Si.
NIP. 196402261989032004

Pembimbing II

Hj. Tina Safaria Nilawati, S.Si., M.Si.
NIP. 197303172001122002

Mengetahui,

Ketua Program Studi Biologi,

Dr. Wahyu Surakusumah, S.Si., M.T.
NIP. 197212301999031001

PERNYATAAN

Dengan ini saya menyatakan bahwa skripsi dengan judul “**Profil Metabolit serta Aktivitas Antioksidan pada Akar dan Daun Tanaman Hanjeli (*Coix lacryma-jobi* L.) Desa Wisata Hanjeli Sukabumi**” ini beserta seluruh isinya merupakan karya saya sendiri. Saya tidak melakukan penjiplakan atau pengutipan dengan cara-cara yang tidak sesuai dengan etika ilmu yang berlaku dalam masyarakat keilmuan. Atas pernyataan ini, saya siap menanggung risiko/sanksi apabila di kemudian hari ditemukan adanya pelanggaran etika keilmuan atau ada klaim dari pihak lain terhadap keaslian karya saya ini.

Bandung, Agustus 2023

Pembuat Pernyataan

Zahra Apriyani Pratiwi

NIM. 1902025

UCAPAN TERIMA KASIH

Puji syukur penulis panjatkan kehadirat Allah SWT yang senantiasa memberikan rahmat dan karunia-Nya yang begitu melimpah, sehingga penulis diberi kesabaran, kekuatan, kemudahan, dan keikhlasan dalam menyelesaikan tugas akhir dan penulisan skripsi yang berjudul **“Profil Metabolit serta Aktivitas Antioksidan pada Akar dan Daun Tanaman Hanjeli (*Coix lacryma-jobi* L.) Desa Wisata Hanjeli Sukabumi”**.

Dalam penyusunan skripsi ini tidak terlepas dari bantuan dan bimbingan dari berbagai pihak. Oleh karena itu, dengan segala kerendahan hati dalam kesempatan ini penulis hendak mengucapkan terima kasih dan menyampaikan perhargaan sebesar-besarnya kepada semua pihak yang telah berkontribusi dengan memberikan dukungan baik berupa do'a, materi maupun gagasan. Ucapan terima kasih yang tak terukur ditujukan kepada:

1. Ibu Dr. R. Kusdianti, M.Si. selaku dosen pembimbing I yang selalu memberikan arahan, bimbingan, masukan, motivasi dan memfasilitasi hal yang dapat membantu pelaksanaan tugas akhir dan penulisan skripsi ini.
2. Ibu Hj. Tina Safaria, M.Si. selaku dosen pembimbing II yang selalu memberikan bimbingan dan membantu pelaksanaan tugas akhir dan penulisan skripsi ini.
3. Bapak Dr. Wahyu Surakusumah, S.Si., M.T. selaku ketua Program Studi Biologi FPMIPA UPI.
4. Ibu Prof. Dr. Hj. RR. Hertien Koosbandiah, M.Sc. selaku dosen wali atas bimbingan dan motivasi yang telah diberikan.
5. Seluruh dosen Departemen Pendidikan Biologi FPMIPA UPI atas segala ilmu, bimbingan, pengalaman, dan motivasi yang telah diberikan selama perkuliahan.
6. Kepada seluruh staf akademik yang selalu membantu dalam memberikan fasilitas, ilmu, serta pengalaman pada penulis hingga dapat menunjang dalam menyelesaikan skripsi ini.

7. Kepada Bapak Asep *founder* Desa Wisata Hanjeli beserta seluruh warga yang telah memberikan kesempatan dan membantu kepada kami untuk melakukan penelitian terkait hanjeli di Desa Wisata Hanjeli.
8. Frita Annisa Reina Aziz dan Hanna Yustianisa sebagai rekan penelitian yang telah sabar melewati setiap tahap penggerjaan skripsi ini bersama-sama dan selalu bersedia membantu serta mendukung dalam penelitian ini.
9. Kepada sahabat satu atap saya Sifa Aulia Rahma, Siti Wardah Khofifah, Armeta,Witasari, dan Luna Nuranisa Zakiah yang senantiasa menemani, mendukung, dan memberi semangat dalam menyelesaikan skripsi ini.
10. Kepada sahabat seperjuangan saya Shevira Arista, Syifa Indah, Fadhilah Haifa, dan Andi Milenia yang selalu mendukung, memberi semangat, dan berjuang bersama dalam menyelesaikan perkuliahan ini.
11. Kepada sahabat-sahabat kecil saya Raisa, Evi, dan Ertasya yang selalu mendukung dan memberi semangat dalam menyelesaikan skripsi ini.
12. Teman-teman Biologi C 2019 yang telah berjuang bersama-sama dari awal perkuliahan sampai akhirnya lulus.

Penulis juga mengucapkan banyak terima kasih kepada kedua orang tua, Bapak Hasan Basri dan Ibu Anih Suryanih, adik Nazila Asyabani, sepupu Sella Asmaul Insani, beserta keluarga besar yang selalu memberi motivasi, dukungan, kepercayaan, dan pengorbanan yang tidak dapat terukur, serta kepada seluruh pihak yang sangat berpengaruh dalam proses penyusunan skripsi ini yang tidak bisa penulis sebutkan satu persatu. Semoga Allah SWT senantiasa membalas semua kebaikan yang telah diberikan. Semoga penelitian ini dapat memberikan manfaat khususnya bagi penulis dan umumnya kepada semua yang membaca.

Profil Metabolit serta Aktivitas Antioksidan pada Akar dan Daun Tanaman Hanjeli (*Coix lacryma-jobi* L.) Desa Wisata Hanjeli Sukabumi

ABSTRAK

Hanjeli (*Coix lacryma-jobi* L.) telah banyak digunakan sebagai obat tradisional di berbagai negara. Tanaman ini diketahui mengandung metabolit yang beragam di setiap daerah. Desa Wisata Hanjeli Kabupaten Sukabumi merupakan tempat eduwisata yang bergerak di bidang budidaya tanaman lokal hanjeli. Penelitian ini bertujuan untuk mendapatkan profil metabolit yang terkandung pada akar dan daun hanjeli kultivar ketan dan batok serta aktivitas antioksidan dalam akar dan daun hanjeli batok. Sampel akar dan daun diambil dari Desa Wisata Hanjeli, Kabupaten Sukabumi. Sampel diekstrak menggunakan metode maserasi dengan pelarut etanol p.a. 70%. Kandungan metabolit akar dan daun hanjeli dianalisis menggunakan *Gas Chromatography-Mass Spectrometry* (GC-MS) dan diidentifikasi menggunakan pustaka WILLEY09TH. Hasil penelitian menunjukkan akar hanjeli ketan mengandung 9 senyawa, sedangkan akar hanjeli batok mengandung 5 senyawa. Kedua akar hanjeli didominasi oleh senyawa 4-vinilfenol (Akar hanjeli ketan = 24.40%, akar hanjeli batok = 35.12%) yang termasuk ke dalam golongan fenolik. Daun hanjeli ketan mengandung 8 senyawa yang didominasi oleh senyawa metil linoleat (17.78%) yang termasuk ke dalam golongan asam lemak, sedangkan daun hanjeli batok mengandung 9 senyawa yang didominasi oleh senyawa metil linolelaidat (12.49%) yang termasuk golongan asam lemak. Nilai IC₅₀ ekstrak akar (315,72 ppm) dan daun (390,64 ppm) hanjeli batok menunjukkan aktivitas antioksidan yang tergolong kategori lemah. Penelitian ini menunjukkan adanya perbedaan kandungan metabolit pada akar dan daun hanjeli dalam kultivar yang berbeda. Aktivitas antioksidan ekstrak akar lebih tinggi dibanding ekstrak daun hanjeli batok.

Kata kunci: Akar hanjeli, aktivitas antioksidan, daun hanjeli, Desa Wisata Hanjeli, *Gas Chromatography-Mass Spectrometry* (GC-MS), metabolit

Profile of Metabolites and Antioxidant Activity of Roots and Leaves of Hanjeli (*Coix lacryma-jobi* L.) Desa Wisata Hanjeli Sukabumi

ABSTRACT

Hanjeli (*Coix lacryma-jobi* L.) has been widely used as a traditional medicine in various countries. This plant is known to contain metabolites that vary in each region. Desa Wisata Hanjeli, Sukabumi Regency, is an educational destination engaged in the cultivation of local hanjeli plants. The purpose of the study to obtain profiles of metabolites contained in the roots and leaves and leaves of hanjeli cultivars of ketan and batok as well as antioxidant activity in the roots and leaves of hanjeli batok. Samples of roots and leaves were taken from Desa Wisata Hanjeli, Sukabumi Regency. Samples were extracted using the maceration method with ethanol p.a as solvent. 70%. The metabolites of hanjeli roots and leaves were analyzed using *Gas Chromatography-Mass Spectrometry* (GC-MS) and identified using the WILLEY09TH library. The results showed that the root of the hanjeli ketan contains 9 compounds, while the root of the hanjeli batok contains 5 compounds. Both hanjeli roots are dominated by 4-vinylphenol compounds (hanjeli ketan roots = 24.40%, hanjeli batok roots = 35.12%) which belong to the phenolic group. Hanjeli ketan leaves contain 8 compounds which are dominated by methyl linoleic compounds (17.78%) which belong to the fatty acid group, while hanjeli batok leaves contain 9 compounds which are dominated by methyl linoleic acid compounds (12.49%) which belong to the fatty acid group. The IC₅₀ value of the root extract (315.72 ppm) and leaves (390.64 ppm) of hanjeli batok showed antioxidant activity which was classified as a weak category. This study showed that there were differences in the content of metabolites in the roots and leaves of hanjeli in different cultivars. The antioxidant activity of the root was higher than that of the hanjeli batok leaf.

Keywords: Hanjeli roots, antioxidant activity, hanjeli leaves, *Gas Chromatography – Mass Spectrophotometry* (GC-MS), metabolite

DAFTAR ISI

LEMBAR PENGESAHAN	i
PERNYATAAN.....	ii
UCAPAN TERIMA KASIH	iii
ABSTRAK	v
ABSTRACT	vi
DAFTAR ISI.....	vii
DAFTAR TABEL	ix
DAFTAR GAMBAR.....	x
DATA LAMPIRAN	xii
BAB I PENDAHULUAN.....	1
1.1 Latar Belakang	1
1.2 Rumusan Masalah	3
1.3 Pertanyaan Penelitian	4
1.4 Tujuan Penelitian.....	4
1.5 Batasan Penelitian	4
1.6 Manfaat Penelitian.....	4
1.7 Struktur Organisasi Penelitian.....	5
BAB II ANALISIS METABOLIT TANAMAN HANJELI.....	7
2.1 Deskripsi Hanjeli (<i>Coix lacryma-jobi</i> L.).....	7
2.2 Metabolit	8
2.3 Ekstraksi	12
2.4 Gas Chromatography-Mass Spectrometry (GC-MS).....	13
2.5 Aktivitas Antioksidan.....	14
2.6 Uji Aktivitas Antioksidan dengan Metode DPPH.....	15
2.7 Deskripsi Desa Wisata Hanjeli	17
BAB III METODE PENELITIAN	18
3.1 Jenis Penelitian	18
3.2 Waktu dan Lokasi Penelitian.....	18
3.3 Populasi dan Sampel	19
3.4 Prosedur Penelitian.....	19
3.4.1 Pengambilan sampel	20

3.4.2 Pengukuran Faktor Abiotik.....	20
3.4.3 Persiapan Bahan.....	21
3.4.4 Pembuatan Ekstrak Akar dan Daun Hanjeli Ketan dan Hanjeli Batok.	23
3.4.5 Analisis Senyawa Bioaktif dengan GC-MS.....	29
3.4.6 Uji Aktivitas Antioksidan	25
3.4.7 Analisis Data Uji Aktivitas Antioksidan	28
3.5 Alur Penelitian.....	29
BAB IV TEMUAN DAN PEMBAHASAN	30
4.1 Kandungan Metabolit Akar Hanjeli	32
4.1.1 Kandungan Metabolit Akar Hanjeli Batok	32
4.1.2 Kandungan Metabolit Akar Hanjeli Ketan	39
4.1.3 Perbandingan Metabolit Akar Hanjeli Batok dan Hanjeli Ketan	44
4.2 Kandungan Metabolit Daun Hanjeli.....	48
4.2.1 Kandungan Metabolit Daun Hanjeli Batok	48
4.2.2 Kandungan Metabolit Daun Hanjeli Ketan	54
4.2.3 Perbandingan Metabolit Daun Hanjeli Batok dan Hanjeli Ketan.....	59
4.3 Perbandingan Kandungan Metabolit pada Akar Hanjeli Batok, Akar Hanjeli Ketan, Daun Hanjeli Batok, dan Daun Hanjeli Ketan	62
4.4. Aktivitas Antioksidan Akar dan Daun Hanjeli Batok	72
BAB V SIMPULAN, IMPLIKASI, DAN REKOMENDASI	76
5.1 Simpulan.....	76
5.2 Implikasi	76
5.3 Rekomendasi	77
DAFTAR PUSTAKA	78
LAMPIRAN.....	106

DAFTAR TABEL

Tabel	Halaman
3.1. Pembuatan Larutan Seri Ekstrak Akar Hanjeli Batok.....	28
3.2. Pembuatan Larutan Seri Ekstrak Daun Hanjeli Batok	28
4.1. Hasil Analisis Metabolit Ekstrak Etanol Akar Hanjeli Batok.....	32
4.2. Hasil Analisis Metabolit Ekstrak Etanol Akar Hanjeli Ketan.....	40
4.3. Hasil Analisis Metabolit Ekstrak Akar Hanjeli Batok dan Hanjeli Ketan.	46
4.4. Hasil Analisis Metabolit Ekstrak Etanol Daun Hanjeli Batok	49
4.5. Hasil Analisis Metabolit Ekstrak Etanol Daun Hanjeli Ketan	55
4.6. Hasil Analisis Metabolit Ekstrak Daun Hanjeli Batok dan Hanjeli Ketan.	62
4.7. Perbandingan Faktor Abiotik Penelitian Sebelumnya	69
4.8. Nilai IC ₅₀ Ekstrak Akar dan Daun Hanjeli Batok.....	72

DAFTAR GAMBAR

Gambar	Halaman
2.1. Tanaman Hanjeli	7
2.2. Ringkasan Jalur Utama Biosintesis Metabolit Sekunder	9
2.3. Jalur Sikimat	10
2.4. Jalur Mevalonat.....	11
2.5. Jalur Metileritritol-fosfat (MEP).....	11
2.6. Reaksi antara DPPH dan Antioksidan.....	16
3.1. Peta Lokasi Pengambilan Sampel	18
3.2. Lokasi Desa Wisata Hanjeli	19
3.3. Organ yang digunakan	19
3.4. Pengambilan Sampel.....	20
3.5. Pengukuran Faktor Abiotik	21
3.6. Proses Analisis MOT	21
3.7. Hasil Titrasi MOT	21
3.8. Simplisia Hasil Pengeringan	22
3.9. Pengeringan Sampel menggunakan Inkubator Suhu 50°C	22
3.10. Penghalusan Sampel menggunakan Blender	22
3.11. Pengayakan Sampel menggunakan Saringan 100 mesh	22
3.12. Serbuk Simplisia	23
3.13. Maserasi Serbuk Simplisia.....	23
3.14. Penyaringan Ekstrak.....	24
3.15. Ekstrak Hasil Maserasi.....	24
3.16. Penguapan Ekstrak menggunakan Waterbath.....	24
3.17. Ekstrak Akar dan Daun Hanjeli Hasil Penguapan	24
3.18. GC-MS AGILENT 5973.....	24
3.19. Spektrofotometer UV-Vis 1800	25
3.20. Penghomogenan Larutan DPPH	26
3.21. Larutan Ekstrak Sampel	27
3.22. Penghomogenan Larutan dengan Ultrasonic Cleaner	27
3.23. Penghomogenan Larutan dengan Centrifuge	27

3.24. Larutan Ekstrak Hanjeli Sebelum Diinkubasi.....	28
3.25. Larutan Ekstrak Hanjeli Setelah Diinkubasi	28
3.26. Pengukuran Absorbansi dengan Spektrofotometer UV-Vis	28
3.27. Alur Penelitian	29
4.1. Buah Hanjeli: A. Batok; B. Ketan.....	30
4.2. Sampel Daun Hanjeli: A. Batok; B. Ketan	30
4.3. Sampel Akar Hanjeli: A. Batok; B. Ketan	30
4.4. <i>Heatmap</i> Kandungan Senyawa pada Akar dan Daun Hanjeli	31
4.5. Golongan Senyawa yang Ditemukan pada Akar Hanjeli Batok	32
4.6. Biosintesis lignin	33
4.7. Biosintesis Koniferil Alkohol	35
4.8. Golongan Senyawa yang Ditemukan pada Akar Hanjeli Ketan	39
4.9. Kandungan Senyawa pada Akar Hanjeli Batok dan Hanjeli Ketan.....	44
4.10. Golongan Senyawa yang Ditemukan pada Daun Hanjeli Batok	49
4.11. Golongan Senyawa yang Ditemukan pada Daun Hanjeli Ketan	55
4.12. Kandungan Senyawa pada Daun Hanjeli Batok dan Hanjeli Ketan	60
4.13. Kandungan Senyawa Akar dan Daun Hanjeli Batok dan Hanjeli Ketan.	64

DATA LAMPIRAN

Lampiran	Halaman
1. Pengukuran Faktor Abiotik	106
2. Hasil Uji Antioksidan.....	107
3. Dokumentasi Penelitian	109
4. Hasil Analisis Metabolit Akar dan Daun Hanjeli dengan GC-MS	113

DAFTAR PUSTAKA

- Abd-ElGawad, A.M., Elshamy, A.I., Al-Rowaily, S.L., & El-Amier, Y.A. (2019). Habitat Affects the Chemical Profile, Allelopathy, and Antioxidant Properties of Essential Oils and Phenolic Enriched Extracts of the Invasive Plant *Heliotropium Curassavicum*. *Plants*, 8(11), 482. doi: [10.3390/plants8110482](https://doi.org/10.3390/plants8110482)
- Abdillah, D. (2015). *Pengaruh Cekaman Kekeringan terhadap Kandungan Fenolik dan Antioksidan Tanaman Sorgum (Sorghum bicolor L. Moench) pada Fase Awal Vegetatif*. (Skripsi). Fakultas Pertanian, Universitas Jember, Jawa Timur.
- Abubacker, M.N. & Deepalakshmi, T. (2013). In vitro Antifungal Potentials of Bioactive Compound Methyl Ester of Hexadecanoic Acid Isolated from *Annona muricata* Linn. (Annonaceae) Leaves. *Biosciences Biotechnology Research*, 10(2), 879-884. doi: <http://dx.doi.org/10.13005/bbra/1211>
- Acamovic, T. & Brooker J.D. (2005). Biochemistry of Plant Secondary Metabolites and Their Effects in Animals. *Proceeding of Nutrition Society*, 64, 403-412.
- Alasalvar, C., Grigor J.M., Zhang, D.L., Quantic, P.C., & Shahidi, F. (2001). Comparison of Volatiles, Phenolics, Sugars, Antioxidant Vitamins, and Sensory Quality of Different Colored Carrot Varieties. *Journal of Agriculture and Food Chemistry*, 49, 1410-1416.
- Agostini-Costa, T.Da., Vieira, R.F., Bizzo, H.R., Silveira, D., & Gimenes, M.A. (2012). Secondary Metabolites. Dalam S. Dhanarasu. (Penyunting), *Chromatography and Its Applications* (hlm. 131-166). Brazil: Intechope. doi: [10.5772/3570](https://doi.org/10.5772/3570)
- Agusta, A. (2000). *Minyak Atsiri Tumbuhan Tropika Indonesia*. Bandung: Institut Teknologi Bandung.
- Ahmed, S.H., Jamil, K.B., Ahmed, F.H., Mutlib, H.S., Ibrahim, T.Y., & Al-Newani, H.R.H. (2023). Study of the Antioxidant and Anti-Cancer Activity and Biological Activity of (*Punicagranatum* L.) Extract on Pathogenic Microorganisms Using Alkaline Energy Water. *South Asian Research Journal of Biology and Applied Biosciences*, 5(2). doi: [10.36346/sarjbab.2023.v05i02.005](https://doi.org/10.36346/sarjbab.2023.v05i02.005)
- Alharthy, K.M., Balaha, M.F., Devi, S., Altharawi, A., Yusufoglu, H.S., Aldossari, R.M., Alam, A., & Giacomo, V.D. (2023). Ameliorative Effects of Isoeugenol and Eugenol against Impaired Nerve Function and Inflammatory and Oxidative Mediators in Diabetic Neuropathic Rats. *Biomedicines*, 11, 1203. doi: <https://doi.org/10.3390/biomedicines11041203>

- Ampadu, G.A.A., Mensah, J.O., Darko G., & Borquaye, L.S. (2022). Essential Oils from the Fruits and Leaves of *Spondias mombin* Linn: Chemical Composition, Biological Activity, and Molecular Docking Study. *Evidence-Based Complementary and Alternative Medicine*. doi: <https://doi.org/10.1155/2022/7211015>
- Andriana, Y., Fajriani, N.A., Iwansyah, A.C., & Xuan, T.D. (2023). Phytochemical Constituents of Indonesian Adlay (*Coix lacrima-jobi* L.) and Their Potential as Antioxidants and Crop Protection Agents. *Agrochemicals*, 2, 135–149. doi: <https://doi.org/10.3390/agrochemicals2010010>
- Anggraito, Y.U., Susanti, R., Iswari, R.S., Yuniaستuti, A., Lisdiana, Nugrahaningsih, Habibah, N.A., Bintari, S.H., & Dafip, M. (2018). *Metabolit Sekunder dari Tanaman: Aplikasi dan Produksi*. Semarang: Universitas Negeri Semarang.
- Antunes, G.L., Matzenbacher, L.S., Costa, B.P., Basso, B.S., Levorse, V.G.S., Antunes, K.H., Costa-Ferro, Z.S.M., & Oliveira, J.R. (2022). Methoxyeugenol Protects Against Lung Inflammation and Suppresses Neutrophil Extracellular Trap Formation in an LPS-Induced Acute Lung Injury Model. *Inflammation*, 45(4), 1534–1547. doi: [10.1007/s10753-022-01639-8](https://doi.org/10.1007/s10753-022-01639-8)
- Antúnez-Mojica, M., Romero-Estrada, A., Hurtado-Díaz, I., Miranda-Molina, A., & Alvarez, L. (2021). Lignans from *Bursera fagaroides*: Chemistry, Pharmacological Effects and Molecular Mechanism. A Current Review. *Life*, 11, 685. doi: <https://doi.org/10.3390/life11070685>
- Anwar, T., Qureshi, H., Mahnashi, M.H., Kabir, F., Parveen, N., Ahmed, D., Afzal, U., Batool, S., Awais, M., & Ahmed, A.S. (2021). Bioherbicidal Ability and Weed Management of Allelopathic Methyl Esters from *Lantana camara*. *Saudi Journal of Biological Sciences*, 28, 4365–4374.
- Ao, C., Deba, F., Tako, M., & Tawata, S. (2009). Biological Activity and Composition of Extract from Aerial Root of *Ficus microcarpa* L. fil. *International Journal of Food Science & Technology*, 44(2), 349–358. doi: [10.1111/j.1365-2621.2008.01732.x](https://doi.org/10.1111/j.1365-2621.2008.01732.x)
- Arafa, N.M., Girgis, N.D., Ibrahim, M.M., Mohamed, S.S., & El-Bahr, M.K. (2022). Phytochemical Profiling by GC-MS Analysis and Antimicrobial Activity Potential of In vitro Derived Shoot Cultures of Some Egyptian Herbal Medicinal Plant. *Egyptian Journal of Chemistry*, 65(9), 155-169. doi: [10.21608/EJCHEM.2022.115045.5230](https://doi.org/10.21608/EJCHEM.2022.115045.5230)
- Asami, E., Kitami, M., Ida, T., Kobayashi, T., & Saeki, M. (2023). Anti-inflammatory Activity of 2-methoxy-4-vinylphenol Involves Inhibition of Lipopolysaccharide-induced Inducible Nitric Oxidase Synthase by Heme Oxygenase-1. *Immunopharmacology and immunitoxicology*. doi: <https://doi.org/10.1080/08923973.2023.2197141>

- Ayaz, M., Junaid, M., Ullah, F., Ovais, M., Ahmad, W., Ahmad, S., & Zeb, A. (2016). Chemical Profiling, Antimicrobial and Insecticidal Evaluations of *Polygonum hydropiper* L. *BMC Complementary and Alternative Medicine*, 16, 502. doi: [10.1186/s12906-016-1491-4](https://doi.org/10.1186/s12906-016-1491-4)
- Azizah, B. & Salamah, N. (2013). Standarisasi Parameter Non Spesifik dan Perbandingan Kadar Kurkumin Ekstrak Etanol dan Ekstrak Terpurifikasi Rimpang Kunyit. *Pharmaciana*, 3(1).
- Azwanida, N.N. (2015). A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation. *Med Aromat Plants*, 4(196), 2167-0412. doi: [10.4172/2167-0412.1000196](https://doi.org/10.4172/2167-0412.1000196)
- Badan Pusat Statistik Kabupaten Sukabumi. (2016). *Kabupaten Sukabumi Dalam Angka 2016*. (Online). Diakses dari <https://sukabumikab.bps.go.id/publication/2017/08/04/5ac28d5fefb8e7a6de6bf7ab/kabupaten-sukabumi-dalam-angka-2016.html>
- Badaring, R.D., Sari S.P.M., Nurhabiba, S., & Wulan, W. (2020). Uji Ekstrak Daun Maja (*Aegle marmelos* L.) terhadap Pertumbuhan Bakteri *Escherichia coli* dan *Staphylococcus aureus*. *Indonesian Journal of Fundamental Science*, 6(1). doi: <http://dx.doi.org/10.26858/ijfs.v6i1.13941>
- Bamigboye, C.O., Fatoki, I.O., Yakubu, O.F., & Biodum, R. (2021). Antimicrobial Activity and Phytochemical Analysis of Some Selected Plants against Clinical Pathogens. *Tropical Journal Natural Product Research*, 5(4), 732-738.
- Basso, D.S.B., Haute, G. V., Ortega-Ribera, M., Luft, C., Antunes, G. L., Bastos, M.S., Carlessi, L.P., Levorse, V.G., Cassel, E., Donadio, M.V.F., Santarem, E.R., Grachia-Sancho, J., & de Oliveira, J.R. (2021). Methoxyeugenol Deactivates Hepatic Stellate Cells and Attenuates Liver Fibrosis and Inflammation through a PPAR- γ and NF-kB Mechanism. *Journal of Ethnopharmacology*, 280, 114433. doi: [10.1016/j.jep.2021.114433](https://doi.org/10.1016/j.jep.2021.114433)
- Bebia, P.G., Edet, U.O., Eyo, A.-A.O., Ogar, A.Y., Ugwu, J.C., Mbim, E.N., Ogbag, O.M., & Nwaokonie, F.O. (2023). Antimicrobial Activity of *Tetrapleura tetraptera* (Uyayak) Extracts, ADMET Profiling and Molecular Docking of its Bioactive Compounds Against Dihydropteroate Synthase of *Escherichia coli*. *Research Square*. doi: <https://doi.org/10.21203/rs.3.rs-2622083/v1>
- Belkheir, A.K., Gaid, M., Liu, B., Hänsch, R., & Beerhues, L. (2016). Benzophenone Synthase and Chalcone Synthase Accumulate in the Mesophyll of *Hypericum perforatum* Leaves at Different Developmental Stages. *Frontiers in Plant Science*, 7, 921. doi: <https://doi.org/10.3389/fpls.2016.00921>

- Bendre, R.S., Rajput, J.D., Bagul, S.D., & Karandikar, P.S. (2016). Outlooks on Medicinal Properties of Eugenol and Its Synthetic Derivatives. *Natural Products Chemistry & Research*, 4(3). doi: [10.4172/2329-6836.1000212](https://doi.org/10.4172/2329-6836.1000212)
- Bi, C., Chen, F., Jackson, L., Gill, B.S., & Li, W. (2011). Expression of Lignin Biosynthetic Genes in Wheat during Development and Upon Infection by Fungal Pathogens. *Plant Molecular Biology Reporter*, 29, 149–61.
- Borges, C.V., Minatel, I.O., Gomez-Gomez, H.A., & Lima, G.P.P. (2017). Medicinal Plants: Influence of Environmental Factors on the Content of Secondary Metabolites. *Medicinal Plants and Environmental Challenges*, 259–277.
- Bouket, A.C., Narmani, A., Sharifi, K., Naeimi, S., Mogaddam, M.R.A, Hamidi, A.A., Luptakova, L., Alenezi, F.N., & Belbahri, L. (2023). Semi-VOCs of Wood Vinegar Display Strong Antifungal Activities against Oomycete Species *Globisporangium ultimum* and *Pythium aphanidermatum*. *Microbiol. Res.*, 14, 371–389.
- Bhandari, S.R., Park, S.K., Cho, Y.C., & Lee, Y.S. (2012). Evaluation of Phytonutrients in Adlay (*Coix lacryma-jobi* L.) Seeds. *African Journal of Biotechnology*, 11(8), 1872–1878. doi: [10.5897/AJB11.2416](https://doi.org/10.5897/AJB11.2416)
- Boerjan, W., Ralph, J., & Baucher, M. (2003). Lignin Biosynthesis. *Annu. Rev. Plant Biol.*, 54: 519–546. doi: [10.1146/annurev.arplant.54.031902.134938](https://doi.org/10.1146/annurev.arplant.54.031902.134938)
- Bribi, N. (2018). Pharmacological Activity of Alkaloids: A Review Pharmacological Activity of Alkaloids: A Review. *Asian Journal of Botany*, 1. doi: [10.63019/ajb.v1i2.467](https://doi.org/10.63019/ajb.v1i2.467)
- Brighente, I.M.C., Dias, M., Verdi, L.G., & Pizzolatti, MG. (2007). Antioxidant Activity and Total Phenolic Content of Some Brazilian Species. *Pharmaceutical Biology*, 45(2), 156–161.
- Budiarso, F.S., Suryanto, E., & Yudishtira, A. (2017). Ekstraksi dan Aktivitas Antioksidan dari Biji Jagung Manado Kuning (*Zea mays* L.). *Pharmacon*, 6(3).
- Buhian, W.P.C., Rubio, R.O., Jr. Valle, D.L., & Martin-Puzin, J.J. (2016). Bioactive Metabolite Profiles and Antimicrobial Activity of Ethanolic Extracts from *Muntingia calabura* L. Leaves and Stems. *Asian Pacific Journal of Tropical Biomedicine*, 6(8), 682–685. doi: [10.1016/j.apjtb.2016.06.006](https://doi.org/10.1016/j.apjtb.2016.06.006)
- Cahyadi, W. (2006). *Kedelai Khasiat dan Teknologi*. Bandung: Bumi Aksara.
- Calixto, P.S., Almeida, R.N., Salvadori, M.G.S.S., Maia, M.D.S., Filho, J.M.B., Scotti, M.T., & Scotti, L. (2021). In Silico Study Examining New Phenylpropanoids Targets with Antidepressant Activity. *Curr Drug Targets*, 22(5), 539–554. doi: [10.2174/1389450121666200902171838](https://doi.org/10.2174/1389450121666200902171838)

- Carullo, G., Ramunno A, Sommella, E.M., Luca, M.D., Belsito, E.L., Frattaruolo, L., Brindisi, M., Campiglia, P., Cappello, A.R. & Aiello, F. (2022). Ultrasound-Assisted Extraction, Chemical Characterization, and Impact on Cell Viability of Food Wastes Derived from Southern Italy Autochthonous Citrus Fruits. *Antioxidants*, 11, 285. doi: <https://doi.org/10.3390/antiox11020285>
- Cavoski, I., Caboni, P., & Miano, T. (2011). Natural pesticides and future perspectives. Dalam Margarita Stoytcheva (Penyunting), *Pesticides in the Modern World - Pesticides Use and Management* (hlm 169- 190). Rijeka: InTech Europe.
- Chairunnisa, S., Wartini, N.M., & Suhendra, L. (2019). Pengaruh Suhu dan Waktu Maserasi terhadap Karakteristik Ekstrak Daun Bidara (*Ziziphus mauritiana* L.) sebagai Sumber Saponin. *Rekayasa dan Manajemen Agroindustri*, 7(4), 551–560.
- Cheng, G., Zhang, X., Chen, Y., Lee, R.J., Wang, J., Yao, J., Zhang, Y., Zhang, C., Wang, K., & Yu, B. (2019). Anticancer Activity of Polymeric Nanoparticles Containing Linoleic Acid-SN38 (LA-SN38) Conjugate in a Murine Model of Colorectal Cancer. *Colloids and Surfaces B Biointerfaces*, 181, 822–829. doi: [10.1016/j.colsurfb.2019.06.0](https://doi.org/10.1016/j.colsurfb.2019.06.0)
- Chhabra, D. & Gupta, R.K. (2015). Formulation and Phytochemical Evaluation of Nutritional Product Containing Job's tears (*Coix lachryma-Jobi* L.). *Journal of Pharmacognosy and Phytochemistry*, 4(3), 291-298.
- Choi, J., Shin, K-M., Park, H-J., Jung, H-J., Kim, H-J., Lee, Y-S., Rew, J.H., & Lee, K.T. (2004). Anti-Inflammatory and Antinociceptive Effects of Sinapyl Alcohol and its Glucoside Syringin. *Planta Medica*, 70, 1027-1032.
- Close, D.C. & McArthur, C. (2002). Rethinking the Role of Many Plant Phenolics – Protection from Photodamage, Not Herbivores?. *Oikos*, 99, 166–172.
- Costa, B.P., Nassr, M.T., Diz, F.M., Fernandes, K.H.A., Antunes, G.L., Grun, L.K., Barbé-Tuana, F.M., Nunes, F.B., Branchini, G., & de Oliveira, J.R. (2021). Methoxyeugenol Regulates the p53/p21 Pathway and Suppresses Human Endometrial Cancer Cell Proliferation. *Journal of Ethnopharmacology*, 267, 113645. doi: [10.1016/j.jep.2020.113645](https://doi.org/10.1016/j.jep.2020.113645)
- Croteau, R., Kutchan, T.M., & Lewis, N.G. (2000). Natural Products (Secondary metabolites). Dalam Bob B. Buchanan, Wilhelm Gruissem, & Russell L. Jones (Penyunting), *Biochemistry & Molecular Biology of Plants* (hlm. 1250- 1318). Amerika Serikat: Wiley.
- Da Silva Port's, P., Chisté, R.C., Godoy, H.T., & Prado, M.A. (2013). The Phenolic Compounds and the Antioxidant Potential of Infusion of Herbs from the

- Brazilian Amazonian Region. *Food Research International*, 53(2), 875–881.
- Dalawai, D., Murthy, H.N., Dewir Y.H., Sebastian, J.K., & Nag, A. (2023). Phytochemical Composition, Bioactive Compounds, and Antioxidant Properties of Different Parts of Andrographis macrobotrys Nees. *Life*, 13, 1166. doi: <https://doi.org/10.3390/life13051166>
- Daldiyono, Ismail, A., Rani A.A., Manan, C., & Sumadibrata, R. (1990). Kanker Kolon dan Peran Diet Tinggi Serat: Kejadian di Negara Barat. *Gizi Indonesia*, 15(1), 73- 75.
- Dalimartha S. (2008). *Atlas tumbuhan obat Indonesia*. Jakarta: Pustaka Bunda.
- Darmapatni, K.A.G., Basori, A., & Suaniti, N.M. (2016). Pengembangan Metode GC-MS untuk Penetapan Kadar Acetaminophen pada Spesimen Rambut Manusia. *Jurnal Biosains Pascasarjana*, 18(3). doi: <http://dx.doi.org/10.20473/jbp.v18i3.2016.255-266>
- Das, G., Gouda, S., Kerry, R.G., Cortes, H., Prado-Audelo, M.L, Leyva-Gomez, G., Fokou, P.V.T., Gutierrez-Grijalva, E.P., Heredia, J.B., Shin, H.S., & Patra, J.K. (2022). Study of Traditional Uses, Extraction Procedures, Phytochemical Constituents, and Pharmacological Properties of *Tiliacora triandra*. *Journal of Chemistry*. doi: <https://doi.org/10.1155/2022/8754528>
- Das, S., Akhter, R., Khandaker, S., Huque, S., Das, P., Anwar, & Shahriar, M. (2017). Phytochemical Screening, Antibacterial and Anthelmintic Activities of Leaf and Seed Extracts of *Coix lacryma-jobi* L. *Journal of Coastal Life Medicine*, 5(8), 360-364.
- Das, S., Vasudeva, N., & Sharma, S. (2014). Chemical Composition of Ethanol Extract of *Macrotyloma uniflorum* (Lam.) Verdc. Using GC-MS Spectroscopy. *Organic and Medicinal Chemistry Letters*, 4(13).
- De Abreu, N. & Mazzafera, P. (2005). Effect of Water and Temperature Stress on the Content of Active Constituents of *Hypericum brasiliense* Choisy. *Plant Physiol. Biochem*, 43, 241–248.
- Dehpour, A.A., Ebrahimzadeh, M.A., Fazel, N.S., & Mohammad, N.S. (2009). Antioxidant Activity of Methanol Extract of *Ferula assafoetida* and Its Essential Oil Composition. *Grasas Aceites*, 60(4), 405-412.
- Dhaniaputri, R., Suwono, H., Amin, M., & Lukiat, B. (2021). Introduction to Plant Metabolism, Secondary Metabolites Biosynthetic Pathway, and In-Silico Molecular Docking for Determination of Plant Medicinal Compounds: An Overview. *Advances in Biological Sciences Research*, 22. doi: <https://dx.doi.org/10.2991/absr.k.220406.053>

- Diab, T.A., Donia, T.K., & Saad-Allah, K. (2021). Characterization, Antioxidant, and Cytotoxic Effects of Some Egyptian Wild Plant Extracts. *Beni-Suef University Journal of Basic and Applied Sciences*, 10(1). doi: <http://dx.doi.org/10.1186/s43088-021-00103-0>
- Dicosmo, F. & Tower, G.H.N. (1984). Stress and Secondary Metabolism in Culture. Dalam Barbara, Cornelius Steelink, Frank, A (Penyunting), *Plant Cell in Phytochemical Adaption to Stress*. Toronto: Plenum Publishing Co.
- Dinde, A.V., Lokhande, P.B., & Mujawar, H.A (2018). Essential Oil Extraction, Characterization and Antimicrobial Study of *Blumea laciniata* DC from Konkan Region. *J. Biol. Chem. Chron*, 4(2), 70-76.
- Diningrat, S.D., Harahap, N.S., Risfandi, M., Zulfahri, Sari, A.N., & Kusdianti. (2021). Antioxidant and Antibacterial Activities of *Coix lacryma-jobi* Seed and Root Oil Potential for Meningitis Treatment. *Jordan Journal of Biological Sciences*, 14(5). doi: <https://doi.org/10.54319/jjbs/140501>
- Diningrat, S.D., Risfandi, M., Harahap, N.S., Sari, A.N., Kusdianti, & Siregar, H.K. (2020). Phytochemical Screening and Antibacterial Activity *Coix lacryma-jobi* Oil. *Journal Plant Biotechnology*, 47, 100-106. doi: <https://doi.org/10.5010/JPB.2020.47.1.100>
- Dixon, RA, Achnine, L., Kota, P., Liu, C.J., Reddy, M.S.S., & Wang, L. (2002). The Phenylpropanoid Pathway and Plant Defence—a Genomics Perspective. *Mol Plant Pathol*, 3, 371–90.
- Duke, J.A. (Penyunting). (2017). *Handbook of Phytochemical Constituent Grass, Herbs and Other Economic Plants*. New York: Routledge. doi: <https://doi.org/10.1201/9780203752623>
- Dutra, R.C., Leite, M.N., & Barbosa, N.R. (2008). Quantification of Phenolic Constituents and Antioxidant Activity of *Pterodon Emarginatus* Vogel Seeds. *International journal of molecular sciences*, 9(4), 606–614.
- ECHO Staff. (2013). Job's Tears (*Coix lacryma-jobi* L.), a Resilient and Multipurpose Grain. *ECHO Development Notes* No. 120.
- Edreva A, Velikova V, Tsonev T, Dagnon S, Gürel AL & Aktas L. (2008). Stress-protective Role of Secondary Metabolites: Diversity of Functions and Mechanisms. *Genetic Application of Plant Physiology*, 34, 67-78.
- El-Demerdash, E. (2011). Anti-inflammatory and Antifibrotic Effects of Methyl Palmitate. *Toxicol Appl Pharmacol*, 254(3), 238-244. doi: [10.1016/j.taap.2011.04.016](https://doi.org/10.1016/j.taap.2011.04.016)

- Erviani, A.E. & Arif, A.R. (2017). Rendemen Analysis and Phytochemical Screening of *Perinereis aibuhitensis* Extracts. *International Journal of Current Research and Academic Review*, 5(11), 25-29.
- Eze, I., Madubunyil, I.I., Udem, S.C., & Peter-Ajuzie, I.K. (2021). The Hepatoprotective Activity of the Methanol Leaf Extract of *Lasimorpha Senegalensis* against Liver Injury Induced by Paracetamol (Acetaminophen) in Albino Wistar Rats. *The Proceedings of the Nigerian Academy of Science*, 14(1). doi: [10.57046/ZOSX1419](https://doi.org/10.57046/ZOSX1419)
- Fernandes, R.M.N, Rodrigues M.AM., Panontin, J.F., Alves, D.R., Morais, S.M., Soares, I.M., & Scapin, E. (2021). Chemical Investigation, Toxic Potential and Acetylcholinesterase Inhibitory Effect of *Parkia platycephala* Leaf and Seed Extracts. *Journal Medicinal Plants Research*, 15(9), 401-412. doi: [10.5897/JMPR2021.7158](https://doi.org/10.5897/JMPR2021.7158)
- Fessenden, R.J. & Fessenden, J.S. (1982). *Kimia Organik*. Jakarta: Penerbit Erlangga.
- Furchan (2004). *Pengantar Penelitian dalam Pendidikan* (hlm. 54). Yogyakarta: Pustaka Pelajar Offset.
- Fuzzati, N., Sutarjadi, Dyatmiko, W., Rahman, A., & Hostettmann, K. (1995). Phenylpropane Derivatives from Roots of *Cosmos caudatus*. *Phytochemistry*, 39(2), 409–412. doi:[10.1016/0031-9422\(95\)00031-2](https://doi.org/10.1016/0031-9422(95)00031-2)
- Gandjar, I.G. & Rohman, A. (2012). *Analisis Obat Secara Spektrofotometri dan Kromatografi*. Yogyakarta: Pustaka Pelajar.
- Gecibesler, I.H., Yaglioglu, A.S., Gul, F., Temirturk, M., & Demirtas, I. (2017). Phytochemicals of *Chrysophthalmum montanum* (DC.) Boiss. Roots and Their Antiproliferative Activities Against HeLa and C6 Cell Lines. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*. doi: [10.1007/s40011-017-0925-1](https://doi.org/10.1007/s40011-017-0925-1)
- Gismar, A.Y., (2021). *Analisis Metabolit Sekunder dari Akar dan Daun Hanjeli (*Coix lacryma-jobi* L.) Ketan dan Hanjeli Putih Menggunakan GC-MS*. (Skripsi). Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam, Universitas Pendidikan Indonesia, Bandung.
- Gowda, M.D.M., Jayachandra, K., Joshi, V., Manjunprashanna, V.N., Rudresha, G.V., Velmurugan, D., Pachaiappan, R., Jameel, N.M., & Vishwanath, B.S. (2021). Gaertn Bran Suppresses Inflammatory Response Through the Down-Regulation of cPLA2, COX-2, IkB α , p38 and MPO Signaling in sPLA2 Induced Mice Paw Edema. *Research Square*. doi: <https://doi.org/10.21203/rs.3.rs-755579/v1>

- Green, R.J. (2004). *Antioxidant Activity of Peanut Plant Tissues.* (Tesis). North Caroline State University, Department of Food Science, Raleigh.
- Ghahari, S., Alinezhad, H., Nematzadeh, Gh. A., Tajbakhsh, M., & Baharfar, R. (2017). Biochemical Composition, Antioxidant and Biological Activities of the Essential Oil and Fruit Extract of *Xanthium strumarium* Linn. From Northern Iran. *Journal of Agricultural Scince and Technology*, 19, 1603-1616.
- Gruenbacherand, G. & Thurnher, M. (2017) Mevalonate Metabolism Immuno in Oncology. *Frontiers Immunology*, 8, 1-8. doi: [10.3389/fimmu.2017.01714](https://doi.org/10.3389/fimmu.2017.01714)
- Gutzeit, H.O. & Ludwig-Muller, J. (2014). *Plant Natural Products: Synthesis, biological functions and practical applications, First Edition.* New York: Wiley Blackwell.
- Hamed, A.B., Mantawy, E.M., El-Bakly, W.M., Abdel-Mottaleb, Y., & Azab, S.S. (2020). Methyl Palmitate: the Naturally Occurring Cardioprotective Agent. *Archives of Pharmaceutical Sciences Ain Shams University*, 24(1), 47-62.
- Hammerschmidt, R. & Kuć, J. (1982). Lignification as a Mechanism for Induced Systemic Resistance in *Cucumber*. *Physiological Plant Pathology*, 20(1), 61–71. doi: [10.1016/0048-4059\(82\)90024-8](https://doi.org/10.1016/0048-4059(82)90024-8)
- Hamzah, N., Ismail, I., & Sandi, A.D. (2014). Pengaruh Emulgalator terhadap Aktivitas Antioksidan Krim Ekstrak etanol Kelopak Bunga Rosella (*Hibiscus sabdariffa*). *Jurnal Kesehatan*, 7(2), 377-385. doi: <https://doi.org/10.24252/kesehatan.v7i2.57>
- Han, X., Zhang, X., Wang, Q., Wang, L., & Yu, S. (2020). Antitumor Potential of *Hedyotis diffusa* Willd: A Systematic Review of Bioactive Constituents and Underlying Molecular Mechanisms. *Biomedicine & Pharmacotherapy*, 130. doi: <https://doi.org/10.1016/j.bioph.2020.110735>
- Handayani, F., Sumarmiyati, & Rahayu, S.P. (2019). Karakteristik Morfologi jelai (*Coix lacryma-jobi* L.) lokal Kalimantan Timur. *Pros Sem Nas Masy Biodiv Indon*, 5(2), 228-233. doi: [10.13057/psnmbi/m0](https://doi.org/10.13057/psnmbi/m0)
- Hanani, E., Mun'im, A., & Sekarini, R. (2005). Identifikasi Senyawa Antioksidan Dalam Spons *Callyspongia* sp. Dari Kepulauan Seribu. *Majalah Ilmu Kefarmasian*, 2(3), 127-133.
- Harada, H., Sakagami, H., Konno, K., Sato, T., Osawa, N., Fujimaki, M., & Komatsu, N. (1988). Induction of Antimicrobial Activity by Antitumor Substances from Pine Cone Extract of *Pinus parviflora* Sieb. et Zucc. *Anticancer Res*, 8, 581–588.
- Harborne, J.B. (1987). *Metode Fitokimia*. Bandung: Penerbit ITB.

- He, M., He, C.Q., & Ding, N.Z. (2018). Abiotic Stresses: General Defenses of Land Plants and Chances for Engineering Multistress Tolerance. *Frontiers in Plant Science*, 9, 1771. doi: [10.3389/fpls.2018.01771](https://doi.org/10.3389/fpls.2018.01771)
- Hermanto. (2008). *Aplikasi Alat HPTLC dan GC-MS*. Jakarta: UI Press.
- Hien, T.T., Ha, D.T., Truong, D.M., Duc, L., Dao, T., & Hai, N. (2016). Triolein from *Coix lacryma-jobi* Induces Cell Cycle Arrest Through p53/p21 Signaling Pathway. *Biomed and Pharmacology Journal*, 9(2), 519-524. doi: <http://dx.doi.org/10.13005/bpj/967>.
- Hu, J., Wenzhe, H., Fantao, Z., Xiangdong, L., Yaling, C., & Jiankun, X. (2020). Variability of Volatile Compounds in the Medicinal Plant *Dendrobium officinale* from Different Regions. *MDPI Molecules*, 25(21), 5046. doi: <https://doi.org/10.3390/molecules25215046>
- Hu, Q., Zhou, M., & Wei, S. (2018). Progress on the Antimicrobial Activity Research of Clove Oil and Eugenol in the Food Antiseptics Field. *Journal of Food Science*, 83(6), 1476–1483. doi: [10.1111/1750-3841.14180](https://doi.org/10.1111/1750-3841.14180)
- Huang, C.B., George, B., & Ebersole, J.L. (2010). Antimicrobial Activity of n-6, n-7 and n-9 Fatty Acids and Their Esters for Oral Microorganisms. *Archives of Oral Biology*, 55, 555–560.
- Husna, R.S.N., Effendi, M.E., & Maheswari, H. (2016). Efek Samping Ekstrak Etanol 96% dan 70% Herba Kemangi (*Ocimum americanum* L.) yang Bersifat Estrogenik Terhadap Kadar Asam Urat pada Tikus Putih. *Ekologia*, 16(2), 32-38.
- Ide, P. (2008). *Gaya Hidup Penghambat Alzheimer*. Jakarta: PT. Elex Komputindo.
- Illahi, A.K., Yusniwati, & Swasti, E. (2021). Eksplorasi dan Karakterisasi Hanjeli (*Coix lacryma-jobi* L.). *Lumbung*, 20(1). doi: <http://dx.doi.org/10.32530/lumbung.v20i1.245>
- İnci, S., Sancar, P.Y., Demirpolat, A., Kirbag, S., & Civelek, S. (2022). Chemical Compositions of Essential Oils, Antimicrobial Effect and Antioxidant Activity Studies of *Hyoscyamus niger* L. from Turkey. *BioRxiv preprint*. doi: <https://doi.org/10.1101/2022.08.07.503024>
- Inggrid, M. & Santoso, H. (2014). *Ekstraksi Antioksidan dan Senyawa Aktif dari Buah Kiwi (Actinidia deliciosa)*. Bandung: Research Report Engineering Science
- Irawan, B. (2010). *Peningkatan Mutu Minyak Nilam dengan Ekstraksi dan Destilasi pada Berbagai Komposisi Pelarut*. (Tesis). Universitas Diponegoro, Semarang.

- Ismail, N. Z., Arsal, H., Samian, M. R., & Hamdan, M. R. (2017). Determination of Phenolic and Flavonoid contents, Antioxidant Activities and GC-MS Analysis of *Clinacanthus nutans* (Acanthaceae) in Different Locations. *AGRIVITA Journal of Agricultural Science*, 39(3), 335–344. doi: <http://doi.org/10.17503/agrivita.v39i3.1076>
- ITIS. (2022). *Coix lacryma-jobi L.* [Online]. Diakses dari https://www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=41586#null (24 Oktober 2022).
- Jeong, J.B., Hong, S.C., Jeong, H.J., & Koo, J.S. (2011). Anti-inflammatory Effect of 2-methoxy-4-vinylphenol via The Suppression of NF-κB and MAPK Activation, and Acetylation of Histone H3. *Archives of Pharmacal Research*, 34(12), 2109–2116. doi: [10.1007/s12272-011-1214-9](https://doi.org/10.1007/s12272-011-1214-9)
- Julianto, T.S. (2019). *Fitokimia Tinjauan Metabolit Sekunder dan Skrining Fitokimia*. Yogyakarta: Universitas Islam Indonesia.
- Jumlongkul, P. & Chaikul, P. (2021). Antioxidant Activity of Acidified Ethanolic Adlay (*Coix lacryma-jobi* L) Seed Extract In Vitro and Cell Culture Assays. *Proceedings of RSU International Research Conference*. Thailand: RSU International Research Conference.
- Kachroo, A. & Kachroo, P. (2009). Fatty Acid-Derived Signals in Plant Defense. *Annual Review of Phytopathology*, 47(1), 153–176. doi: [10.1146/annurev-phyto-080508-081820](https://doi.org/10.1146/annurev-phyto-080508-081820)
- Karpinski, S., Gabrys, H., Mateo, A., Karpinska, B., & Mullineaux, P.M. (2003). Light Perception in Plant Disease Defence Signalling. *Current Opinion in Plant Biology*, 6, 390–396.
- Katrin & Bendra, A. (2015). Aktivitas Antioksidan Ekstrak, Fraksi, dan Golongan Senyawa Kimia Daun *Premna oblongata* Miq. *Pharmaceutical Science and Research*, 2(1), 21-31.
- Kchaou, W., Abbès, F., Blecker, C., Attia, H., & Besbes, S. (2013). Effects of Extraction Solvents on Phenolic Contents and Antioxidant Activities of Tunisian Date Varieties (*Phoenix dactylifera* L.). *Industrial Crops and Products*, 45, 262–269. doi: [10.1016/j.indcrop.2012.12.028](https://doi.org/10.1016/j.indcrop.2012.12.028)
- Khare, S., Singh, N. B., Singh, A., Hussain, I., Niharika, K., Yadav, W., Bano, C., Yadav, R.K., & Amist, N. (2020). Plant Secondary Metabolites Synthesis and their Regulations Under Biotic and Abiotic Constraints. *Journal of Plant Biology*. doi: [10.1007/s12374-020-09245-7](https://doi.org/10.1007/s12374-020-09245-7)
- Kementerian Pariwisata dan Ekonomi Kreatif. (2023). *Desa Wisata Hanjeli*. [Online]. Diakses dari <https://jadesta.kemenparekraf.go.id/desa/hanjeli> (27 Januari 2023).

- Kemit, N., Rai W.I., & Nocianitri, K. (2016). Pengaruh Jenis Pelarut dan Waktu Maserasi terhadap Kandungan Senyawa Flavonoid dan Aktivitas Antioksidan Ekstrak Daun Alpukat (*Persea americana mill.*). *Jurnal Ilmu Dan Teknologi Pangan (ITEPA)*, 5(2), 130-141.
- Keawsa-ard, S., Liawruangrath, B., Liawruangrath, S., Teerawutgulrag, A., & Pyne, S.G. (2012). Chemical Constituents and Antioxidant and Biological Activities of the Essential Oil from Leaves of *Solanum spirale*. *Natural Product Communications*, 7(7).
- Keen, N.T. & Littlefield, L.J. (1979). The Possible Association of Phytoalexins with Resistance Gene Expression in Flax to *Melampsora lini*. *Physiological Plant Pathology*, 14(3), 265–280. doi: [10.1016/0048-4059\(79\)90048-1](https://doi.org/10.1016/0048-4059(79)90048-1)
- Khiralla, A., Spina, R., Varbanov, M., Philippot, S., Lemiere, P., Slezack-Deschaumes, S., André, P., Mohamed, I., Yagi, S.M., & Laurain-Mattar, D. (2020). Evaluation of Antiviral, Antibacterial and Antiproliferative Activities of the Endophytic Fungus *Curvularia papendorfii*, and Isolation of a New Polyhydroxyacid. *Microorganisms*, 8(9), 1353. doi:[10.3390/microorganisms809135](https://doi.org/10.3390/microorganisms809135)
- Kilian, J., Whitehead, D., Horak, J., Wanke, D., Weinl, S., Batistic, O., D'Angelo, C., Bornberg-Bauer, E., Kudla, J., & Harter, K. (2007). The At Gen Express Global Stress Expression Data Set: Protocols, Evaluation and Model Data Analysis of UV-B light, Drought and Cold Stress Responses. *Plant Journal*, 50, 347–363.
- Kim, D.H., Han, S.I., Go, B., Oh U.H., Kim, C.S., Jung, Y.H., Lee, J., & Kim, J.H. (2019). 2-Methoxy-4-vinylphenol Attenuates Migration of Human Pancreatic Cancer Cells via Blockade of FAK and AKT Signaling. *Anticancer Research*, 39(12), 6685–6691. doi: [10.21873/anticanres.13883](https://doi.org/10.21873/anticanres.13883)
- Kizilay, Z., & Kahraman Cetin N. (2018). Effect of Methyl Palmitate on the Formation of Epidural Fibrosis in an Experimental Epidural Fibrosis Model. *J Invest Surg*, 31(6), 469-474. doi: [10.1080/08941939.2017.1356403](https://doi.org/10.1080/08941939.2017.1356403).
- Ko, G.-A., Shrestha, S., & Kim Cho, S. (2018). *Sageretia thea* Fruit Extracts Rich in Methyl Linoleate and Methyl Linolenate Downregulate Melanogenesis via the Akt/GSK3 β Signaling Pathway. *Nutrition Research and Practice*, 12(1), 3. doi: [10.4162/nrp.2018.12.1.3](https://doi.org/10.4162/nrp.2018.12.1.3)
- Koeduka, T., Fridman, E., Gang, D.R., Vassao, D.G., Jackson, B.L., Kish, C.M., Orlova, I., Spassova, S.M., Lewis, N.G., & Noel, J.P. (2006). Eugenol and Isoeugenol, Characteristic Aromatic Constituents of Spices, are Biosynthesized Via Reduction of a Coniferyl Alcohol Ester. *Proceedings of the National Academy of Sciences, USA*, 103, 10128–10133.

- Ksouri, R., Megdiche, W., Debez, A., Falleh, A., Grignon, C., & Abdelly, C. (2007). Salinity Effects on Polyphenol Content and Antioxidant Activities in Leaves of the Halophyte *Cakile maritima*. *Plant Physiol. Biochem*, 45, 244–249.
- Kulusic, T., Radonic, A., Katalinic, V., & Milos, M. (2004). Use of Different Methods for Testing Antioxidative Activity of Oregano Essential Oil. *Food Chemistry*, 85, 633-640.
- Kumar, N., Biswas, S., Shrungeswara, A.H., Mallik, S.B., Viji, M.H., Mathew, J.E., Mathew, J., Nandakumar, K., & Lobo, R. (2017). Pinocembrin Enriched Fraction of *Elytranthe parasitica* (L.) Danser Induces Apoptosis in HCT 116 Colorectal Cancer Cells. *Journal of Infection and Chemotherapy*, 23(6), 354–359. doi: [10.1016/j.jiac.2017.02.009](https://doi.org/10.1016/j.jiac.2017.02.009)
- Kumar, S., Abedin, M., Singh, A.K., & Das, S. (2020). Role of Phenolic Compounds in Plant-Defensive Mechanisms. Dalam Rafiq Lone, Razia Shuab, & Azra N. Kamili (Penyunting), *Plant Phenolics in Sustainable Agriculture* (hlm. 517-528). Singapore: Springer.
- Kuo, C.-C., Chiang, W., Liu, G.-P., Chien, Y.-L., Chang, J.-Y., Lee, C.-K., Lo, M.-J., Huang, S.-H., Shih, M.C., & Kuo, Y.-H. (2002). 2,2'-Diphenyl-1-picrylhydrazyl Radical-Scavenging Active Components from Adlay (*Coix lachryma-jobi* L. Var.ma-yuen Stapf) Hulls. *Journal of Agricultural and Food Chemistry*, 50(21), 5850–5855. doi: [10.1021/jf020391w](https://doi.org/10.1021/jf020391w)
- Laekeman, G.M., Van Hoof, L., Haemers, A., Berghe, D.A.V., Herman, A.G., & Vlietinck, A.J. (1990). Eugenol a Valuable Compound Forin Vitro Experimental Research and Worthwhile for Furtherin Vivo Investigation. *Phytotherapy Research*, 4(3), 90–96. doi: [10.1002/ptr.2650040304](https://doi.org/10.1002/ptr.2650040304)
- Lalthanpuii, P.B., Zarzokimi, & Lalchhandama, K. (2020). Analysis of Chemical Constituents and Antiparasitic Activities of the Extracts of *Imperata cylindrical*. *Res. J. Pharm. Technol*, 13(2). doi: [10.5958/0974-360X.2020.00125.0](https://doi.org/10.5958/0974-360X.2020.00125.0)
- Langa-Lomba, N., Buzón-Durán, L., Sánchez-Hernández, E., Martín-Ramos, P., Casanova-Gascón, J., Martín-Gil, J., & González-García, V. (2021). Antifungal Activity against *Botryosphaeriaceae* Fungi of the Hydro-Methanolic Extract of *Silybum marianum* Capitula Conjugated with Stevioside. *Plants*, 10(7), 1363. doi: [10.3390/plants10071363](https://doi.org/10.3390/plants10071363)
- Lathifah, U. (2020). *Identifikasi Senyawa Metabolit Sekunder Dan Aktivitas Antioksidan Daun Tebu*. (Skripsi). Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam, Universitas Negeri Semarang, Semarang.
- Leiss, K.A., Choi, Y.H., Verpoorte, R., & Klinkhamer, P.G.L. (2011). An Overview of NMR-based Metabolomics to Identify Secondary Plant Compounds

- Involved in Host Plant Resistance. *Phytochemistry Reviews*, 10, 205-216. doi: [10.1007/s11101-010-9175-z](https://doi.org/10.1007/s11101-010-9175-z)
- Leung, H., Ko, C., Yue, G.G.L., Herr, I., Lau, C.B. (2018). The Natural Agent 4-vinylphenol Targets Metastasis and Stemness Features in Breast Cancer Stem-like Cells. *Cancer Chemotherapy and Pharmacology*, 82(2), 185-197. doi: <https://doi.org/10.1007/s00280-018-3601-0>
- Li, B., Qiao, L., Li, L., Zhang, Y., Li, K., Wang, L., & Qiao, Y. (2017). Novel Antihypertensive Peptides Derived from Adlay (*Coix lachryma-jobi* L. var. ma-yuen Stapf) Glutelin. *Molecules*, 22(4), 534. doi: [10.3390/molecules22040534](https://doi.org/10.3390/molecules22040534)
- Liang, J., Ning, A., Lu, P., An, Y., Wang, Z., Zhang, J., He, C., & Wang, Y. (2021). Biological Activities and Synergistic Effects of *Elsholtzia stauntonii* Essential Oil from Flowers and Leaves and their Major Constituents Against *Tribolium castaneum*. *European Food Research and Technology*. doi: <https://doi.org/10.1007/s00217-021-03829-4>
- Lin, J., Dou, J., Xu, J., & Aisa, H.A. (2012). Chemical Composition, Antimicrobial and Antitumor Activities of the Essential Oils and Crude Extracts of *Euphorbia macrorrhiza*. *Molecules*, 17(5), 5030–5039. doi: [10.3390/molecules17055030](https://doi.org/10.3390/molecules17055030)
- Lim, T.K. (2012). *Coix lachryma-jobi. Edible Medicinal And Non-Medicinal Plants*, 5, 243-261. doi: [10.1007/978-94-007-5653-3_14](https://doi.org/10.1007/978-94-007-5653-3_14)
- Lima, N.M., Preet, G., Marqui, S.R., Falcóski, T.O.R., Navegante, G., Soares, C.P., Andrade, T.J.A.S., Porta, F.A.L., Rakotondraie, H.L.R., Jaspars, M., & Silva, D.H.S. (2022). Metabolic Profiling of Inga Species with Antitumor Activity. *Molecules*, 27. doi: <https://doi.org/10.3390/molecules27154695>
- LIPI. (1986). *Jenis Rumput Dataran Rendah Lembaga Biologi nasional*. LIPI-Bogor.
- Liu, L., Duncan, N. A., Chen, X., & Cui, J. (2018). Exploitation of Job's Tears in Paleolithic and Neolithic China: Methodological Problems and Solutions. *Quaternary International*. doi: [10.1016/j.quaint.2018.11.019](https://doi.org/10.1016/j.quaint.2018.11.019)
- Luo, H. (2011). *Extraction of Antioxidant Compounds from Olive (Olea europaea) Leaf*. (Tesis). Master of Technology in Food Technology, Massey University, Albany, New Zealand.
- Ma, Z.Q. & Zhang, S.S. (2010). Light Intensity Affects Growth, Photosynthetic Capability, and Total Flavonoid Accumulation of *Anoectochilus* plants. *HortScience*, 45, 863–867.

- Madhavi, D.L., Dhespande, S.S., & Salunkhe, D.K. (1996). *Food Antioxidant Technological, Toxicological and Healt Perpectives*. New york: Marcel dekker inc.
- Manosroi, A., Sainakham, M., Chankhampan, C., Manosroi, W., & Manosroi, J. (2016). In Vitro Anti-cancer Activities of Job's tears (*Coix lachryma-jobi* Linn.) Extracts on Human Colon Adenocarcinoma. *Saudi Journal of Biological Sciences*, 23(2), 248–256. doi: [10.1016/j.sjbs.2015.03.008](https://doi.org/10.1016/j.sjbs.2015.03.008)
- Manosroi, J., Khositsuntiwong, N., & Manosroi, A. (2011). Biological Activities of Fructooligosaccharide (FOS)-containing *Coix lachryma-jobi* Linn. Extract. *Journal of Food Science and Technology*, 51(2), 341–346. doi: [10.1007/s13197-011-0498-6](https://doi.org/10.1007/s13197-011-0498-6)
- Manzano, P., Valmaña García, O., Malusín, J., Villamar, J., Quijano, M., Viteri, R., Barragan, A., & Orellana-Manzano, A. (2020). Larvicidal activity of ethanolic extract of Azadirachta indica against *Aedes aegypti* larvae. *Revista Facultad Nacional de Agronomía Medellín*, 73(3), 9315–9320. doi: [10.15446/rfnam.v73n3.80501](https://doi.org/10.15446/rfnam.v73n3.80501)
- Martins, J.L.R., Silva, D.M., Gomes, E.H., Fava1, S.A., Carvalho, M.F., Macedo, I.Y.L., Gil, E.S., Ghedini, P.C., Rocha, F.F., Silva, O.N., Fajemiroye, J.O., Pinto, E.M.H., & Costa, E.A. (2020). Evaluation of Gastroprotective Activity of Linoleic acid on Gastric Ulcer in a Mice Model. *Current Pharmaceutical Design*, 26, 1-6.
- Marxen, K., Vanselow, K.H., Lippemeier, S., & Hintze, R. (2007). Determination of DPPH Radical Oxidation Caused by Methanolic Extracts of Some Microalgal Species by Linear Regression Analysis of Spectrophotometric Measurements. *Sensors*, 7(10), 2080-2095.
- Matsuura, H.N. & Fett-neto, A.G. (2015). Plant Alkaloids: Main Features, Toxicity, and Mechanisms of Action. Dalam P. Gopalakrishnakone, C.R., Carlini, & R. Braun-Ligabue (Penyunting), *Plant Toxins* (hlm. 243-261). Springer Dordrecht. doi: [10.1007/978-94-007-6728-7](https://doi.org/10.1007/978-94-007-6728-7)
- Maungchanburi, S., Rattanaburee, T., Sukpondma, Y., Tedasen, A., Tipmanee, V., & Graidi, P. (2022). Anticancer Activity of *Piper cubeba* L. Extract on Triple Negative Breast Cancer MDA-MB-231. *Journal of Pharmacy & Pharmacognosy Research*, 10(1), 39-51. doi: https://doi.org/10.56499/jppres21.1160_10.1.39
- Mellway, R.D., Tran, L.T., Prouse, M.B., Campbell, M.M., & Constabel, C.P. (2009). The wound-, Pathogen-, and Ultraviolet B-responsive MYB134 Gene Encodes a R2R3 Transcription Factor that Regulates Proanthocyanidin Synthesis in Poplar. *Plant Physiology*, 150, 924–941.

- Modi, B., Koirala, N., Aryal, S.P., Shrestha, J., Koirala, S., Upadhyaya, J., Basnyat R.C., Nassan, M.A., Alqarni, M., & Batiha, G.E. (2020). *Tinospora cordifolia* (Willd.) Miers: Phytochemical Composition, Cytotoxicity, Proximate Analysis and their Biological Activities. *Cellular and Molecular Biology*. 67(1), 50-57. doi: <http://dx.doi.org/10.14715/cmb/2021.67.1.8>
- Mohammed, H. A., Al-Omar, M. S., Aly, M. S. A., & Hegazy, M. M. (2019). Essential Oil Constituents and Biological Activities of the Halophytic Plants, *Suaeda Vermiculata* Forssk and *Salsola Cyclophylla* Bakera Growing in Saudi Arabia. *Journal of Essential Oil Bearing Plants*, 1–12. doi: [10.1080/0972060x.2019.1574611](https://doi.org/10.1080/0972060x.2019.1574611)
- Mohiuddin, AK. (2019). Impact of Various Environmental Factors on Secondary Metabolism of Medicinal Plants. *J of Pharmacol & Clin Res*, 7(1), 555704. doi: [10.19080/JPCR.2019.07.555704](https://doi.org/10.19080/JPCR.2019.07.555704)
- Molyneux, P. (2004). The Use Of The Stable Free Radical Diphenylpicrylhydrazyl (DPPH) For Estimating Antioxidant Activity. *Songklanakarin Journal of Science and Technology*, 26 (2), 211-219.
- Mukhriani, 2014, Ekstraksi, Pemisahan Senyawa, dan Identifikasi Senyawa Aktif. *Jurnal Kesehatan*, 7(2). doi: <https://dx.doi.org/10.24252/kesehatan.v7i2.55>
- Nandika, D., Karlinasari, L., Arinana, A., Batubara, I., Sitanggang, P.S., Santoso, D., Witasari, L.D., Rachmayanti, Y., Firmansyah, D., Sudiana, I.K., & Hertanto, D.M. (2021). Chemical Components of Fungus Comb from Indo-Malayan Termite Macrotermes gilvus Hagen Mound and Its Bioactivity against Wood-Staining Fungi. *Forests*, 12, 1591. doi: <https://doi.org/10.3390/f12111591>
- Nielsen, C.K., Kjems, J., Mygind, T., Snabe, T., Schwarz, K., Serfert, Y., & Meyer, R.L. (2017). Antimicrobial Effect of Emulsion-Encapsulated Isoeugenol Against Biofilms of Food Pathogens and Spoilage Bacteria. *International Journal of Food Microbiology*, 242, 7–12. doi: [10.1016/j.ijfoodmicro.2016.11.002](https://doi.org/10.1016/j.ijfoodmicro.2016.11.002)
- Nishantini, A., Ruba, A.A., & Mohan, V.R. (2012). Total Phenolic, Flavonoid Contents and In Vitro Antioxidants Activity of Leaf Of *Suaeda monoica* Forssk ex Gmel (Chenopodiaceae). *International Journal of Advanced Life Science (IJALS)*, 1(5), 34-43.
- Novitasari, A.E. & Putri, D.Z. (2016). Isolasi dan Identifikasi Saponin pada Ekstrak Daun Mahkota Dewa dengan Ekstraksi Maserasi. *Jurnal Sains*. 6(12), 10-14.
- Nghiem, C.T., Jiang, G.L., Shen, K.F., & Wang, Z. (2016). Effect of Dose Fertilizer and Cultvars to the Active Compound Glyceryl Trioleate of *Coix lacryma-jobi L.* *Agrivita Journal of Agricultural Science*. 38(3), 261-268.

- Nguyen, H.T. & Cao, D.N. (2022). Antimicrobial Compounds of One *Streptomyces cellulosflavus* Strain Isolated from Can Gio Mangrove Soil, Vietnam. *GSC Biological and Pharmaceutical Sciences*, 19(3), 120–126.
- Nguyen, T.N., Son, S., Jordan, M.C., Levin, D.B., & Ayele, B.T. (2016). Lignin Biosynthesis in Wheat (*Triticum aestivum* L.): Its Response to Waterlogging and Association with Hormonal Levels. *BMC Plant Biology*, 16(28). doi: [10.1186/s12870-016-0717-4](https://doi.org/10.1186/s12870-016-0717-4).
- Nguta, J.M., Appiah, O.R., Nyarko, A.K., Yeboah, M.D., & Addo, P.G.A. (2015). Medicinal Plants Used to Treat TB in Ghana. *Intl J Mycobacteriol*, 4(2), 116-123. doi: <https://doi.org/10.1016/j.ijmyco.2015.02.003>
- Nurhalimah, S. (2020). *Etnobotani dan Aktivitas Antioksidan pada Biji dan Tangkai Buah Hanjeli (Coix lacryma-jobi L.)*. (Skripsi). Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam, Universitas Pendidikan Indonesia, Bandung.
- Nurmala, T. & Irwan, A.W. (2007). *Pangan Alternatif Berbasis Sorealia Minor*. Bandung: PT. Giratuna.
- Nurmala, T. (2011). Potensi dan Prospek Pengembangan Hanjeli (*Coix lacryma-jobi L*) sebagai Pangan Bergizi Kaya Lemak untuk Mendukung Diversifikasi Pangan Menuju Ketahanan Pangan Mandiri. *Jurnal Pangan*. 20(1), 41-48.
- Oentarini, T., Taty, R., & Zulhipri. (2011). “Uji Aktivitas Antioksidan dan Profil Fitokimia Rambutan Rapiyah (*Nephelium lappaceum*)” *Prosiding Simposium Penelitian Bahan Obat Alami* (hlm. 185-196). Solo: UPT Penerbitan dan Percetakan UNS.
- Ogidi, C.O., Ojo, A.E., Ajayi-Moses, O.B., Aladejana, O.M., Thonda, O.A., & Akinyele, B.J. (2021). Synergistic Antifungal Evaluation of Over-the-Counter Antifungal Creams with Turmeric Essential Oil or Aloe Vera Gel Against Pathogenic Fungi. *BMC Complementary Medicine and Therapies*, 21(1). doi: [10.1186/s12906-021-03205-5](https://doi.org/10.1186/s12906-021-03205-5)
- Okiki, P.A., Egbebi, A., Akharaiyi, F.C., Adewole, E., Asoso, SO.O. (2022). Drug Properties and Antimicrobial Evaluations of Extracts from *Phyllanthus amarus*. *Journal of Microbiology & Experimentation*, 10(1), 10–16. doi: [10.15406/jmen.2022.10.00346](https://doi.org/10.15406/jmen.2022.10.00346)
- Olakunle, O.M. & Bola, A. (2017). GC-MS Anylsis of Phyto Components from the Stem Bark of *Cola nitida* Schott and Endl. *Journal of Plant Sciences*, 5(4), 99-103.doi: [10.11648/j.jps.20170504.11](https://doi.org/10.11648/j.jps.20170504.11)
- Oo, T., Saiboonjan, B., Srijampa, S., Srisrattakarn, A., Sutthanut, K., Tavichakorntrakool, R., Chanawong, A., Lulitanond, A., & Tippayawat, P.

- (2021). Inhibition of Bacterial Efflux Pumps by Crude Extracts and Essential Oil from *Myristica fragrans* Houtt. (Nutmeg) Seeds against *Methicillin-Resistant Staphylococcus aureus*. *Molecules*, 26(15), 4662. doi: [10.3390/molecules26154662](https://doi.org/10.3390/molecules26154662)
- Omoroyi, B.E., Afolayan, A.J., & Bradley, G. (2014). The Inhibitory Effect of *Mesembryanthemum edule* (L.) Bolus Essential Oil on Some Pathogenic Fungal Isolates. *BMC Complementary and Alternative Medicine*, 14, 168.
- Otsuka, H., Hirai, Y., & Nagao, T. (1988). Anti-inflammatory Activity of Benzoazinoids from Roots of *Coix lachryma-jobi* var. *ma-yuen*. *Journal Nat. Prod.*, 51(1), 74-79.
- Palanisamy, C.P., Alugoju, P., Jayaraman S., & Poompradub, S. (2023). *Nigella sativa* L. Seed Extracts Promote Wound Healing Progress by Activating VEGF and PDGF Signaling Pathways: An In Vitro and In Silico Study. *F1000 Research*, 12, 436. doi: <https://doi.org/10.12688/f1000research.132106.1>
- Pangemanan, D.A.M Suryanto, E., & Yamlean, P.V.Y. (2020). Skrining Ditokimia, Uji Aktivitas Antioksidan dan Tabir Surya pada Tanaman Jagung (*Zea mays* L.). *Parmachon*, 9(2).
- Pangemanan, K., Bodhi, W., Datu, O.S., Fatimawali, Kalalo, M.J., & Windah, A.L. (2022). Uji Aktivitas Antidiabetes Daging Buah Alpukat Mentega (*Parsea Americana*) Sebagai Inhibitor Enzim Alfa Glukosidase Secara In Silico. *Pharmacy Medical Journal*, 5(2).
- Pangerang, F. (2021). Kandungan Gizi dan Aktivitas Antioksidan Beras Merah dan Beras Hitam Padi Ladang Lokal dari Kabupaten Bulungan, Provinsi Kalimantan Utara. *Journal of Tropical Agrifood*, 3(2), 93-100.
- Pekal, A. & Pyrzynska. (2012). Estimation of antioxidant properties of teas using DPPH Assay. *Analytical Chemistry*, 9, 39-48.
- Perangin-Angin, Y., Purwaningrum, Y., Asbur, Y., Rahayu, M.S., & Nurhayati. (2019). Pemanfaatan Kandungan Metabolit Sekunder yang Dihasilkan Tanaman pada Cekaman Biotik. *Agriland*, 7(1), 39-47. doi: <https://doi.org/10.30743/agr.v7i1.3471>
- Pereira, D.M., Valentao, P., Pereira, J., & Andrade, B.P. (2009). Phenolics: From Chemistry to Biology. *Molecules*, 14(6), 2202–2211. doi: [10.3390/molecules14062202](https://doi.org/10.3390/molecules14062202)
- Pinelo, M., Rubilar, M., Jerez, M., Sineiro, J., & Núñez, M.J. (2005). Effect of Solvent, Temperature, and Solvent-to-Solid Ratio on the Total Phenolic Content and Antiradical Activity of Extracts from Different Components of

- Grape Pomace. *Journal of Agricultural and Food Chemistry*, 53(6), 2111–2117. doi: [10.1021/jf0488110](https://doi.org/10.1021/jf0488110)
- Pinto, M.E.A., Araujo, S.G., Morais, M.I., SÁ,N.P., Lima, C.M., Rosa, C.A. Siquera, E.P., Johann, S., & Lima, L.A.R.S. (2017). Antifungal and Antioxidant Activity of Fatty Acid Methyl Esters from Vegetable Oils. *Anais Da Academia Brasileira de Ciências*, 89(3), 1671–1681. doi:[10.1590/0001-3765201720160908](https://doi.org/10.1590/0001-3765201720160908)
- Pott, D.M., Osorio, s., & Vallarino, J.G. (2019). From Central to Specialized Metabolism: An Overview of Some Secondary Compounds Derived from the Primary Metabolism for Their Role in Conferring Nutritional and Organoleptic Characteristics to Fruit. *Frontiers in Plant Science*, 10. doi: [10.3389/fpls.2019.00835](https://doi.org/10.3389/fpls.2019.00835)
- Prakash, A. (2001). Antioxidant Activity. *Medallion Laboratories Analytical Progress*, 19(2).
- Praptiwi, Dewi, P., & Harapini, M. (2006). Nilai Peroksida dan Aktivitas Anti Radikal Bebas Diphenyl Picril Hydrazil Hidrate (DPPH) Ekstrak Metanol *Knema laurina*. *Majalah Farmasi Indonesia*, 17(1), 32-36.
- Prasad, S. N. & Muralidhara. (2012). Neuroprotective Efficacy of Eugenol and Isoeugenol in Acrylamide-Induced Neuropathy in Rats: Behavioral and Biochemical evidence. *Neurochemical Research*, 38(2), 330–345. doi: [10.1007/s11064-012-0924-9](https://doi.org/10.1007/s11064-012-0924-9)
- Prasathkumar, M., Raja, K., Vasanth, K., Khusro, A., Sadhasivam, S., Sahibzada, M.U.K., Gawwad, M.R.A., Al Farraj, D.A., & Elshikh, M.S. (2021). Phytochemical Screening and In Vitro Antibacterial, Antioxidant, Anti-inflammatory, Anti-diabetic, and Wound healing Attributes of *Senna auriculata* (L.) Roxb. Leaves. *Arabian Journal of Chemistry*, 14(9), 103345. doi: [10.1016/j.arabjc.2021.103345](https://doi.org/10.1016/j.arabjc.2021.103345)
- Pratimasari, D. (2009). *Uji Aktivitas Penangkap Radikal Buah Carica papaya L. Dengan Metode DPPH dan Penetapan Kadar Fenolik Serta Flavonoid Totalnya*. (Skripsi). Fakultas Farmasi, Universitas Muhammadiyah, Surakarta.
- Pratt, D.E. & Hudson, B.J.F. (1990). Natural Antioxidants Not Exploited Commercially. Dalam B.J.F. Hudson (Penyunting), *Food Antioxidants* (hlm. 171-192). Amsterdam: Elsevier Applied Science. doi: <https://doi.org/10.1007/978-94-009-0753-95>
- Prayitno, S.A., Kusnadi, J., & Murtini, E.S. (2016). Antioxidant Activity of Red Betel Leaves Extract (*Piper crocatum Ruiz and Pav.*) By Different Concentration of Solvents. *Journal of Pharmaceutical, Biological and Chemical Science*, 7(5), 1836-1843.

- Prayitno, S.A. & Rahim, A.R. (2020). The Comparison of Extracts (Ethanol And Aquos Solvents) *Muntingia calabura* Leaves on Total Phenol, Flavonid And Antioxidant (IC_{50}) Properties. *Kontribusia (Research Dissemination for Community Development)*, 3(2), 319. doi: [10.30587/kontribusia.v3i2.1451](https://doi.org/10.30587/kontribusia.v3i2.1451).
- Prayoga G. (2013). *Fraksinasi, Uji Aktivitas Antioksidan dengan Metode DPPH dan Identifikasi Golongan Senyawa Kimia dari Ekstrak Teraktif Daun Sambang Darah (Excoecaria cochinchinensis Lour)*. (Skripsi). Fakultas Farmasi, Program Studi Sarjana Ekstensi, Universitas Indonesia, Jakarta.
- Priyanto, A. & Islamiyati, R. (2018). Uji Aktivitas Antioksidan pada Batang Tebu Hijau dan Batang Tebu Merah Menggunakan Metode Peredaman Radikan Bebas DPPH. *Cendekia Journal of Pharmacy*, 2(1).
- Prommaban, A., Utama-ang, N., Chaikitwattana, A., & Uthaipibull, C. (2019). Linoleic Acid-rich Guava Seed Oil: Safety and Bioactivity. *Phytotherapy Research*, 1–16.
- Promraksa, B., Katrun, P., Phetcharaburanin, J., Kittirat, Y., Namwat, N., Techasen, A., Li, J.V., & Loilome, W. (2021). Metabolic Changes of Cholangiocarcinoma Cells in Response to Coniferyl Alcohol Treatment. *Biomolecules*, 11, 476. doi: <https://doi.org/10.3390/biom11030476>
- Puspitasari, A. D. & Proyogo, L., S. (2017). Perbandingan Metode Ekstraksi Maserasi dan Sokletasi terhadap Kadar Flavonoid Total Ekstrak Etanol Daun Kersen (*Muntingia calabura*). *Jurnal Ilmiah Cendekia Eksata*, 2(1). doi: [10.3194/ce.v2i1.1791](https://doi.org/10.3194/ce.v2i1.1791)
- Putra, A.A.B., Bogoriani, N.W., Diantariani, N.P., & Sumadewi, N.L. (2014). Ekstraksi Zat Warna Alam dari Bonggol Tanaman Pisang (*Musa Paradisiaca* L.) dengan Metode Maserasi, Refluks, dan Sokletasi. *Jurnal Kimia*, 8(1), 113-119. doi: <https://doi.org/10.24843/JCHEM.2014.v08.i01.p18>
- Radušienė, J., Karpavičienė, B., Stanius, Ž. (2012) Effect of External and Internal Factors on Secondary Metabolites Accumulation in St. John's worth. *Bot Lith.* doi: <https://doi.org/10.2478/v10279-012-0012-8>
- Raj, N., Sekaran, Aneesha, & Mathew, C.K. (2020). Evaluation of Phytochemicals in Leaf Part of *Atalantia monophylla* (Wild Lemon) and Bioinformatic approach for Evaluating its Medicinal Properties. *Advances in Bioresearch*, 11(6), 197-205.
- Ratnasari, E. (2019). *Kandungan Metabolit Sekunder pada Akar dan Daun Hanjeli (Coix lacryma-jobi L.) Liar dan Budidaya dengan GC-MS*. (Skripsi). Fakultas Pendidikan Matematika dan Ilmu Pengetahuan Alam, Universitas Pendidikan Indonesia, Bandung.

- Renganathan, S., Sahu, S.K., & Kathiresan, K. (2015). Phytochemical and Molecular Docking Analyses of *Prosopis juliflora* Derived Phytochemicals Against Anti-apoptotic Protein (BCL2). *World Journal of Pharmaceutical Research*, 4(5).
- Riwanti, P., Izazih, F., & Amaliyah. (2020). Pengaruh Perbedaan Konsentrasi Etanol pada Kadar Flavonoid Total Ekstrak Etanol 50,70, dan 96% *Sargassum Polycystum* dari Madura. *Journal of Pharmaceutical Care Anwar Medika*, 2(2). doi: <https://dx.doi.org/10.36932/jpcam.v2i2.1>
- Roberts, M.R. & Paul, N.D. (2006). Seduced by the Dark Side: Integrating Molecular and Ecological Perspectives on the Influence of Light on Plant Defence Against Pests and Pathogen. *New Phytologist*, 170, 677–699.
- Roopa, M.S., Shubharani, R., Rhetsø, T., & Sivaram, V. (2020). Comparative Analysis of Phytochemical Constituents, Free Radical Scavenging Activity and GC-MS Analysis of Leaf and Flower Extract of *Tithonia diversifolia* (Hemsl.) A. Gray. *International Journal of Pharmaceutical Sciences and Research*, 11(10), 5081-5090.
- Rubab, M., Chelliah, R., Saravanakumar, K., Barathikannan, K., Wei, S., Kim, J.-R., Yoo, D., Wang, M.H., & Oh, D.H. (2020). Bioactive Potential of 2-Methoxy-4-vinylphenol and Benzofuran from *Brassica oleracea L. var. capitata f. rubra* (Red Cabbage) on Oxidative and Microbiological Stability of Beef Meat. *Foods*, 9(5), 568. doi: [10.3390/foods9050568](https://doi.org/10.3390/foods9050568)
- Ruminta, Yuwariah, Y., & Sabrina, N. (2017). Respon Pertumbuhan dan Hasil Tanaman Hanjeli (*Coix lacryma-jobi L.*) terhadap Jarak Tanam dan Pupuk Pelengkap Cair. *Jurnal Agrikultura*, 28(2), 82-89.
- Saeed, N.M., El-Demerdash, E., Abdel-Rahman, H.M., Algandaby, M.M., Al-Abbasi, F.A., & Abdel Naim, A.B. (2012). Anti-inflammatory Activity of Methyl Palmitate and Ethyl Palmitate in Different Experimental Rat Models. *Toxicology Appl. Pharmacology*, 264(1), 84-93. doi: [10.1016/j.taap.2012.07.020](https://doi.org/10.1016/j.taap.2012.07.020)
- Saffaryazdi, A., Ganjeali, A., Farhoosh, R., & Cheniany, M. (2020). Variation in Phenolic Compounds, α -linolenic Acid and Linoleic acid Contents and Antioxidant Activity of Purslane (*Portulaca oleracea L.*) During Phonological Growth Stages. *Physiology and Molecular Biology of Plants*. doi: <https://doi.org/10.1007/s12298-020-00836-9>
- Saifudin, A. (2014). *Senyawa Alam Metabolit Sekunder: Teori, Konsep, dan Teknik Pemurnian*. Sleman: Deepublish.
- Saibabu, V., Fatima, Z., Khan, L.A., & Hameed, S. (2015). Therapeutic Potential of Dietary Phenolic Acids. *Advances in Pharmacological Sciences*. doi: <http://dx.doi.org/10.1155/2015/823539>

- Sakagami, H., Hashimoto, K., Suzuki, F., Ogiwara, T., Satoh, K., Ito, H., Hatano, T., Takashi, Y., & Fujisawa, S. (2005). Molecular Requirements of lignin–Carbohydrate Complexes for Expression of Unique Biological Activities. *Phytochemistry*, 66(17), 2108–2120. doi: [10.1016/j.phytochem.2005.05.0](https://doi.org/10.1016/j.phytochem.2005.05.0)
- Saltveit, M.E. (2017). Synthesis and Metabolism of Phenolic Compounds. Dalam Elhadi M. Yahia (Penyunting), *Fruit and Vegetable Phytochemicals: Chemistry and Human Health*, 2nd Edition (hlm. 115–124). Amerika Serikat: John Wiley & Sons Ltd. doi: [10.1002/9781119158042.ch5](https://doi.org/10.1002/9781119158042.ch5)
- Sanchez, S. & Demain, A.L. (2011). Secondary Metabolites. Dalam Murray Moo-Young (Penyunting), *Comprehensive Biotechnology* (hlm. 131–143). Amerika Serikat: Elsevier. doi: [10.1016/B978-0-444-64046-8.00012-4](https://doi.org/10.1016/B978-0-444-64046-8.00012-4)
- Sanchita & Sharma, A. (2018). Gene Expression Analysis in Medicinal Plants under Abiotic Stress Conditions. *Plant Metabolites and Regulation under Environmental Stress*, 407–414.
- Sari, L.O.R.K. (2006). Pemanfaatan Obat Tradisional dengan Pertimbangan Manfaat dan Keamanannya. *Majalah Ilmu Kefarmasian*, 3(1), 1-7.
- Sarveswaril, H.B., Gupta, K.K., Durai, R., & Solomon, A.P. (2023). Development of a Smart pH-responsive Nano-polymer Drug, 2-methoxy-4-vinylphenol Conjugate Against the Intestinal Pathogen, *Vibrio cholera*. *Scientific Reports*, 13,1250. doi: <https://doi.org/10.1038/s41598-023-28033-0>
- Sembiring, B.B. (2007). *Satus Teknologi Pasca Panen Sambiloto (Andrographis paniculata Needs)*. Jakarta: Balai Penelitian Tanaman Obat dan Aromatik, 134-144.
- Senja, R.Y., Issusilaningtyas, E., Nugroho, A.K., & Setyowati, E.P. (2014). The Comparison of Extraction Method and Solvent Variation on Yield and Antioxidant Activity of *Brassica oleracea L. var. capitata f. rubra* Extract. *Traditional Medicine Journal*, 19(1), 43-48. doi: <https://doi.org/10.24843/itepa.2019.v0 8.i01.p04>
- Setiani, L.A., Sari, B.L., Indriani, L., & Jupersio. (2017). Penentuan Kadar Flavanoid Ekstrak Etanol 70% Kulit Bawang Merah (*Allium cepa L.*) dengan Metode Maserasi dan MAE (*Microwave Assister Extract*). *Fitokimia*. 7(2), 15-22.
- Setiawan, I. R., Isa, I. G. T., Hestiana, S., & Tsani, A. (2020). Kampung Eduwisata Hanjeli di Desa Waluran Mandiri Kabupaten Sukabumi. *Abdimas:Jurnal Pengabdian Masyarakat Universitas Merdeka Malang*, 5(3), 300-311. doi: <https://doi.org/10.26905/abdimas.v5i3.4382>

- Shaaban, M.T., Ghaly, M.F., & Fahmi, S.M. (2021). Antibacterial Activities of Hexadecanoic Acid Methyl Ester and Green-Synthesized Silver Nanoparticles Against Multidrug-Resistant Bacteria. *Journal of Basic Microbiology*, 61(6), 557–568. doi: [10.1002/jobm.202100061](https://doi.org/10.1002/jobm.202100061)
- Shadmani, A., Azhar, I., Mzhar, F., Hassan, M.M., Ahmed, S.W., Iqbal, A., Usmanghani, K., & Shamim, S. (2004). Kinetic Studies on *Zingiber officinale*. *Journal of Pharmaceutical Sciences*, 17(1), 47-54.
- Sharifi-Rad, J., Quispe, C., Bouyahya, A., Meniyi, N.E., Omari, N.E., Md Shahinuzzaman, Md., Ovey, M.H.A., Koirala, N., Panthi, M., Ertani, A., Nicola, S. Lapava, N., Herrera-Bravo, J., Salazar, L.A., Changan, S., Kumar, M. & Calina, D. (2022). Ethnobotany, Phytochemistry, Biological Activities, and Health-Promoting Effects of the Genus *Bulbophyllum*. *Evidence-Based Complementary and Alternative Medicine*. doi: <https://doi.org/10.1155/2022/6727609>
- Shobier, A.H., Ghani, S.A.A., & Barakat, K.M. (2016). GC/MS Spectroscopic Approach and Antifungal Potential of Bioactive Extracts Produced by Marine Macroalgae. *Egyptian Journal of Aquatic Research*. doi: <http://dx.doi.org/10.1016/j.ejar.2016.07.003>
- Soledad, C.-P.T., Paola, H.-C., Carlos Enrique, O.-V., Israel, R.-L.I., GuadalupeVirginia, N.-M., & Raúl, Á.-S. (2021). Avocado Seeds (*Persea americana* cv. Criollo sp.): Lipophilic Compounds Profile and Biological Activities. *Saudi Journal of Biological Sciences*, 28(6), 3384–3390. doi: [10.1016/j.sjbs.2021.02.087](https://doi.org/10.1016/j.sjbs.2021.02.087)
- Souliman, S.I., Kiran, B., Gur, M., Guney, K., Altuner, E.M., Cetter, T., Hajar, & Mohammed. (2019). Chemical Composition and Antimicrobial Activities of Cold-Pressed Oils Obtained From *Nigella sativa* and *Prunus amygdalus*. *Tobruk University Journal of Medicine Science*, 2(1).
- Sotuboa, S.E., Lawal, O.A., Osunsamia, A.A., & Ogunwande, I.A. (2016). Constituents and Insecticidal Activity of *Deinbollia pinnata* Essential Oil. *Natural Product Communications*, 11(12).
- Suhendra, C., Widarta, I., & Wiadnyani, A. (2019). Pengaruh Konsentrasi Etanol terhadap Aktivitas Antioksidan Ekstrak Rimpang Ilalang (*Imperata cylindrica* (L) Beauv.) Pada Ekstraksi Menggunakan Gelombang Ultrasonik. *Jurnal Ilmu Dan Teknologi Pangan (ITEPA)*, 8(1), 27-35. doi: <https://doi.org/10.24843/itepa.2019.v0>
- Sukmawaty, E., Masri, M., Sijid, S.S., Afni, N., & Nasaruddin, N. (2019). Aktivitas Antioksidan Ekstrak Etanol Bekatul Sorgum (*Sorghum bicolor* L.) Super 2 Varietas dalam Menurunkan Kadar MDA Mencit (*Mus musculus*). *Prosiding Seminar Nasional Sains dan Teknologi Terapan*, 2.

- Sumardika I.W. & Jawi I.M. (2011). Ekstrak Air Daun Ubijalar Ungu Memperbaiki Profil Lipid dan Meningkatkan Kadar SOD Darah Tikus Yang Diberi Makanan Tinggi Kolesterol. *Jurnal Ilmiah Kedokteran*, 43(2), 67-70.
- Sunarni, T. (2005). Aktivitas Antioksidan Penangkap Radikal Bebas Beberapa Kecambah dari Biji Tanaman Familia Papilionaceae. *Jurnal Farmasi Indonesia*, 2(2), 53-61.
- Suresh, Y. & Das U.N. (2003). Long-chain Polyunsaturated Fatty Acids and Chemically Induced Diabetes mellitus: Effect of Omega-6 Fatty Acids. *Nutrition*, 19(2), 93–114.
- Suryani, N., Permana, D., & Jambe, A. (2016). Pengaruh Jenis Pelarut terhadap Kandungan Total Flavonoid dan Aktivitas Antioksidan Ekstrak Daun Matoa (*Pometia pinnata*). *Jurnal Ilmu Dan Teknologi Pangan (ITEPA)*, 5(1).
- Syafrida, M., Darmanti, S., & Izzati, M. (2018). Pengaruh Suhu Pengeringan Terhadap Kadar Air, Kadar Flavonoid, dan Aktivitas Antioksidan Daun dan Umbi Rumput Teki (*Cyperus rotundus L.*). *Bioma*, 20(1), 44-50.
- Syed, R.U., Moni, S.S., Huwaimel, B., Alobaida, A., Almarshdi, A.A., Abouzied, A.S., Lila, A.S.A., Abdallah, M.H., Banu, H., Hadi, M.A., El-Horany H.E., Abdelwahab, S.I., & Taha, M.M.E. (2022). Bioactive Principles, Anti-diabetic, and Anti-ulcer Activities of *Ducrosia Anethifolia* Boiss leaves from the Hail region, Saudi Arabia. *Arabian Journal of Chemistry*, 15, 104308. doi: <https://doi.org/10.1016/j.arabjc.2022.104308>
- Taiz , L. & Zeiger, E. (2015). *Plant Physiology*. USA: Sinauer Associates, Inc. Massachusetts, 545-582.
- Teixeira, R. & Silva, L.R. (2013). Bioactive Compounds and In Vitro Biological Activity of *Euphrasia rostkoviana* Hayne Extracts. *Industrial Crops and Products*, 50, 680–689. doi: [10.1016/j.indcrop.2013.08.035](https://doi.org/10.1016/j.indcrop.2013.08.035)
- Tetti, M. (2014). Ekstraksi, Pemisahan Senyawa, dan Identifikasi Senyawa Aktif. *Jurnal Kesehatan*, 7(2), 361-367.
- Twaji, B.M. & Hasan, Md.N. (2022). Bioactive Secondary Metabolites from Plant Sources: Types, Synthesis, and Their Therapeutic Uses. *Int. J. Plant Bio*, 13, 4–14. doi: <https://doi.org/10.3390/ijpb13010003>
- Tzin, V., Galili, G., & Aharoni, A. (2012). Shikimate Pathway and Aromatic Amino Acid Biosynthesis. *Encyclopedia of Life Sciences*, 1–10. doi: [10.1002/9780470015902.a0001315.pub2](https://doi.org/10.1002/9780470015902.a0001315.pub2)
- Vagueresse, M.H.D.L., Romiti, C., Grosclaude, C., & Bounias, M. (2000). Coevolutionary Toxicity as Suggested by Differential Coniferyl Alcohol Inhibition of Ceratocystis Species Growth. *Toxicon*, 39, 203-208.

- Vanholme, R., Demedts, B., Morreel, K., Ralph, J., & Boerjan, W. (2010). Lignin Biosynthesis and Structure. *Plant Physiology*, 153, 895–905.
- Verma, B., Karakoti, H., Kumar, R., Mahawer, S.K., Prakash, O., Srivastava, R.M., Kumar, S. Shilpi Rawat, S., Rawat, D.S., & de Oliveira, M.S. (2023). Phytochemical Screening and Evaluation of Pesticidal Efficacy in the Oleoresins of *Globba sessiliflora* Sims and In Silico Study. *Evidence-Based Complementary and Alternative Medicine*. doi: <https://doi.org/10.1155/2023/5936513>
- Verma, N. & Shukla, S. (2015). Impact of Various Factors Responsible for Fluctuation in Plant Secondary Metabolites. *Journal of Applied Research on Medicinal and Aromatic Plants*, 2(4), 105–113. doi: [10.1016/j.jarmap.2015.09.002](https://doi.org/10.1016/j.jarmap.2015.09.002)
- Virsangbhai, C.K., Goyal, A., Tanwar, B., & Sihag, M.K. (2019). Potential Health Benefits of Conjugated Linoleic Acid: An Important Functional Dairy Ingredient. *European Journal of Nutrition & Food Safety*, 11(4), 200-213. doi: [10.9734/EJNFS/2019/v11i430162](https://doi.org/10.9734/EJNFS/2019/v11i430162)
- Wahyuni, D.T. & Widjanarko, S.B. (2015). Pengaruh Jenis pelarut dan Lama Ekstraksi terhadap Ekstrak Karotenoid Labu Kuning dengan Metode Gelombang Ultrasonik. *Jurnal Pangan dan Agroindustri*, 3(2), 390-401.
- Wang, Y.N., Wang, H.X., Shen, Z.J., Zhao, L.L., Clarke, S.R., Sun, J.H., Du, Y.Y., & Shi, G.L. (2009). Methyl Palmitate, an Acaricidal Compound Occurring in Green Walnut Husks. *J. Econ. Entomol.*, 102, 196–202.
- Wanti, S., Andriani, M.A.M., Parnanto, N.H.R. (2015). Pengaruh Berbagai Jenis Beras terhadap Aktivitas Antioksidan pada Angkak oleh *Monascus purpureus*. *Biofarmasi*, 13(1), 1-5.
- Wariyapperuma, W.A.N.M., Kannangara, S., Wijayasinghe, Y.S., Subramanium, S., & Jayawardene, B. (2020). In Vitro Anti-diabetic Effects and Phytochemical Profiling of Novel Varieties of *Cinnamomum zeylanicum* (L.) Extracts. *PeerJ*. doi: [10.7717/peerj.10070](https://doi.org/10.7717/peerj.10070)
- Warnis, M., Aprilina, L.A., & Maryanti, L. (2020). Pengaruh Suhu Pengeringan Simplicia terhadap Kadar Flavonoid Total Ekstrak Daun Kelor (*Moringa oleifera* L.). *Prosidiang Seminar Nasional Kahuripan 1 Universitas Kahuripan* (264-268). Kediri: Seminar Nasional Kahuripan.
- Wei, G., Kong, L., Zhang, J., Ma, C., Wu, X., Li, X. and Jiang, H. (2016). Essential Oil Composition and Antibacterial Activity of *Lindera nacusua* (D. Don) Merr. *National Product Research*, 30, 2704-2706. doi: [10.1080/14786419.2015.1135145](https://doi.org/10.1080/14786419.2015.1135145)

- Wijaya, H., Novitasari, & Jubaidah, S. (2018). Perbandingan Metode Ekstraksi terhadap Rendemen Ekstrak Daun Rambai Laut (*Sonneratia caseolaris* L. Engl). *Jurnal Ilmiah Manuntung*, 4(1), 79, 79-83.
- Winangsih., Prihastanti, E. & Parman, S. (2013). Pengaruh Metode Pengeringan Terhadap Kualitas Simplicia Lempuyang Wangi (*Zingiber aromaticum* L.). *Buletin Anatomi dan Fisiologi*, 21(1), 19-25.
- Winarsi, H. (2007). *Antioksidan Alami dan Radikal Bebas: Potensi dan Aplikasinya dalam Kesehatan*. Yogyakarta : Kanisius Media.
- Wu, H., Xue, R., Dong, L., Liu, T., Deng, C., Zeng, H., & Shen, X. (2009). Metabolomic Profiling of Human Urine in Hepatocellular Carcinoma Patients using Gas Chromatography/Mass Spectrometry. *Analytica Chimica Acta*, 648, 98-104.
- Xi, X-J., Zhu, Y-G., Tong, Y-P., Yang, X-L., Tang, N-N, Ma, S-M., Li, S. & Cheng, Z. (2016). Assessment of the Genetic Diversity of Different Job's Tears (*Coix lacryma-jobi* L.) Accessions and the Active Composition and Anticancer Effect of Its Seed Oil. *PLoS ONE*, 11(4), 1-22. doi: [10.1371/journal.pone.0153269](https://doi.org/10.1371/journal.pone.0153269).
- Xu, B.J. & Chang, S.K.C. (2007). A Comparative Study on Phenolic and Antioxidant Activity of Legumes as Affected by Extraction Solvents. *Journal of Food Science*. 72(2), 159 – 166.
- Xuan, T.D., Roni, Y., Andriana, Y., Khanh, T.D., Anh, T.T.T., Kakar, K. & Haqani, M.S. (2018). Chemical Profile, Antioxidant Activities and Allelopathic Potential of Liquid Waste from Germinated Brown Rice. *Allelopathy Journal*, 45(1).
- Xue, R., Lin, Z., Deng, C., Dong, L., Liu, T., Wang, J., & Shen, X. (2008). A serum Metabolomic Investigation on Hepatocellular Carcinoma Patients by Chemical Derivatization Followed by Gas Chromatography/Mass Spectrometry. *Rapid Commun. Mass Spectrom.* 22, 3061–3068. doi: [10.1002/rcm.3708](https://doi.org/10.1002/rcm.3708)
- Yabalak, E., Ibrahim, F., Eliuz, E.A.E., Everest, A., & Gizir, A.M. (2020). Evaluation of Chemical Composition, Trace Element Content, Antioxidant and Antimicrobial Activities of *Verbascum pseudoholothrichum*. *Plant Biosystems - An International Journal Dealing with All Aspects of Plant Biology*, 1–10. doi:[10.1080/11263504.2020.185233](https://doi.org/10.1080/11263504.2020.185233)
- Yang, D., Du, X., Liang, X., Han, R., Liang, Z., Liu, Y., Liu, F., & Zhao, J. (2012). Different Roles of the Mevalonate and Methylerythritol Phosphate Pathways in Cell Growth and Tanshinone Production of *Salvia miltiorrhiza* Hairy Roots. *PLoS One*, 7. doi: [10.1371/journal.pone.0046797](https://doi.org/10.1371/journal.pone.0046797)

- Yang, Y., Du, S.Y., & Sun, Y.Q. (2017). Determination of Effective Contents Triolein and *Coix lacryma-jobi* var. ma-yuen from Different Origins. *Chinese Trad Herbal Drugs*, 48(3), 578-581.
- Yateem, H., Afaneh, I., & Al-Rimawi, F. (2014). Optimum Conditions for Oleurpein Extraction from Olive Leaves. *Int. J. Appl.*, 4(5), 153-157.
- Youngson, R. (2005). *Antioksidan: Manfaat Vitamin C dan E Bagi Kesehatan*. Jakarta : Arcan.
- Youssef, A.M., Maaty, D.A.M., & Al-Saraireh, Y.M. (2023). Phytochemistry and Anticancer Effects of Mangrove (*Rhizophora mucronata Lam.*) Leaves and Stems Extract against Different Cancer Cell Lines. *Pharmaceuticals*, 16, 4. doi: <https://doi.org/10.3390/ph16010004>
- Yu, F., Zhang, J., Li, Y.Z., Zhao, Z.Y., & Liu, C.X. (2017). Research and Application of Adlay in Medicinal Field. *Chinese Herbal Medicines*, 9(2), 126-133.
- Yu, F.-R., Lian, X.-Z., Guo, H.-Y., McGuire, P.M., Li, R.-D., Wang, R., & Yu, F.-H. (2005). Isolation and Characterization of Methyl Esters and Derivatives from *Euphorbia kansui* (Euphorbiaceae) and their Inhibitory Effects on the Human SGC-7901 Cells. *Journal of Pharmacy and Pharmaceutical Science*, 8(3), 528-535.
- Yue, G.G.L., Lee, J.K.M., Kwok, H.F., Cheng, L., Wong, E.C.W., Jiang, L., Yu, H., Leung, H.W., Wong, Y.L., Leung, P.C., Fung, K.P., & Lau, C.B.S. (2015). Novel PI3K/AKT Targeting Anti-angiogenic Activities of 4-vinylphenol, a New Therapeutic Potential of a Well-known Styrene Metabolite. *Scientific Reports*, 5(1). doi:10.1038/srep11149
- Zakaria, Z.A., Jaios, E.S., Omar, M.H., Abd. Rahman, S., Hamid, S.S.A., Ching, S.M., Teh, L.K., Salleh, M.Z., Deny, S., & Taher, M. (2016). Antinociception of Petroleum Ether Fraction Derived from Crude Methanol Extract of *Melastoma malabathricum* Leaves and its Possible Mechanisms of Action in Animal Models. *BMC Complementary and Alternative Medicine*, 16(1). doi: [10.1186/s12906-016-1478-1](https://doi.org/10.1186/s12906-016-1478-1)
- Zakaria, Z.A., Kamisan, F.H., Omar, M.H., Mahmood, N.D., Othman, F., Abdul Hamid, S.S., & Abdullah, M.N.H. (2017). Methanol Extract of *Dicranopteris linearis* L. Leaves Impedes Acetaminophen-induced Liver Intoxication Partly by Enhancing the Endogenous Antioxidant System. *BMC Complementary and Alternative Medicine*, 17(1). doi: [10.1186/s12906-017-1781-5](https://doi.org/10.1186/s12906-017-1781-5)
- Zhang, L., Shan, Y., Tang, K., & Putheti, R. (2009). Ultrasound Assisted Extraction Flavonoid of Lotus (*Nelumbo nuficera Gaertn*) Leaf and Evaluation of its Anti-fatigue Activity. *International Journal of Physical Science*, 4(8).

- Zhang, L.L., Zhang, L.F., Xu, J.G., & Hu, Q.P. (2017). Comparison Study on Antioxidant, DNA Damage Protective and Antibacterial Activities of Eugenol and Isoeugenol Against Several Foodborne Pathogens. *Food & Nutrition Research*, 61(1), 1353356. doi: [10.1080/16546628.2017.1353356](https://doi.org/10.1080/16546628.2017.1353356)
- Zhang, Z.Z., Li, X.X., Chu, Y.N., Zhang, M.X., Wen, Y.Q., Duan, C.Q., & Pan, Q.H. (2012). Three Types of Ultraviolet Irradiation Differentially Promote Expression of Shikimate Pathway Genes and Production of Anthocyanins in Grape Berries. *Plant Physiol. Biochem.*, 57, 74–83. doi: [10.1016/j.plaphy.2012.05.005](https://doi.org/10.1016/j.plaphy.2012.05.005)
- Zhou, W., He, Y., Lei, X., Liao, L., Fu, T., Yuan, Y., Huang, X., Zou, L., Liu, Y., Ruan, R., & Li, J. (2020). Chemical Composition and Evaluation of Antioxidant Activities, Antimicrobial, and Anti-melanogenesis Effect of the Essential Oils Extracted from *Dalbergia pinnata* (Lour.) Prain. *Journal of Ethnopharmacology*, 112731. doi: [10.1016/j.jep.2020.112731](https://doi.org/10.1016/j.jep.2020.112731)
- Zhu, F. (2017). Coix: Chemical Composition and Health Effects. *Trends Food Sci. Technol.*, 61, 160- 175.

